# **Organism and Tissue Growth**



## Course 4: Intrinsic control of growth and patterning

Thomas Lecuit chaire: Dynamiques du vivant



## • Summary: intrinsic and organ specific growth control

— In plants and animals tissue growth is driven by both cell division (increase in cell number) and cell growth (increase in cell size).

-The two processes are coordinated but can be independently regulated

- Cell growth is controlled intrinsically (cellular anabolism: Ras, Pi3K, TOR signalling)
- Cell division is controlled by intrinsic cell cycle regulators (Cdk/cyclins, E2F, Rb...)
- Two modalities of growth compensation:
- Cell Autonomous: The pattern of cell division does not affect tissue size: cell growth is dominant over and compensates for cell division defects (eg. policy, cdc2, endoreplication).
- Cell Non-Autonomous: Perturbations in cell growth (via Minute, Myc mutations) does not affect tissue size due to compensation dependent on cell competition.
- These compensatory mechanisms are specific to each organ
- Compensation reveals organ specific tissue size sensing/measurement





# -How is growth of each organ arrested when the it reaches its appropriate size?



## • Control of growth arrest at target size



- ---What is measured? Intrinsic « ruler » of growth: « size-meter » Organ intrinsic feedback of target organ size on cell growth/proliferation
- -Need to consider scaling between factors that promote growth and size itself
  - I. Energy and mass/volume
  - 2. Pattern and mass/volume
  - 3. Mechanics and mass/volume

-Need to consider feedback mechanisms operating across scales (organ/tissue to cell)



#### • Organ growth arrest and energy delivery constraints

— Hypothesis: An asymptotic mass emerges from imbalance between energy supply and energy demand associated with Kleiber law for each organ? (see course 2, 19 Nov)

$$B = B_0 M^{3/4}$$

- Constraints imposed by energy delivery through a fractal branching network with invariant termini
- Organ-level scaling law: the whole organism power law scaling of metabolic power to mass can be explained by organ-level scaling of metabolic power to organ mass.



## Organ growth arrest and energy delivery constraints

— Hypothesis: An asymptotic organ mass emerges from imbalance between energy supply-side and energy demand given the Kleiber law <u>for each organ</u>.

• Growth equation  
and Kleiber's law
$$\frac{dm}{dt} = \left(\frac{m_c}{E_c}\right) B - \left(\frac{B_c}{E_c}\right) m$$
give
$$\frac{dm}{dt} = am^{3/4} - bm$$
with
$$a \equiv B_0 m_c/E_c$$

$$b \equiv B_c/E_c.$$

$$B = B_0 m^{3/4}$$

$$B_0 \text{ constant within taxon (fish, bird, mammal etc)}$$

- Asymptotic mass/size for dm/dt = 0 M =
- Law of growth:  $\left(\frac{m}{M}\right)^{1/4} = 1 \left[1 \left(\frac{m_0}{M}\right)^1\right]$ ne t oirth • « Universal » growth  $r = 1 - e^{-\tau}$  $(m/M)^{1/4}$ Fraction of metabolic power allotted to maintenance The fraction allotted to growth exponentially decreases to 0 (growth arrests) OLLÈGE Dimensionless time, a  $\tau \equiv at/4M^{1/4} - \ln[1 - (m_0/M)^{1/4}]$ Thomas LECUIT 2019-2020 E FRANCE 6

## Organ growth arrest and energy delivery constraints

-Supply-side constraints versus metabolic demand in Manduca sexta

- The tracheal (ie. respiratory) system is over-built after molting and becomes limiting at the end of each instar
- The demand (which increases) cannot be met by supply
- This size-sensing (critical weight) depends on the limited ability of a fixed tracheal system to sustain the oxygen supply to a growing individual

V. Callier and F. Nijhout Proc Natl Acad Sci U S A. 2011 Aug 30;108(35):14664-9. doi: 10.1073/pnas.1106556108

- Yet, the mass-specific metabolic power at the beginning of each instar decreases from molt to molt and supply is no limiting at onset of each instar (respiratory system is « over-built »)
- So supply-side constraints cannot explain everything

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• Demand also decreases over time (mass-specific mitochondrial activity, cytochrome c oxydase COX) due to reduced proportion of highly metabolic tissues

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Chalones: secreted negative feedback inhibitors of growth



It is suggested that the members of this complex of chemical messengers, which may possibly prove to be a family of proteins, should be called *chalones*. The word chalone ( $\chi \alpha \lambda \dot{\alpha} \omega$ , to make slack) was originally proposed by Schäfer (1016) to distinguish a chemical messenger which has a depressant action and for which he felt the term hormone (Starling, 1906) was inappropriate. This suggestion was never adopted but the word may be most appropriate in the present case since the chalones seem to act most powerfully in slowing growth as the adult state is reached.

