



INSTITUT PASTEUR

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# Quand les microbes dialoguent avec le système immunitaire

**Gérard EBERL**

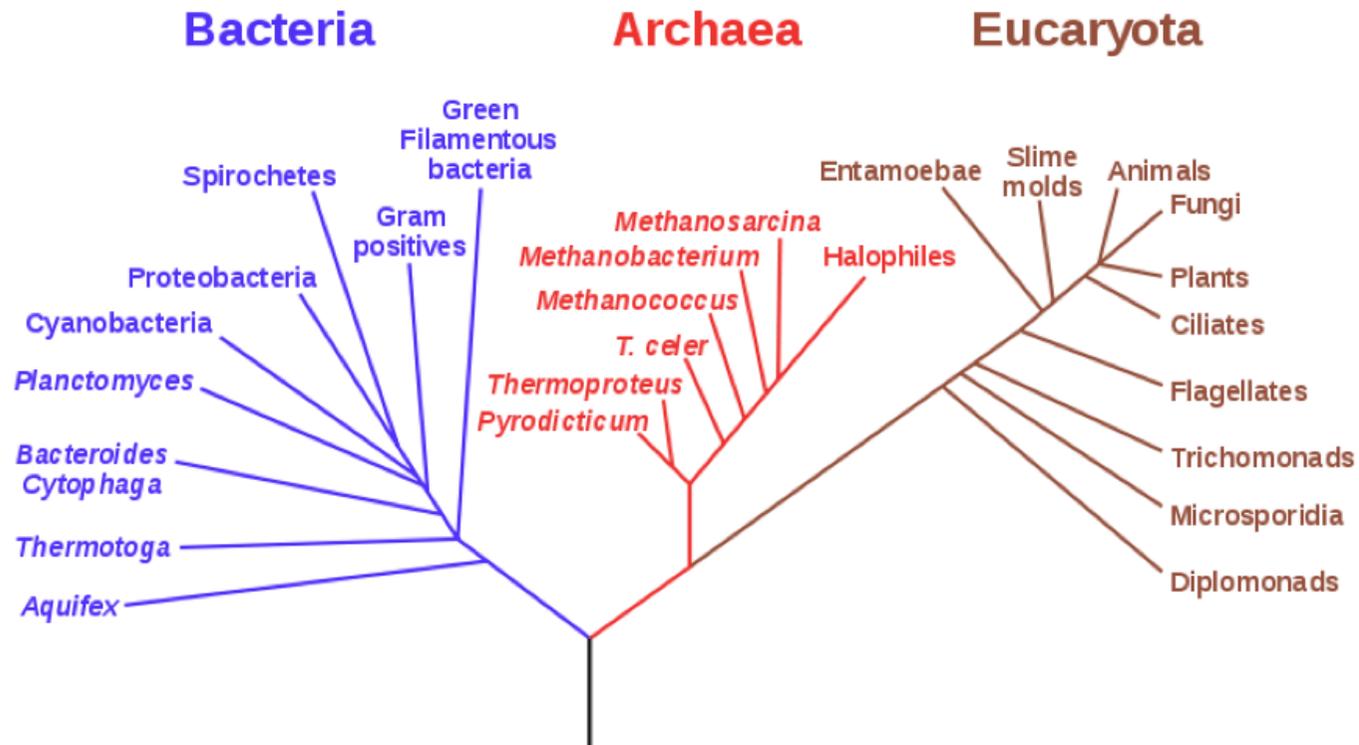
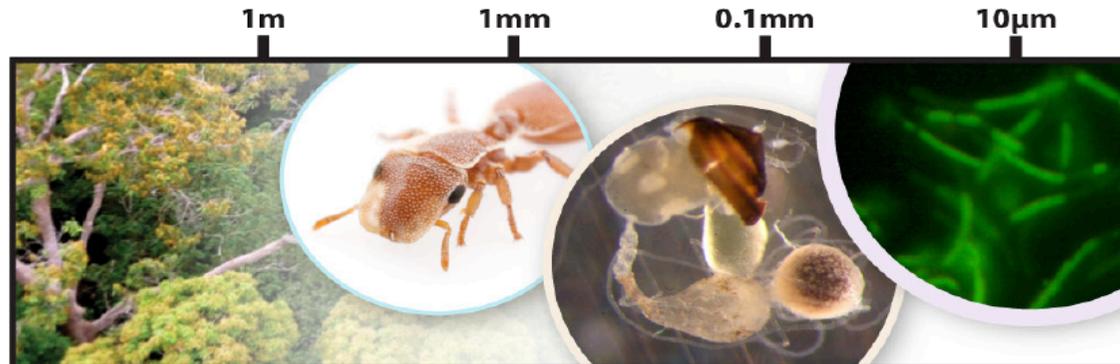
*Unité du Développement des Tissus Lymphoïdes*  
Département d'Immunologie