#### hormone vs chalone

There also arises a question of terminology. Substances of the kind under discussion do not fall within the usual definition of a hormone as a systemically distributed chemical substance 'produced by one tissue with the primary function of exerting a specific effect of functional value on another tissue' (Huxley, 1935). On the contrary, these are substances each of which is produced by a tissue with the primary function of controlling the growth, and perhaps the differentiation, of that same tissue,



CHART 5 .- Diagrams illustrating the different proportions of cell types in different tissues. P, progenitor cells; I, immature cells; M, mature cells; D, mature cells moving towards death. Top, duodenal mucosa with a high rate of cell production and loss; middle, epidermis with a moderate rate of cell production and loss; and bottom, liver with a low rate of cell production and

moderate cholone action

м

strong chalone action

м

s

G.P.

granulocyte

ρ

М

G.Ch.

Gron.D

JOB8

## • Chalones: secreted negative feedback inhibitors of growth

GDF 8/Myostatin:

- TGFB member secreted negative feedback inhibitor of muscle differentiation produced by myocytes. Autocrine inhibition
- Induces cell cycle inhibitor p21 (of cdk1) and promotes cell differentiation





### • Chalones: secreted negative feedback inhibitors of growth

• Different TGFB family secreted factors are produced by terminally differentiated cells and feedback on progenitor growth



McPherron AC, Lawler AM, Lee SJ. Nature. 1997;387:83-90.

Wu, H.H., et al and Calof, A.L. (2003). Neuron 37, 197–207.



## Organ specific negative feedback circuits of growth

Limbs: FGF/Shh/BMP network Transition between positive and negative feedback loop arrests growth of vertebrate limbs



- FGFs promote growth of mesenchymal cells in limb bud
- Positive feedback:
  - -FGF induces polarising signal Shh

—Shh induces, and FGF represses Gremlin an inhibitor of BMP which itself antagonises FGF.

—FGF is maintained through a Positive feedback that promotes growth during limb development

Shh



An FGF 2 bead induces a new limb



Cohn MJ, Izpisua-Belmonte JC, Abud H, Heath JK, Tickle C. (1995). Cell 80: 739 - 746.

## Organ specific negative feedback circuits of growth



#### • Negative Feedback:

—As the limb grows, FGF levels increase and Gremlin repression increases

—Limb growth leads to increased distances between Gremlin and FGF (Mouse) and Shh and Gremlin (Chick): break of positive feedback loop and induction of BMP dependent negative feedback

Scherz, P. J., Harfe, B. D., McMahon, A. P. & Tabin, C. J. (2004) Science 305, 396-399.



J. Verheyden and X. Sun Nature. (2008);454(7204):638-41

## Organ specific negative feedback circuits of growth

#### Bones



HM. Kronenberg (2003) Nature 423:332-336

- Chondrocytes proliferate, then differentiate into hypertrophic chondrocytes and eventually in osteoblasts
- PTHrP sustains proliferation of chondrocytes by blocking lhh expression.
- Following growth, Ihh is expressed at a distance in hypertrophyic chondrocytes (out of reach from PTHrP)
- IHh promotes hypertrophic chondrocyte differenciation and chondrocyte proliferation
- Differentiated cells induce feedback inhibitor of differentiation (via induction of PTHrP)



premature growth arrest and differentiation

Lanske, B. *et al.* Kronenberg HM. *Science* **273**, 663–666 (1996). Vortkamp, A. *et al.* Tabon C. . *Science* **273**, 613–622 (1996).



• The « size-meter» as an organ-scale negative feedback



#### -Measure of tissue size

Amount of inhibitory feedback signal: which scales with cell number/size
 Myocytes (GDF8/Myostatin) repress proliferation of myoblasts
 Olfactory receptor neurons (GDF11) repress proliferation of neuronal precursor
 FGF production during limb growth: inhibition of Gremlin

• Tissue spacing between signal sources Failure to sustain repression (via Gremlin) of negative feedback loop (BMP)





#### • The « size-meter » as an organ-scale negative feedback

#### -How robust is such a system of feedback inhibition based on tissue size?

The number of cells does not necessarily scale with the [concentration] of inhibitor

- Importance of: Tissue Geometry (size of pool/sink)
  - Inhibitor production rate, diffusion/transport and stability
  - What is the sensitivity to geometry (ie. size) given biochemical parameters?
  - What is the sensitivity to biochemical parameters given geometry?
  - Implies: fine tuning of growth rate to biochemical parameters of feedback at the cellular scale.
  - Or interdependency.