1. Où sont les microbes?
2. Que fait le système immunitaire?
3. Les microbes et le développement du système immunitaire

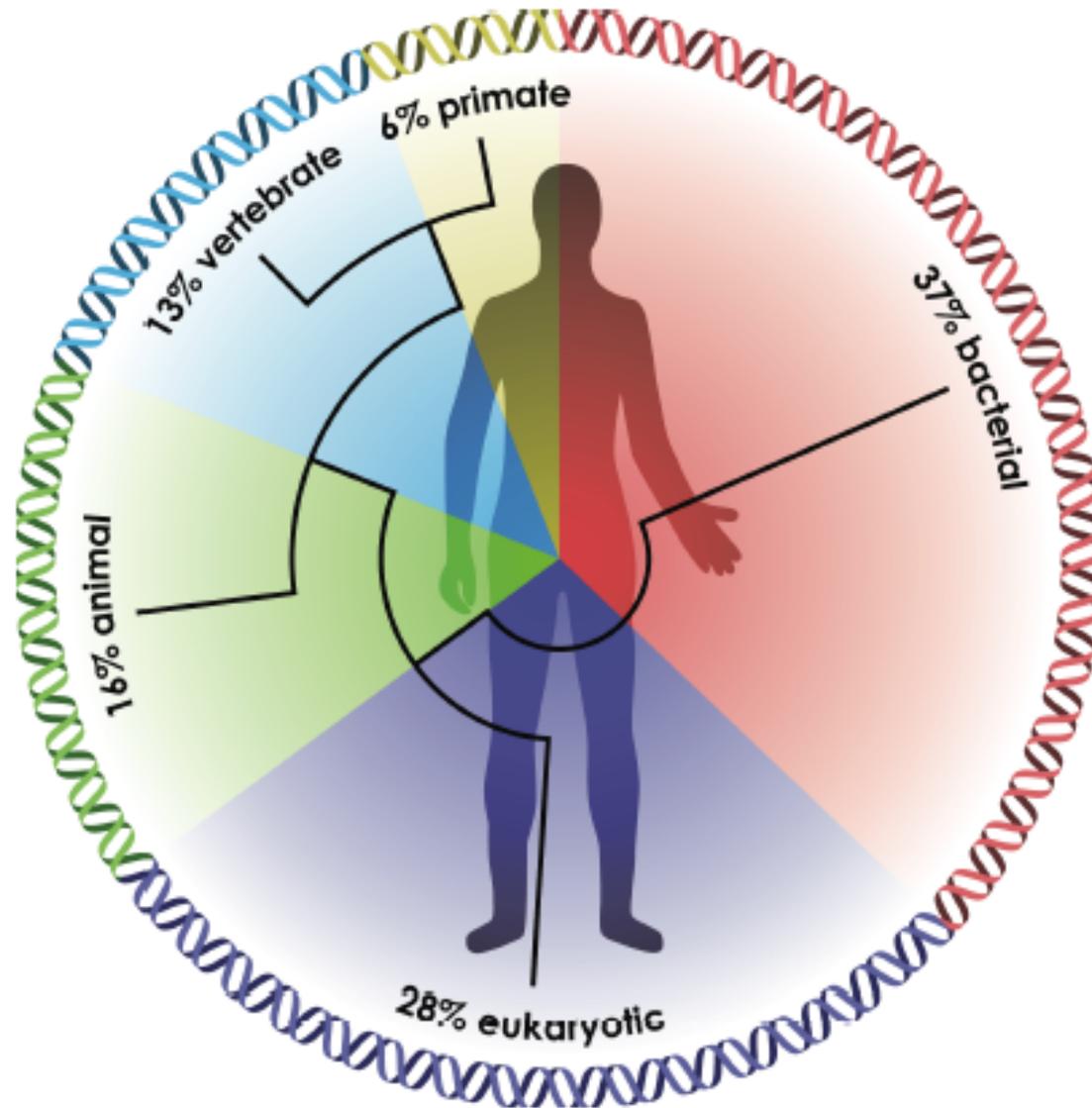
– 1 –

**Où sont les microbes?**

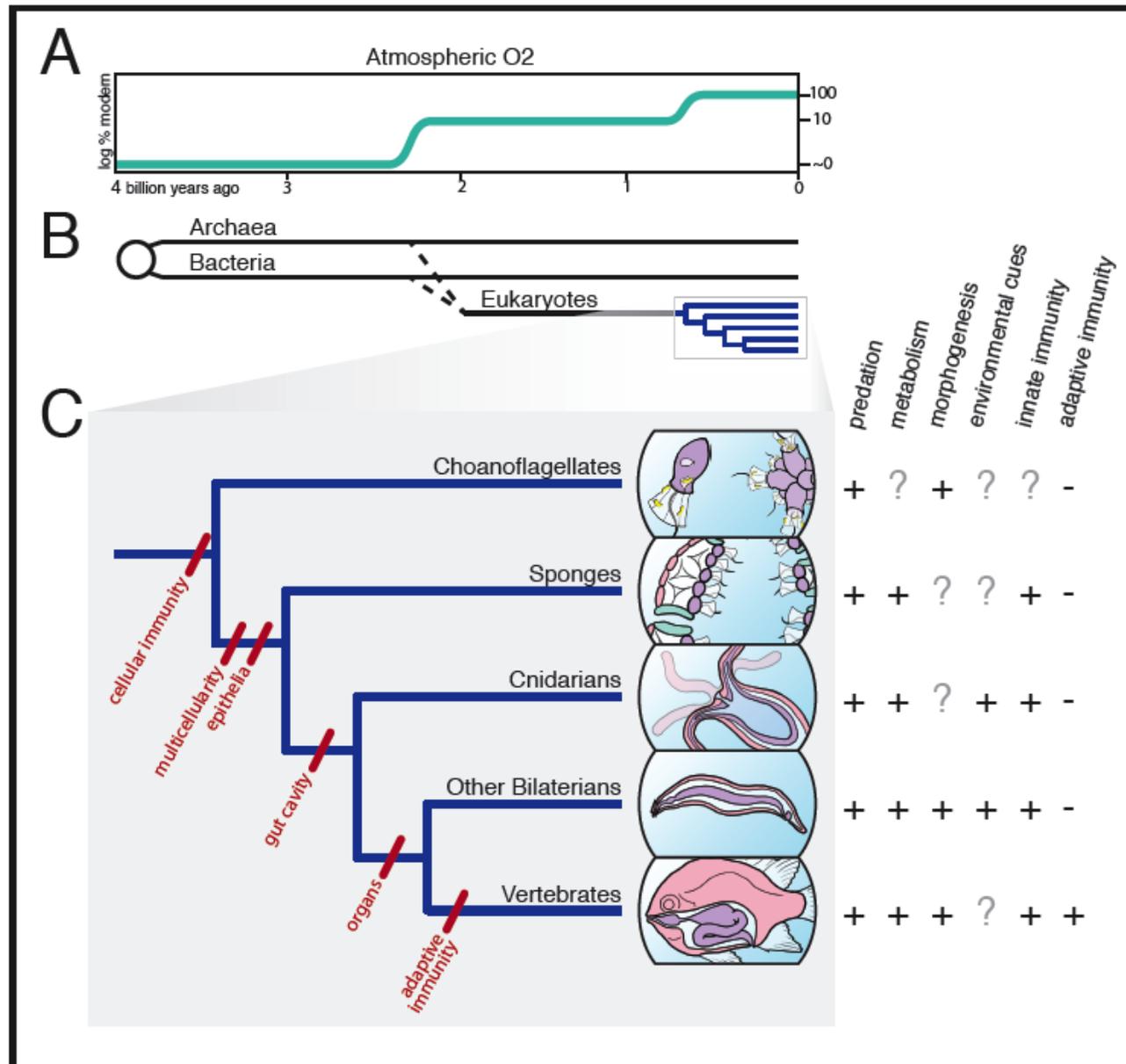
# Notre environnement est essentiellement microbien



Nous sommes génétiquement très microbes



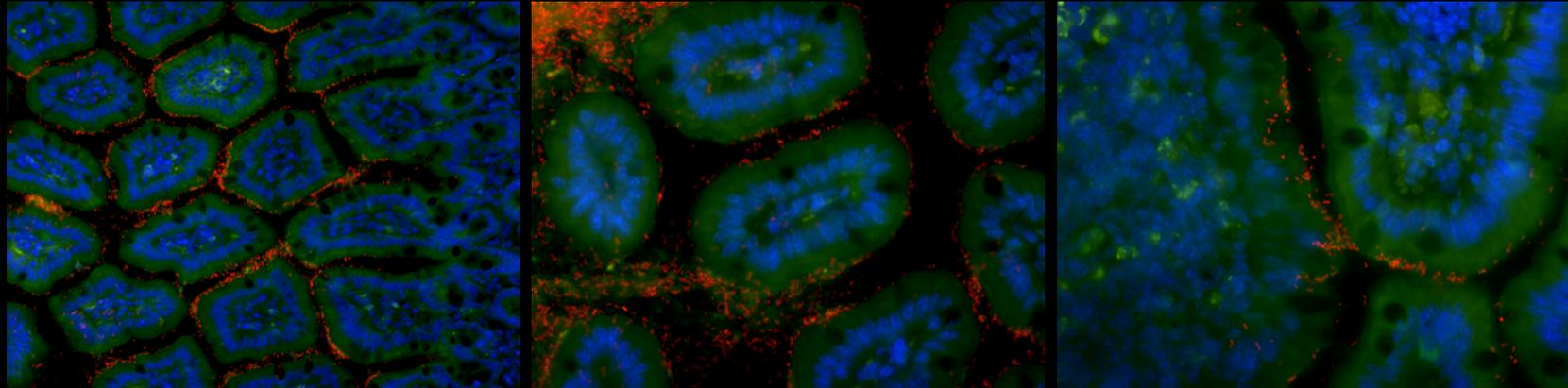
# Les microbes dans l'évolution des animaux