Cell growth/division can potentially advect (transport) and dilute inhibitor, or affect stability, etc



Hypothesis: Feedback of cell growth rate on inhibitor stability (positive feedback) could rescue size:: Robustness Necessity to invoque nested feedbacks: global and local.



Stability of inhibitor ++ Cell growth rate ++

## • Stability of Growth: cellular lineage

-Cell lineages balance growth (of stem cell pool and intermediates) and terminal differentiation



-What ensures the stability of a cell lineage?





Systems sensitivity to  $p_0$  is infinite: unless exactly  $\frac{1}{2}$  of cells remain stem cells and  $\frac{1}{2}$  differentiate ( $p_0=0.5$ ), stem cells pool collapses or explodes. But stem cells do not always divide asymmetrically. They may divide also symmetrically, *i*e. produce 2 or 0 stem cells.

Question: How does  $p_0=0.5$  as a <u>population average</u> if not absolutely true for all cells?

• Negative feedback



A. Lander et al . (2015) PLoS Biol 7(1): e1000015. https://doi.org/10.1371/journal.pbio.1000015

## • Stability of Growth: cellular lineage

#### OPEN a ACCESS Freely available online

#### PLOS BIOLOGY

#### Cell Lineages and the Logic of Proliferative Control

Arthur D. Lander<sup>1,2,3</sup>\*, Kimberly K. Gokoffski<sup>1,4,5</sup>, Frederic Y. M. Wan<sup>3,5</sup>, Qing Nie<sup>2,3,5</sup>, Anne L. Calof<sup>1,3,4\*</sup>

#### • Negative feedback

#### proportional control

(output proportional to error) does not compensate for steady deviation from reference high rate, low gain (amplification from input to output) requires high progenitor load to achieve high rate

#### integral control

(output depends on integral of error) compensates well for steady deviation from reference resistant to external and internal perturbations, high rate does not require high progenitor load









in silico regeneration « experiment »: simulation of return to steady-state after ablation of ORNs (  $_2=$  0)



## • Stability of Growth: cellular lineage

#### -GDFII as a proportional and integral feedback controller of lineage

- GDFII is produced by differentiated olfactory receptor neurons (ORNs)
- BrdU labelling of INPs and assess time course of NCAM immunoreactivity (markers of ORNs).
- INP replication delays differentiation of INPs. Thus, <u>delay in NCAM</u> <u>expression indicative of replication probability p1 of INPs.</u>
- 18h chase: 60% of BrdU+ cells become ORNs without GDF11.
- Addition of low GDF11 increases ORN production indicative of reduced probability of INP replication (*p*<sup>1</sup> lowered)
- Addition of higher GDF11 reduces ORNs indicative of lengthening of cell cycle progression (v1 lowered)
- If so, longer chase should reduce this inhibition, as observed (36h)

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 Stem cell
 INP
 ORN

 (Sox2\* and/or Mash1\*)
 (transit amplifying, Ngn1\*)
 (terminallydifferentiated, Ncam\*)









### • The « size-meter » as an organ-scale negative feedback

#### Mechanical feedback

Synthesis of an external mechanical constraint. For example: Stiff extracellular matrix, contractile tissue (eg. gut), non-growing tissue boundary.

The feedback requires that mechanical constraints are dependent on growth/size of the tissue/organ And that mechanical feedback blocks cell growth/division and/or induces apoptosis.



(Cours #5 Growth and Mechanics)

• Completion of tissue pattern:

Growth is maintained up to a point when pattern is « complete » Ensures scaling of growth and patterning.





### Size regulation and patterning

#### -regeneration and intercalary growth

#### **Pattern Regulation in Epimorphic Fields**

Cells may make use of a polar coordinate system for assessing their positions in developing organs.

Vernon French, Peter J. Bryant, Susan V. Bryant



Positional information

(cf Lewis Wolpert JTB 1969, See also Inaugural lecture)

polar coordinate model

#### When

normally nonadjacent positional values in either the circular or the radial sequence are confronted in a graft combination or as a result of wound healing, growth occurs at the junction until cells with all the intermediate positional values have been intercalated; then growth ceases.