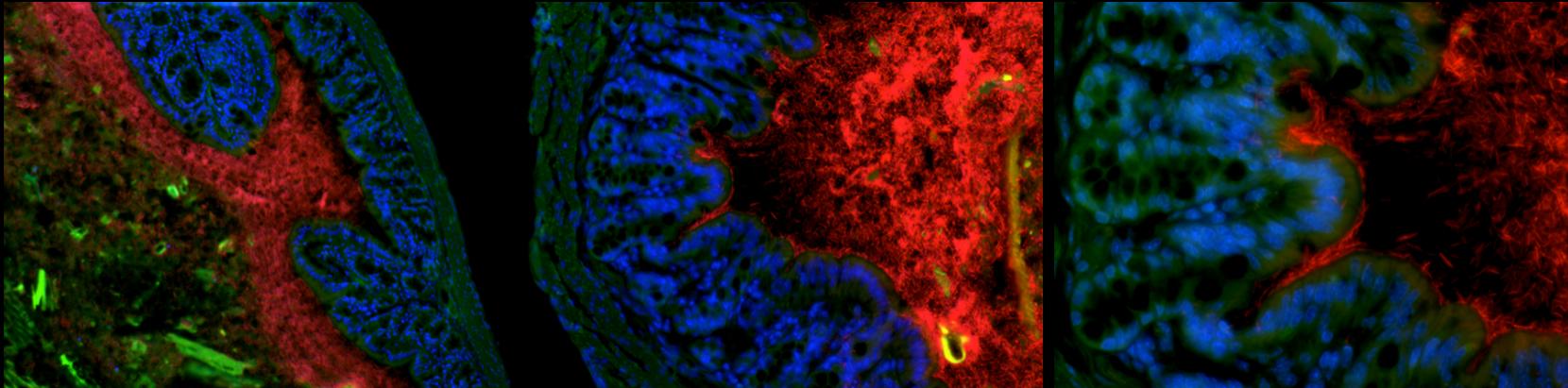
# Les bactéries dans l'intestin

AutoF Bact 16S DAPI

Ileum

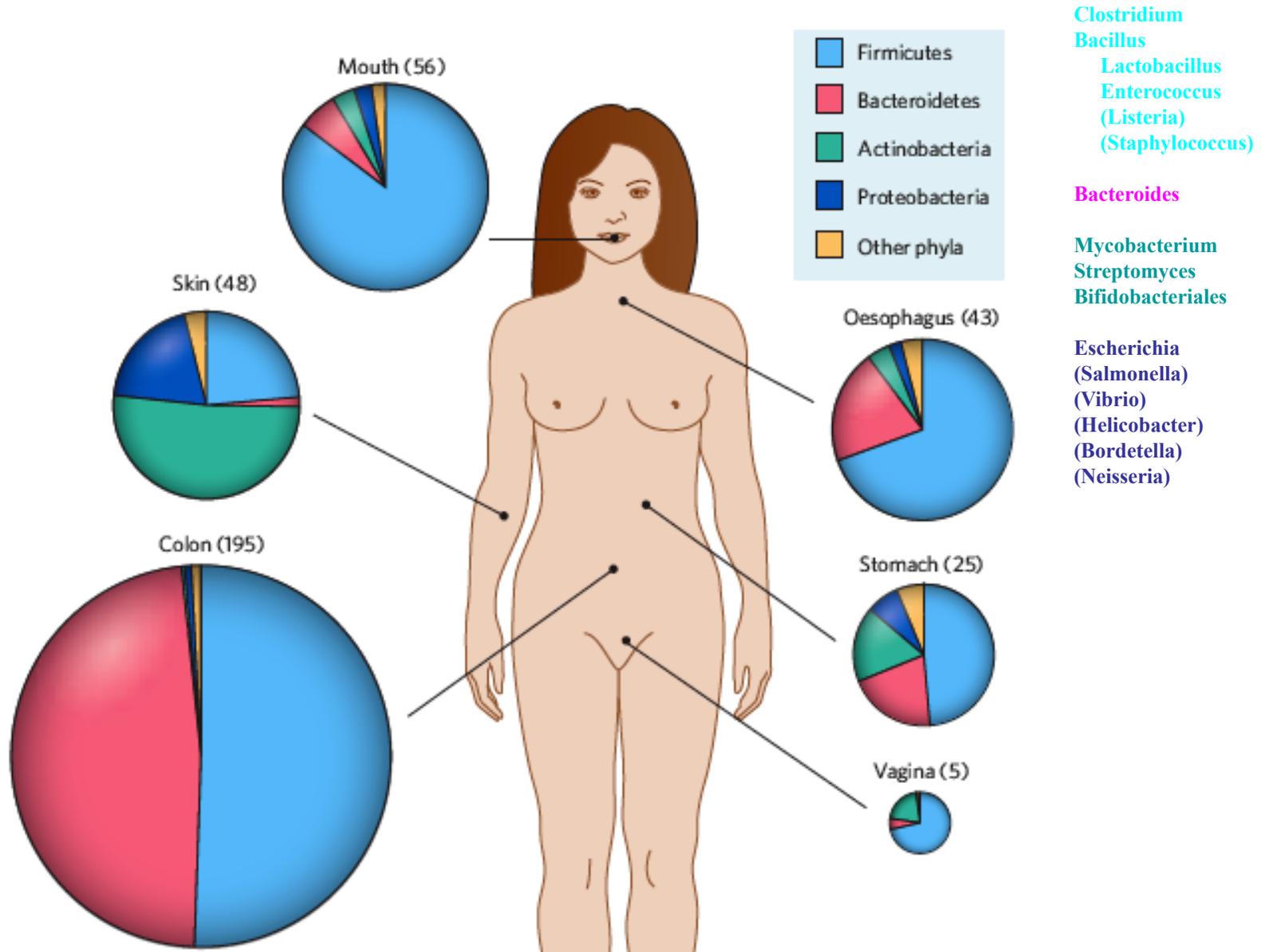


Colon



$\sim 10^{14}$  bactéries and  $\sim 10^{12}$  cellules eucaryotes

# Symbiontes bactériens du corps humain



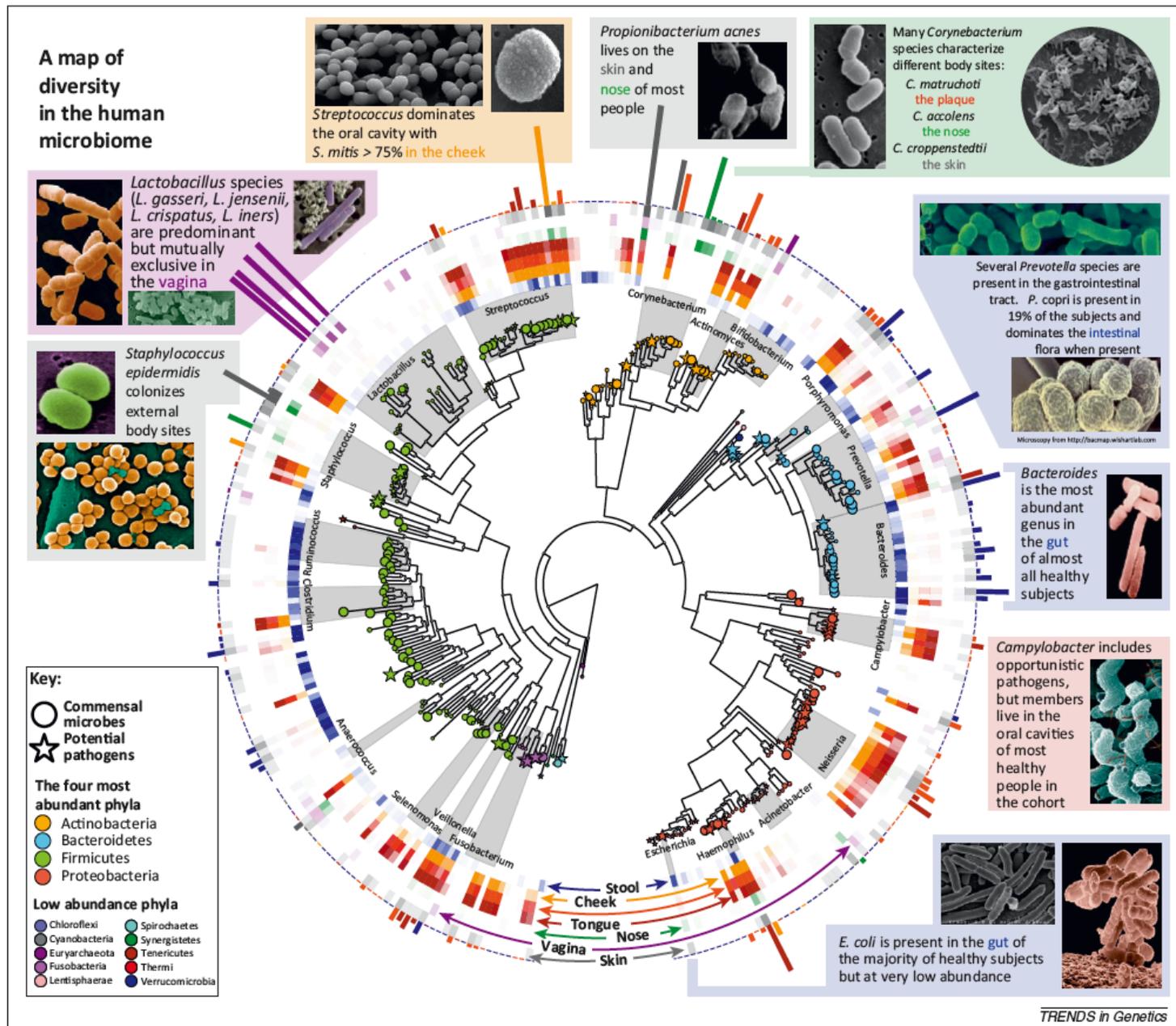
# Symbiontes bactériens du corps humain

## Biodiversity and functional genomics in the human microbiome

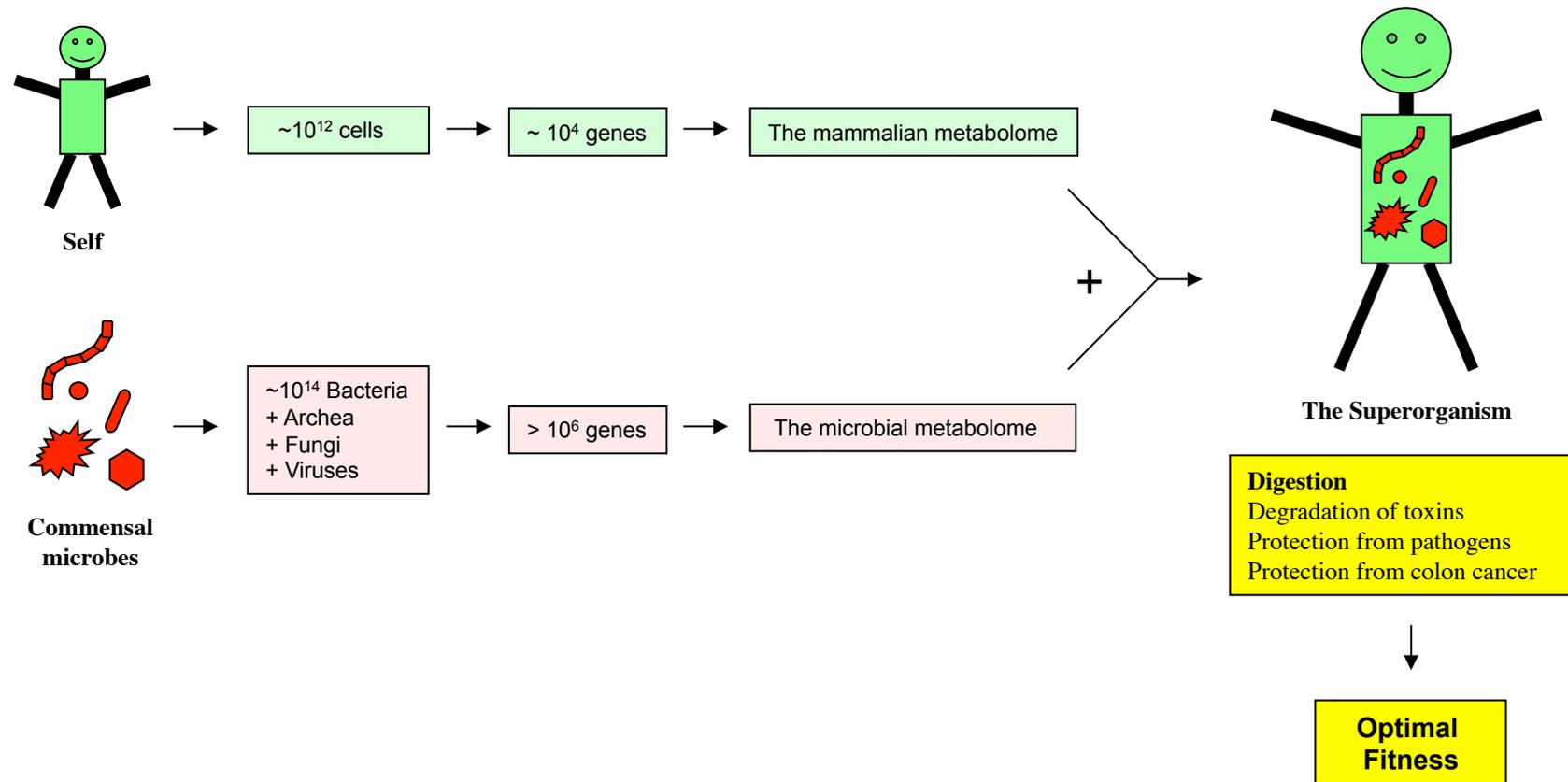
Xochitl C. Morgan<sup>1</sup>, Nicola Segata<sup>1</sup>, and Curtis Huttenhower<sup>1,2</sup>