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## • Size regulation and patterning

-regeneration and intercalary growth

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 Reviews
 V. French, P.J. Bryant and S.V. Bryant Science (1976). 193::969-981

 S.V. Bryant , V. French, and P.J. Bryant Science (1981). 212::993-1002

-regeneration and intercalary growth: possible mechanisms

- Positional information resulting from molecular morphogen gradients
- Growth is coupled to long range diffusion/transport of morphogen gradients



V. French, P.J. Bryant and S.V. Bryant Science (1976). 193::969-981

• Positional information resulting from local cell interactions and cell surface molecules.

S.V. Bryant, V. French, and P.J. Bryant Science (1981). 212::993-1002

**« Entelechia » model** A. C. Garcia-Bellido. and A. Garcia-Bellido. *Int. J. Dev. Biol.* 42: 353-362 (1998) A. Garcia-Bellido. *Int. J. Dev. Biol.* 53: 1291-1303 (2009) doi: 10.1387/ijdb.072459ag



## • Size regulation and patterning

—Morphogen gradients and leg patterning

- Proximal distal patterning occurs in 2D sheets of cells: gene expression in discs of different radii.
- Wg and Dpp are concentration dependent morphogens that pattern the proximo-distal and dorso-ventral axes
- The synergistic and concentration dependent activities of Wg and Dpp control the nested expression of genes in discs of different radii





morphogens





## Size regulation and patterning

#### -Morphogens: Growth factors and with patterning activity

- Gradient of concentration/activity of a molecule (Shh)
- Activity thresholds



Tetsuya Tabata, Yuki Takei Development 2004 131: 703-712



Rolf Zeller et al. Nature Reviews Genetics 10:45-858 (2009)



### • Size regulation and patterning

#### -Morphogens: Growth factors with patterning activity

- Gradient of concentration/activity of a molecule
- Activity thresholds





#### -Morphogens: Growth factors with patterning activity

• A source of expression of the morphogen Dpp induces axis duplication: ie. duplication of patterns AND extra tissue growth (intercalary growth)

#### Dpp inhibition



L. Barrio and M. Milan. eLife (2017);6:e22013. DOI: 10.7554/eLife.22013

#### Dpp ectopic expression



M. Zecca K. Basler and G. Struhl. Development 121, 2265-2278 (1995)



#### —Morphogens: Growth factors with patterning activity

- Dpp acts directly at a distance from its source
- Dose/concentration dependent effect of target gene activation
- Existence of activity thresholds



Ectopic clones of Dpp

D. Nellen et al. and K. Basler. *Cell* 1996; 85(3):357-68.T. Lecuit et al. and S. Cohen. *Nature* 1996;381(6581):387-93



T. Lecuit and S. Cohen Development. 1998 Dec;125(24):4901-7.

#### -Morphogens: Growth factors with patterning activity

• Dpp::GFP forms an exponential gradient from its source



Dpp::GFP morphogen gradient profile

$$\partial_{t}C = D\nabla^{2}C - kC + 2j_{0}\delta(x) \qquad (1) \quad \text{Mass conservation}$$
  
diffusion degradation production  

$$C(x) = C_{0}e^{-\frac{x}{\lambda}} \qquad (2) \quad \text{Steady state solution}$$
  

$$\lambda = \sqrt{D/k} \qquad (3) \quad \text{Decay length}$$
  

$$C_{0} = j_{0}/\sqrt{Dk} \qquad (4) \quad \text{Source concentration}$$
  

$$D = 0.10 \pm 0.05 \ \mu\text{m}^{2}\text{/s}$$
  

$$k = 2.52 \times 10^{-4} \pm 1.29 \times 10^{-4} \text{ s}^{-1}$$
  

$$j_{0} = 3.98 \pm 2.34 \quad \text{molecules/}(\mu\text{m} \times \text{s})$$



Anna Kicheva et al. M. Gonzalez-Gaitan *Science* 315, 521 (2007); doi: 10.1126/science.1135774

- Morphogen: Organ-size dependent growth inducer
- Dpp Signalling pathway and growth promoting activity
  - Dpp controls growth via the Hippo/Yorkie signalling pathway





#### Spatial and temporal patterns of growth rate

Growth rate decays over time and spatial heterogeneities are limited



O. Wartlick et al. F. Jülicher and M. Gonzalez-Gaitan (2011) Science 331:: 1154; doi: 10.1126/science.1200037

O. Wartlick et al. F. Jülicher and M. Gonzalez-Gaitan (2011) Nature Reviews Mol. Cell Biol. 12: 594-604









Hornbruch, A. & Wolpert, L. Cell division in the early growth and morphogenesis of the chick limb. Nature 226, 764-766 (1970).