<sup>1</sup>Department of Biostatistics, Harvard School of Public Health, Boston, MA 02115, USA

<sup>2</sup>Broad Institute of Massachusetts Institute of Technology and Harvard University, Cambridge, MA 02142, USA



# Le superorganisme hôte-microbe



# Symbiosis

*from the Greek:*

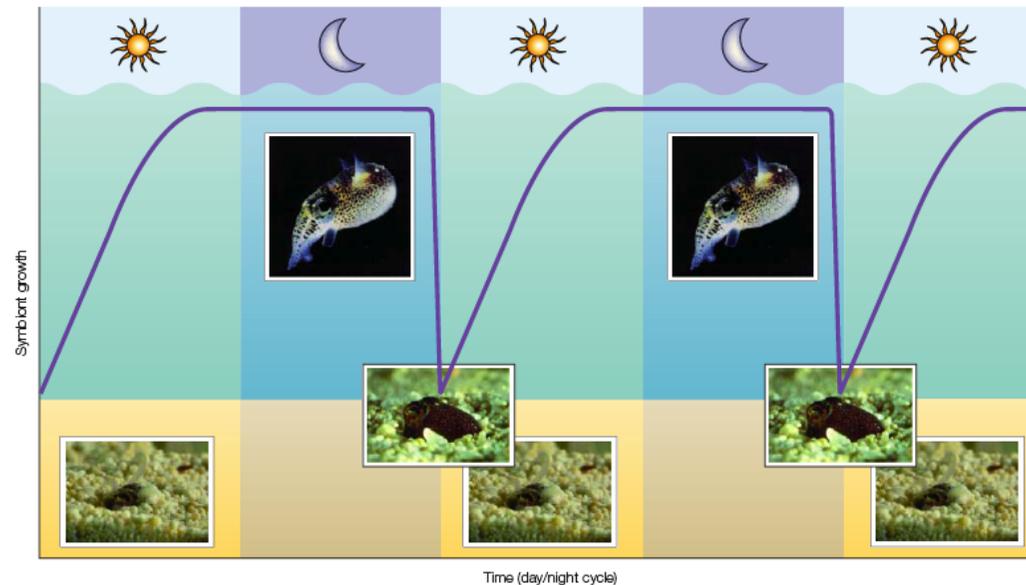
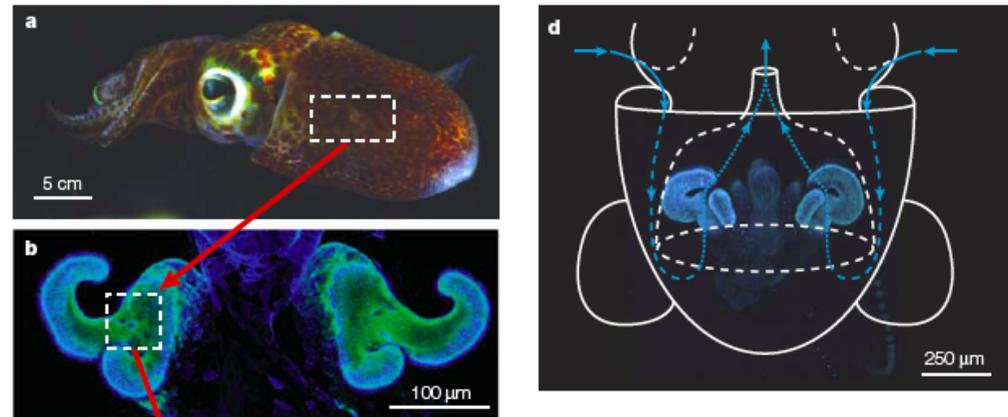
*σύν syn "with"; and βίωσις biosis "living"*

1. Mutualism
2. Commensalism
3. Parasitism

# Microbial Factor-Mediated Development in a Host-Bacterial Mutualism

Tanya A. Koropatnick,<sup>1</sup> Jacquelyn T. Engle,<sup>2</sup> Michael A. Apicella,<sup>3</sup>  
Eric V. Stabb,<sup>4</sup> William E. Goldman,<sup>2</sup> Margaret J. McFall-Ngai<sup>1,5\*</sup>

Science 2004



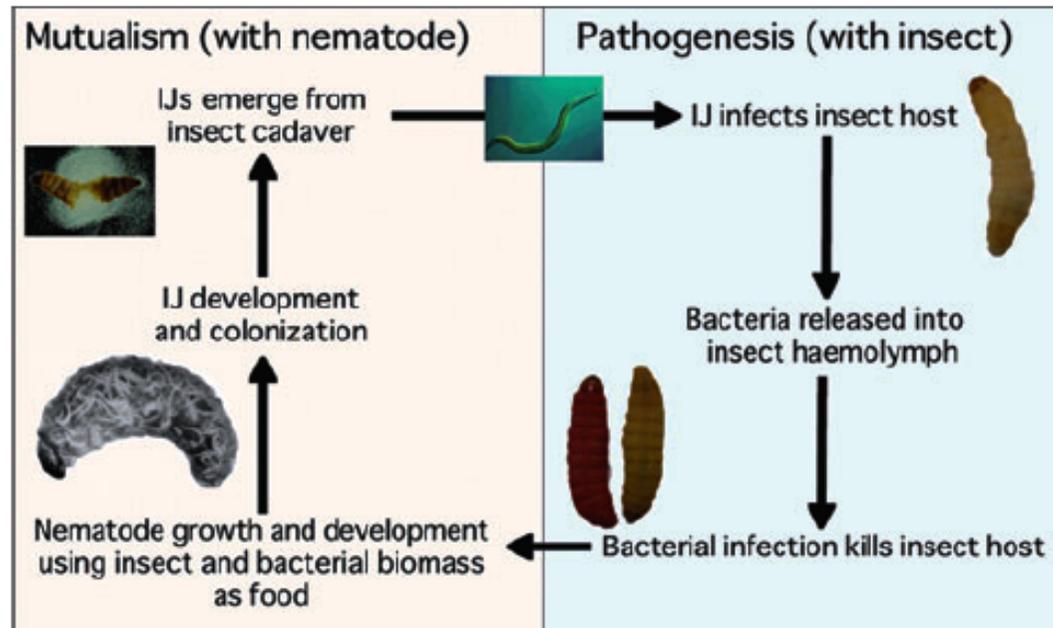
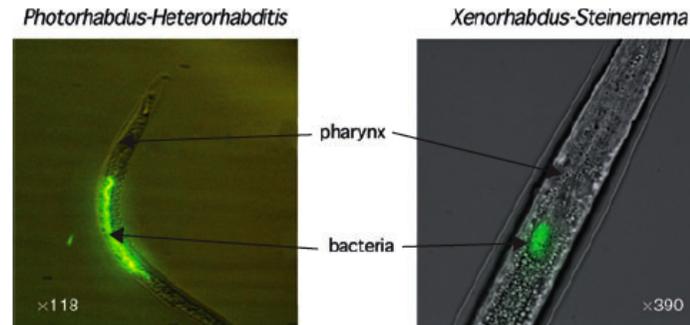
# Mutualism and pathogenesis in *Xenorhabdus* and *Photorhabdus*: two roads to the same destination

Heidi Goodrich-Blair<sup>1</sup> and David J. Clarke<sup>2\*</sup>

<sup>1</sup>Department of Bacteriology, University of Wisconsin, Madison, WI, USA.

<sup>2</sup>Department of Microbiology, University College Cork, Ireland.

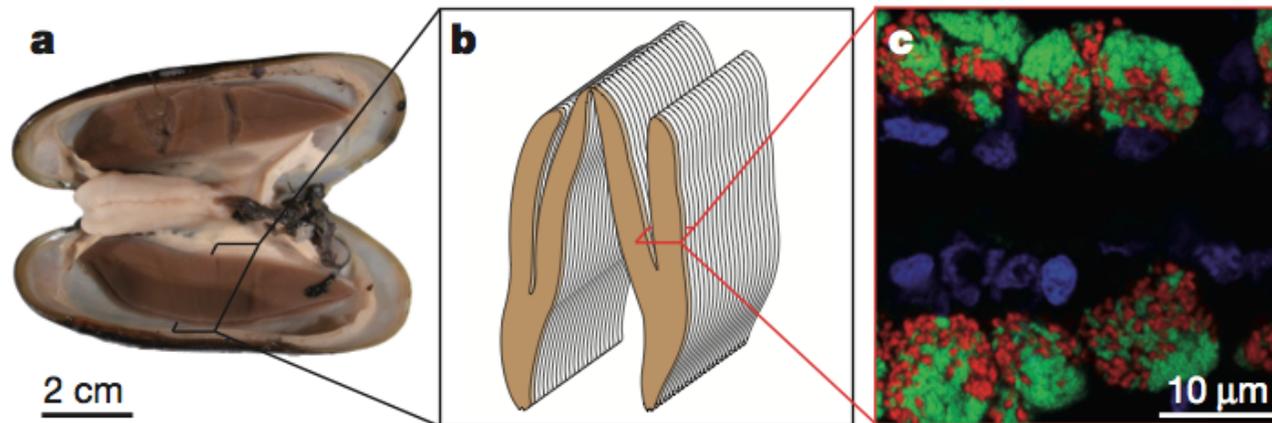
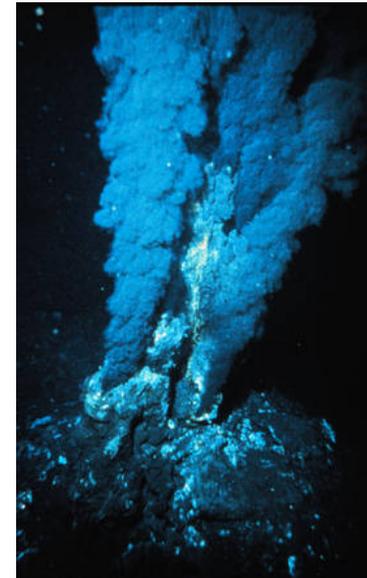
Mol Micr 2007



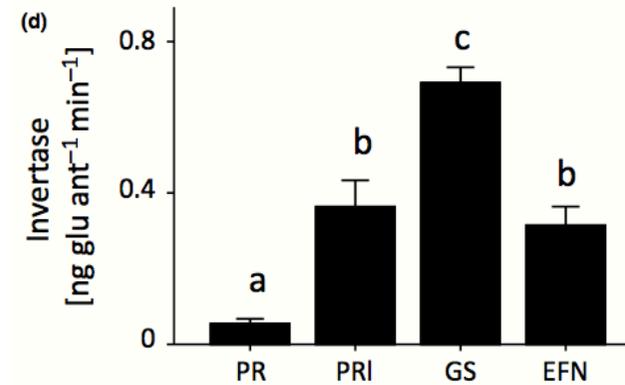
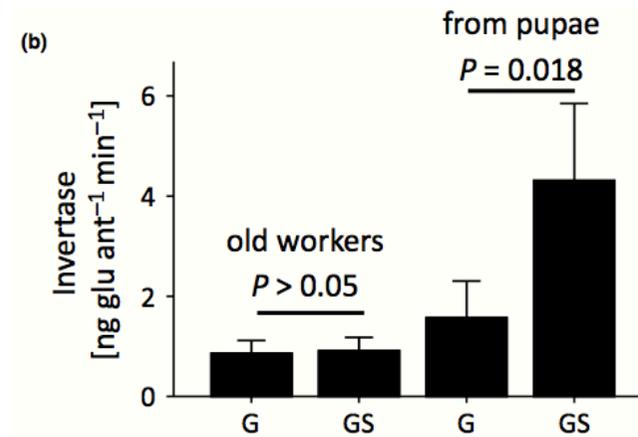
# Hydrogen is an energy source for hydrothermal vent symbioses

Nature 2011

Jillian M. Petersen<sup>1\*</sup>, Frank U. Zielinski<sup>1,2\*</sup>, Thomas Pape<sup>3</sup>, Richard Seifert<sup>4</sup>, Cristina Moraru<sup>1</sup>, Rudolf Amann<sup>1</sup>, Stephane Hourdez<sup>5</sup>, Peter R. Girguis<sup>6</sup>, Scott D. Wankel<sup>6</sup>, Valerie Barbe<sup>7</sup>, Eric Pelletier<sup>7</sup>, Dennis Fink<sup>1</sup>, Christian Borowski<sup>1</sup>, Wolfgang Bach<sup>6</sup> & Nicole Dubilier<sup>1</sup>