Boehm, B. et al. and J. Sharpe PLoS Biol. 8, e1000420 (2010).

— How can a gradient of a growth factor induce *uniform* growth? « instructive model »



D. Rogulja & K. Irvine. (2005) Cell, Vol. 123, 449-461

• Hypothesis: Combined effect of [concentration] and mechanical forces (see course #5)



M. Romanova-Michaelides et al. and F. Jülicher and M. Gonzalez-Gaitan. WIREs Dev Biol 2015, 4:591-608. doi: 10.1002/wdev.195



- Morphogen: Organ-size dependent growth inducer
- How can a gradient of a growth factor induce uniform growth?
   The relative (ie. normalised) spatial difference in [concentration] is spatially uniform with
  - The relative (ie. normalised) spatial difference in [concentration] is spatially uniform with an exponential gradient

• 
$$C'/C = cte$$
  $C(x) = C_0 e^{-\frac{3}{7}}$ 



M. Romanova-Michaelides et al. and F. Jülicher and M. Gonzalez-Gaitan. WIREs Dev Biol 2015, 4:591-608. doi: 10.1002/wdev.195



### • Organ-size dependent growth inducer

- How can a gradient of a growth factor induce *uniform* growth?
  - Spatial model: slope of the gradient C'/C = cte  $C(x) = C_0 e^{-\frac{x}{\lambda}}$



Requires gradient scaling so the gradient covers the whole field
 Scalar gradient causes vectorial/nematic gradient:
 eg. Planar polarity and connection to growth
 A local discontinuity in Dpp signalling induces growth
 Growth is blocked if Dpp is uniform at all time



Rogulja, D., Rauskolb, C. & Irvine, K. D. Morphogen control of wing growth through the Fat signaling pathway. *Dev. Cell* **15**, 309–321 (2008).



- Morphogen: Organ-size dependent growth inducer
- How can a gradient of a growth factor induce *uniform* growth?
  - Spatial model: slope of the gradient C'/C = cte  $C(x) = C_0 e^{-\frac{x}{\lambda}}$



Requires gradient scaling so the gradient covers the whole field
 Scalar gradient causes vectorial/nematic gradient:
 eg. Planar polarity and connection to growth
 A local discontinuity in Dpp signalling induces intercalary growth
 Growth is blocked if Dpp is uniform at all time



Tkv253QD: constitutive activation of Dpp signalling causes non autonomous cell proliferation (BrdU)



D. Rogulja & K. Irvine. (2005) Cell, Vol. 123, 449-461

- Morphogen: Organ-size dependent growth inducer
- How can a gradient of a growth factor induce *uniform* growth?
  - Spatial model: sloj



## • Size regulation by the Hippo pathway

#### —A universal cell and tissue growth promoting pathway

Identification of Hippo, a Ste20Kinase that represses cell and tissue growth •



S. Wu et al. and D. Pan (2003) Cell, Vol. 114, 445-456 K Harvey et al . I. Hariharan. (2003) Cell, Vol. 114, 457-467

- Hippo represses the transcription factor Yorkie
- Yorkie promotes growth via expression of Cyclin E and inhibition of apoptosis



J. Huang et al. and D. Pan. (2005) Cell, Vol. 122, 421-434 J. Dong et al. and D. Pan. (2007) Cell 130, 1120-1133

- Phosphorylation of Yorkie by the kinase Warts blocks nuclear translocation and Yorkie activity
- Mammalian orthologue YAP and Lats control tissue growth in the liver, and tumorogenesis •







cell death

cell cycle cell growt



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Overexpression of YAP

## • Size regulation by the Hippo pathway

#### -A universal cell and tissue growth promoting pathway

- The hippo pathway integrates many cellular inputs:
  - polarity cues: epithelial integrity
  - cell contacts: adhesion molecules
  - mechanical stimuli (see cours #5)
  - growth factor signalling: TFGB/Dpp and Fat





Cell or tissue property that regulates the Hippo pathway Apicobasal polarity Mechanotransduction Cell-cell adhesion Contact inhibition Cell or tissue property that Cellular function regulated
by the Hippo pathway Proliferation Cell survival Cell competition Stem cell maintenance Metastasis Regeneration



DPP

Wing 5

Anterior

Dachsous

Posterior

а

b

DPP

— How can a gradient of a growth factor induce *uniform growth*?

- Spatial model: slope of the gradient C'/C = cte  $C(x) = C_0 e^{-\frac{x}{\lambda}}$
- Dpp controls growth via the Fat/Dachsous signalling pathway
- High Dpp signalling affects Fat signalling



Rogulja, D., Rauskolb, C. & Irvine, K. D. Morphogen control of wing growth through the Fat signaling pathway. *Dev. Cell* **15**, 309–321 (2008).





Yorkie

MAD

MEDEA

Yorkie MAD Warts

 $\rightarrow$  Dad, spalt

**→**brk

Yorkie)

(BRK)

Cell membrane

Cytoplasm

Nucleus

Dachsous

h

DPP

Dachsous

Warts

Yorkie

MAD P MEDEA

Yorkie MAD

— How can a gradient of a growth factor induce *uniform growth*?

- Spatial model: slope of the gradient C'/C = cte  $C(x) = C_0 e^{-\frac{x}{\lambda}}$
- Dpp controls growth via the Hippo/Yorkie signalling pathway



Rogulja, D., Rauskolb, C. & Irvine, K. D. Morphogen control of wing growth through the Fat signaling pathway. *Dev. Cell* **15**, 309–321 (2008).



Dachsous

0

Warts

 $\rightarrow$  Dad, spalt

→ brk

(Yorkie)

(BRK)

Cell membrane

⊢ (Hippo) Cytoplasm

Nucleus

- How can a gradient of a growth factor induce *uniform* growth?
  - Complementary roles of the counter gradients of Dpp and Fat



G. Schwank et al. and K. Basler (2011) Developmental Cell 20, 123-130



— How can a gradient of a growth factor induce uniform growth and growth arrest?

- Complementary roles of the counter gradients Dpp and Fat
- Dpp may not determine the gradient of Fat activity
- How could such a system confer robustness? The two gradients are supposed to be independent and yet have perfectly complementary shapes





G. Schwank et al. and K. Basler Developmental Cell 20, 123–130

Rogulja, D., Rauskolb, C. & Irvine, K. D. Morphogen control of wing growth through the Fat signaling pathway. *Dev. Cell* **15**, 309–321 (2008).



#### — How can a gradient of a growth factor induce *uniform* growth?

- Temporal Model: Consider the dynamical properties of the morphogen gradient  $C(x) = C_0 e^{-\frac{x}{\lambda}}$
- In some conditions, the normalised temporal variation of C is a constant:







— Dynamics of morphogen gradient with growth

• At any time, steady state description of an exponential morphogen:  $C(x) = C_0 e^{-rac{x}{\lambda}}$ 



O. Wartlick *et al.* F. Jülicher and M. Gonzalez-Gaitan *Science* **331**, 1154 (2011); doi: 10.1126/science.1200037

<u>COLLÈGE</u>

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1530

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• Renormalisation shows gradient scaling (decay length increases over time)







• Scaling is not due to change in diffusion or production rate, but to reducing degradation constant



• Expander dilution model of gradient scaling: —degradation is reduced over time as inhibitor of morphogen stability is diluted





time [h]



OLLÈGE

FRANCE



• Expander repression model of gradient scaling:



Ben-Zvi, D. & Barkai, N. Proc. Natl Acad. Sci. USA 107, 6924-6929 (2010)

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Dynamics of morphogen gradient with grow

• Experimental evidence of a negative feedback inhibitor of the Dpp

The expander Pentagon is repressed by Dpp (clones mutant for Dpp signalling express *pent*)





Dpp activity gradient is shaped by Pentagon: expansion when Pent is over expressed, reduced in mutant



Wing patterns are reshaped by Pent





pent<sup>2</sup>/pent<sup>2</sup>



Vuilleumier, R. et al. M. Affolter and G. Pyrowolakis. Nature Cell Biol. 12, 611-617 (2010).