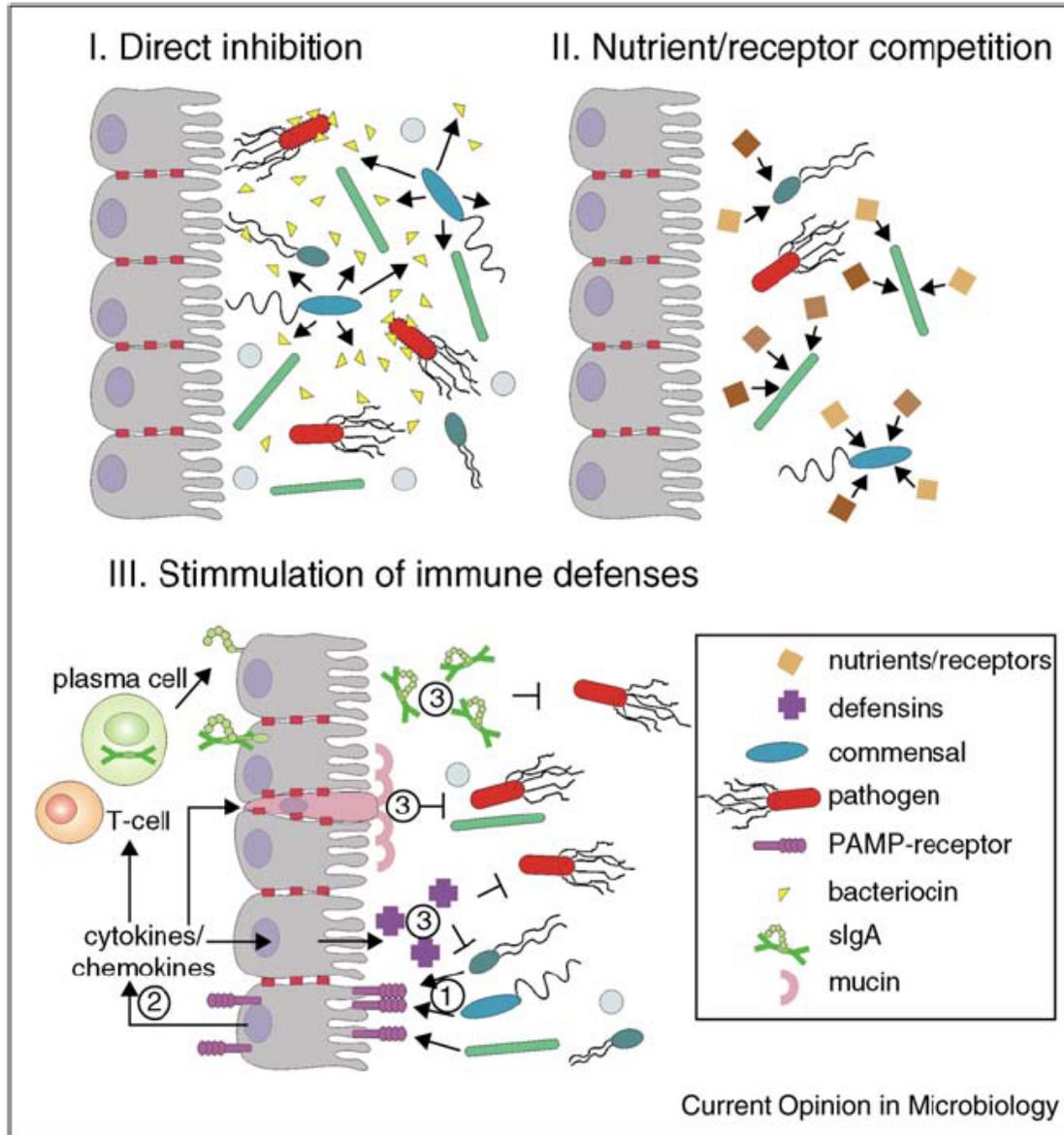
## Partner manipulation stabilises a horizontally transmitted mutualism



# Le rôle des symbiontes bactériens

1. Digestion (extraction d'énergie)
2. Synthèse de métabolites
3. Degradaation de toxines
4. Régulation de l'épithélium
5. Protection contre les pathogènes

# 5. Symbiontes intestinaux et défense



## Mechanisms controlling pathogen colonization of the gut

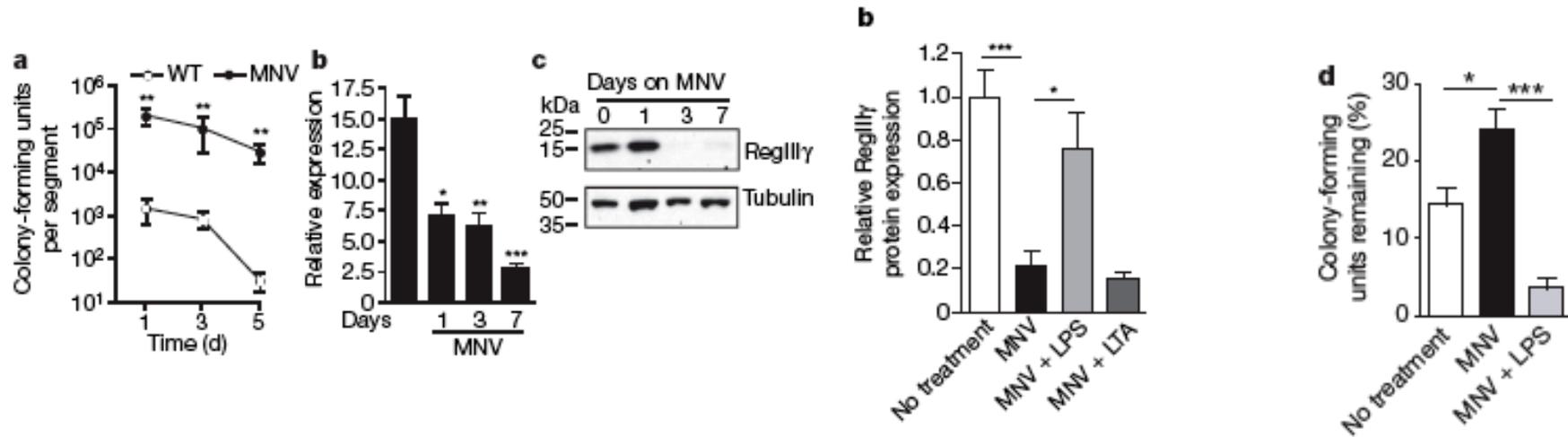
Bärbel Stecher<sup>1</sup> and Wolf-Dietrich Hardt<sup>2</sup>

## 5. Symbiontes intestinaux et défense induite

# Vancomycin-resistant enterococci exploit antibiotic-induced innate immune deficits

Katharina Brandl<sup>1†</sup>, George Plitas<sup>2</sup>, Coralia N. Mihu<sup>1†</sup>, Carles Ubeda<sup>1</sup>, Ting Jia<sup>1</sup>, Martin Fleisher<sup>3</sup>, Bernd Schnabl<sup>4†</sup>, Ronald P. DeMatteo<sup>2</sup> & Eric G. Pamer<sup>1,3</sup>