— Dynamics of morphogen gradient with growth: growth homogeneity

Growth is uniform so the normalised position of a cell down the gradient is invariant

$$r_{\text{cell}} = x_{\text{cell}}(t)/L(t)$$







—Tissue specific, cellular growth rule:

• Power law relates Dpp at the source and tissue size: Implications



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 $C_0(t) \sim A(t)^{\beta}$  (1)  $C_{\text{cell}}(t) \sim A(t)^{\beta}$  since  $C(r_{\text{cell}}t)/C_0(t)$ is a constant

O. Wartlick et al. F. Jülicher and M. Gonzalez-Gaitan (2011)

Science 331:: 1154; doi: 10.1126/science.1200037



where tissue growth rate is  $(g=\dot{A}/A)$ 



And the cell proliferation rate  $g_{cell} \approx \frac{1}{\beta} \frac{C_{cell}}{C_{cell}}$ 

is proportional to the relative temporal change in Dpp concentration

 $\alpha = \Delta C_{\text{cell}}/C_{\text{cell}} \approx (\Delta C_{\text{cell}}/\theta)/C_{\text{cell}} \text{ where the cell cycle time } \theta \approx \ln 2/g_{\text{cell}}$  $\alpha = \frac{\Delta C_{\text{cell}}}{C_{\text{cell}}} \approx \beta \ln 2 = 0.41$ 

• Thus, cells divide when they experience a 40% increase in Dpp concentration or 50% of intracellular signalling S increase



 $\alpha_{\rm s} = 48\% \pm 2\%$  (  $g_{\rm cell} \approx \frac{\ln 2}{\alpha_{\rm s}} \frac{S_{\rm cell}}{S_{\rm cell}}$ 

• Experimental test: Exogenous system of signalling increase using a constitutive activation of the Dpp receptor Tkv (S in the experiment)  $\dot{S}$ 

 $S_{\text{cell}}$  depends on [RU]

 $\dot{S}_{
m cell}/S_{
m cell}$  is greater at tissue edge where s is lower









O. Wartlick *et al.* F. Jülicher and M. Gonzalez-Gaitan (2011) Science 331:: P454:40014460.1126/science.1200037



#### 

Morphogen: Organ-size dependent growth inducer

# —Tissue specific, cellular growth rule: Cell cycle length is set by the timescale of 50% Dpp signalling increase

• Experimental test: Exogenous system of signalling increase using a constitutive activation of the Dpp receptor Tkv (S in the experiment)  $\dot{S}_{cell}$  depends on [RU]

 $\dot{S}_{
m cell}/S_{
m cell}$  is greater at tissue edge where S is lower

Prediction 1: clones grow larger at the periphery as observed Prediction 2:  $\dot{S}_{cell}/S_{cell} = \beta(\dot{A}/A) = \beta g$  as observed



O. Wartlick *et al.* F. Jülicher and M. Gonzalez-Gaitan (2011) *Science* 331:: 1154; doi: 10.1126/science.1200037



#### —Tissue specific, cellular growth rule:

Cell cycle length is set by the timescale of 50% Dpp signalling increase

• Simulations: Dpp production and diffusion are constant as observed.

Dpp degradation rate decays such that its average value becomes inversely proportional to the cell number (expander model).

A cell divides when the relative increase of the local Dpp level reaches a threshold value 0.5 (« growth rule »).











#### —Tissue specific, cellular growth rule:

#### Possible mechanisms for temporal growth rule: measuring the signal increase rate

Dynamics of TGF- $\beta$  signaling reveal adaptive and pulsatile behaviors reflected in the nuclear localization of transcription factor Smad4

Aryeh Warmflash<sup>a,b</sup>, Qixiang Zhang<sup>b</sup>, Benoit Sorre<sup>a,b</sup>, Alin Vonica<sup>b,1</sup>, Eric D. Siggia<sup>a,2</sup>, and Ali H. Brivanlou<sup>b,2</sup> <sup>a</sup>Center for Studies in Physics and Biology and <sup>b</sup>Laboratory for Molecular Vertebrate Embryology, The Rockefeller University, New York, NY 10065

- A. Warmflash et al. and E. Siggia and A. Brivanlou. (2012) www.pnas.org/cgi/doi/10.1073/pnas.1207607109
- R-Smads (Smad I, 2, 3, 5 and 8) phosphorylation quantitatively follows receptor activation
- But co-activator Smad4, which relays R-Smad activation shows transient activation (nuclear translocation)

and Implications for Embryonic Patterning

Laboratory of Molecular Vertebrate Embryology, The Rockefeller University, New York, NY 10065, USA

B. Sorre, A. Warmflash et al, and E. Siggia (2014) Developmental Cell 30, 334-342

<sup>2</sup>Center for Studies in Physics and Biology, The Rockefeller University, New York, NY 10065, USA

—The rate of pathway activation decays with space.

-Evidence of pathway adaptation by negative feedback

Benoit Sorre,<sup>1,2,3</sup> Aryeh Warmflash,<sup>1,2,3</sup> Ali H. Brivanlou,<sup>1,\*</sup> and Eric D. Siggia<sup>2,</sup>

\*Correspondence: brvnlou@rockefeller.edu (A.H.B.), siggiae@rockefeller.edu (E.D.S.)

**Encoding of Temporal Signals** 

by the TGF-β Pathway

http://dx.doi.org/10.1016/j.devcel.2014.05.022













<sup>3</sup>Co-first author

—In an adaptive system, pathway activity is dependent on speed of activation: dC/dt

#### Is the Dpp gradient required for tissue growth?

- Dpp controls growth by inhibiting its target Brk
- Tissue grows in the absence of both Dpp signalling and Brk but growth is no longer uniform and does not normally arrest
- So Dpp is regulating growth
- Assumes complete absence of Dpp in disc center (incorrect)
- Yet Dpp signalling is most likely changing over time.

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Barrio and Milan. *eLife* 2017;6:e22013. DOI: 10.7554/eLife.22013

Bosch et al. eLife 2017;6:e22546. DOI: 10.7554/eLife.22546



Schwank G, Restrepo S, Basler K. 2008. Development 135:4003-4013. doi: 10.1242/dev.025635, PMID: 1902 9041

— How can a gradient of a growth factor induce growth arrest?

• Spatial model: Flattening of a scaling gradient

-Requires that synthesis rate inversely scales with tissue size

• Temporal model: Degradation rate and Growth Feedback Cell division

A gradient of chemical species regulates growth and growth changes the graded chemical pattern by diffusion and advection

-A theoretical model that incorporates:

(1) Production/degradation/diffusion and advection of the morphogen C,

(2) the growth rule,

predicts:

— The interdependency of growth homogeneity and gradient scaling

depending on critical feedback strength  $\beta_c = 2/(1+\varepsilon) = 1$  if isotropic growth

— Growth arrest (bounded growth) depending on degradation rate kand growth feedback parameter  $\beta$ 

For  $\beta = \beta_c$ , growth is homogeneous but unbounded (scale as 1/t) Away from critical point, growth is not perfectly uniform, but becomes bounded if k > 0, depending on  $\beta$ 



final size when growth arrests:

 $\ell^* = \ell(0) [1 + g_0(0)(1 + \beta_c)/k_0]^{1/2}.$   $\frac{\beta < \beta_c}{k = 0} \quad \frac{\beta > \beta_c}{g \sim t^{-1}} \quad \frac{g \sim t^{-1}}{g \sim t^{-1}}$   $\frac{k = k_0}{g \sim e^{-t/\tau}} \quad \frac{g \sim e^{-t/\tau}}{g \sim e^{-t/\tau}}$ 

Aguilar-Hidalgo D, et al and Jülicher F. *Phys Rev Lett.* (2018); 120(19):198102. doi: 10.1103/PhysRevLett.120.198102.



— Spatial temporal regulation of a growth zone by morphogens and beak shape

• BMP4 expression in the mesenchyme induces its growth



The growth zone shrinks at a constant rate over time until it disappears



A. Abzhanov et al, C. Tabin, Science (2004) Nature. (2006)



J. Fritz et al, A. Abzhanov & M. Brenner. Nature Communications | 5:3700 | DOI: 10.1038/ncomms4700

- Cell growth is controlled intrinsically
- -What are the mechanisms of organ specific tissue size sensing/measurement?
- --- What is measured? Intrinsic « ruler » of growth: « size-meter », scaling.



• Energy delivery/demand unbalance.

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- Organ-size dependent negative feedback on cell growth/proliferation (chalone)
- « Size-meter »: scaling of chalone concentration and tissue size distance between activator and feedback inhibitor mechanical signal that scales with size patterning cue: morphogen gradient
- Organ-size dependent growth inducer: Morphogen and growth arrest: Spatial model: gradient scaling and gradient slope. Temporal model: gradient scaling and rate of signal increase
- Importance of considering robustness of mechanisms: e.g. integral feedback



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## Programmed vs Self-organised regulation of Growth



- hierarchical
- modular
- deterministic rules (ie. genetically encoded)



- no hierarchy
- feedbacks
- statistical rules



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