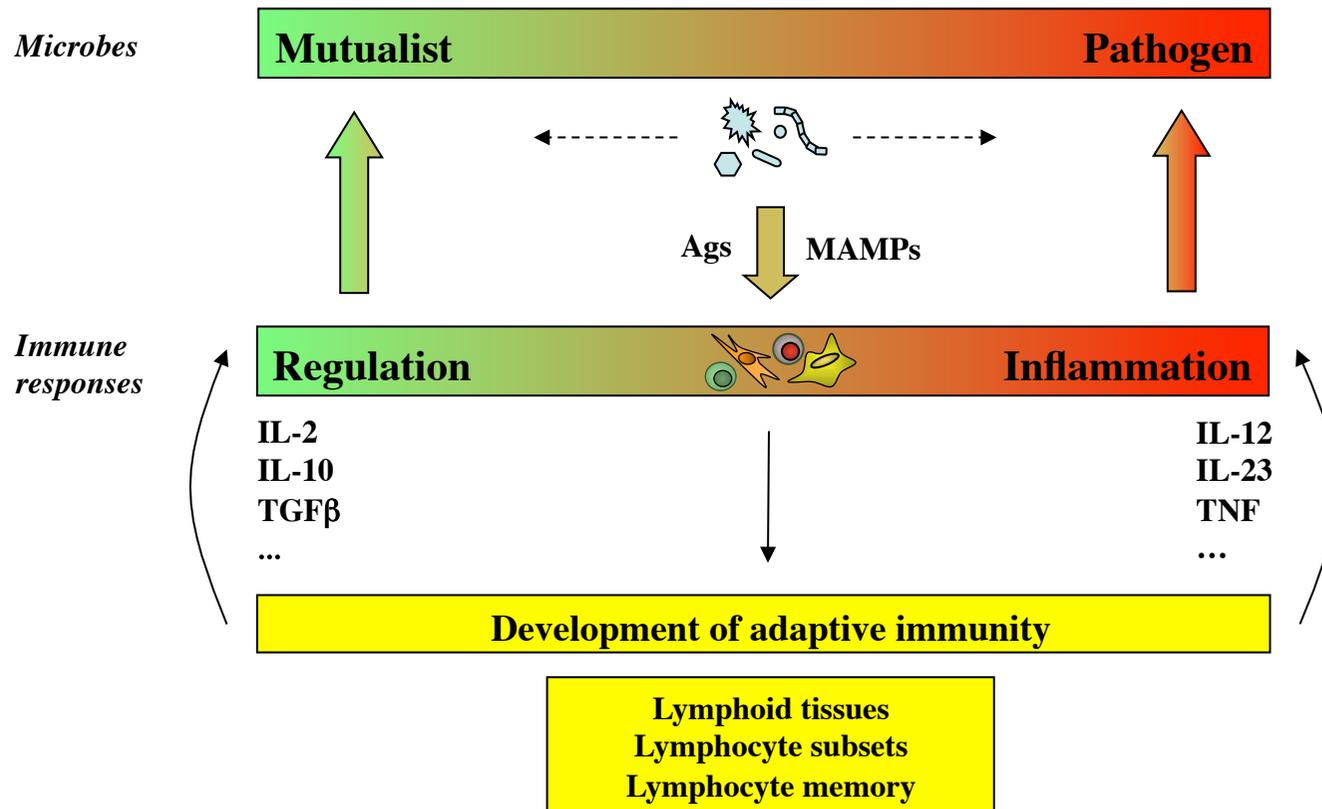
Nature 2008



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**Que fait le système immunitaire?**

# Le système immunitaire maintient l'équilibre

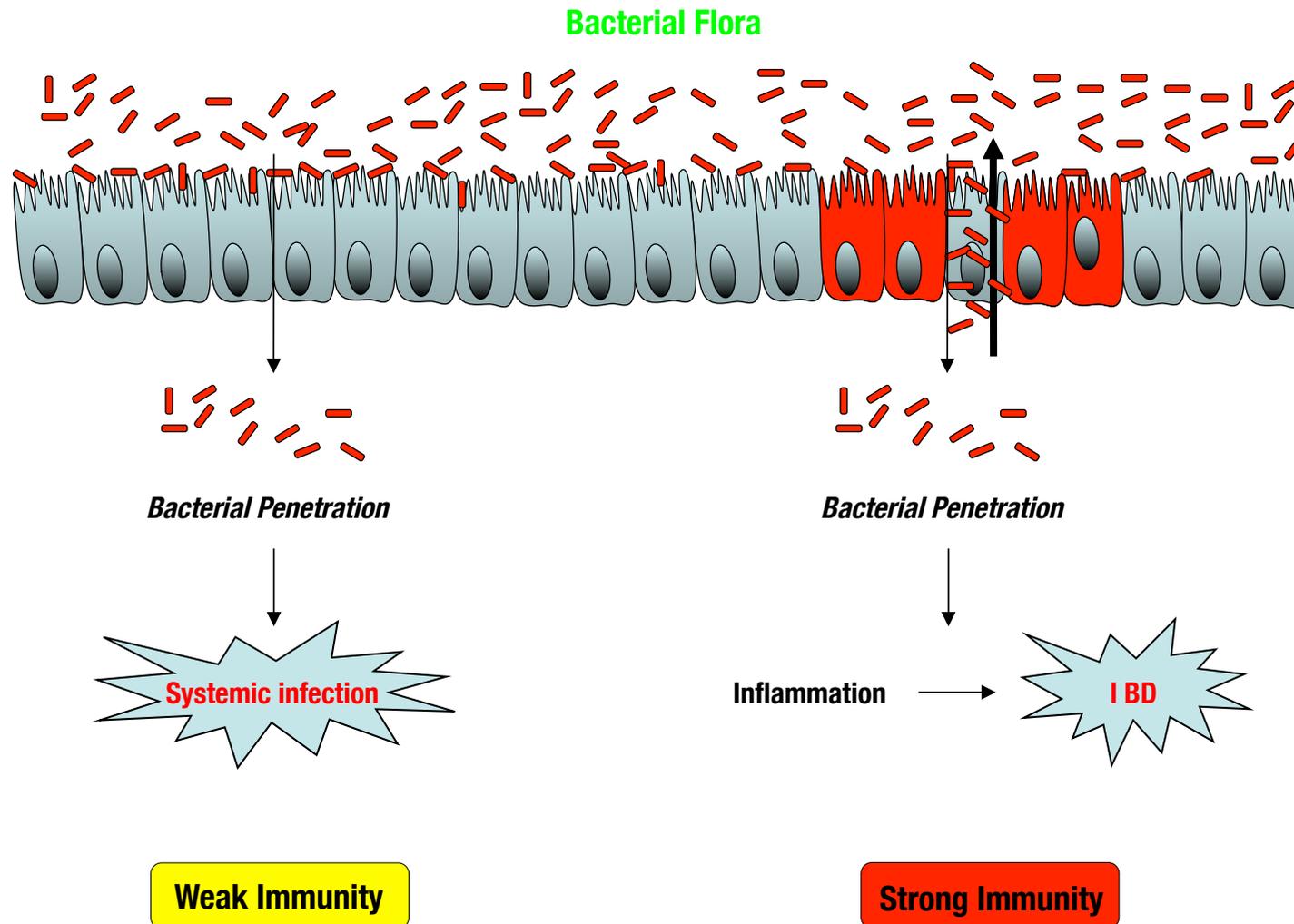


# L'équilibre hôte-symbionte: la clé de la santé

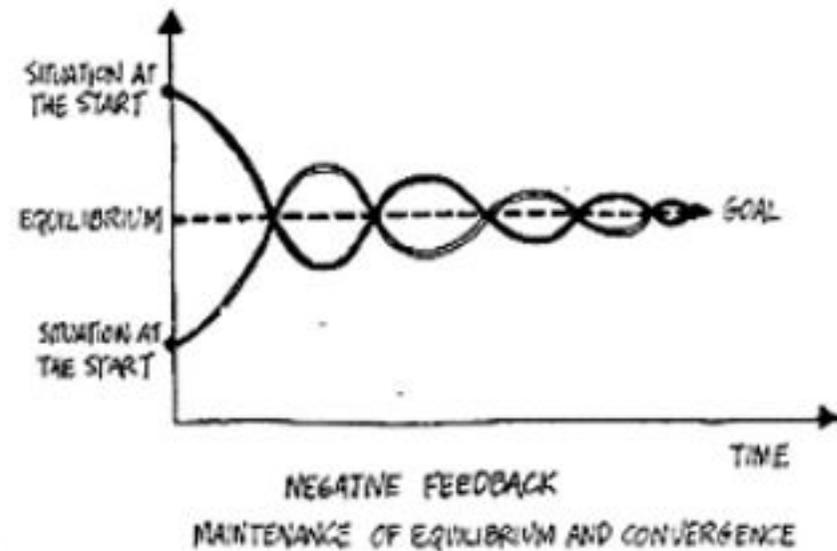
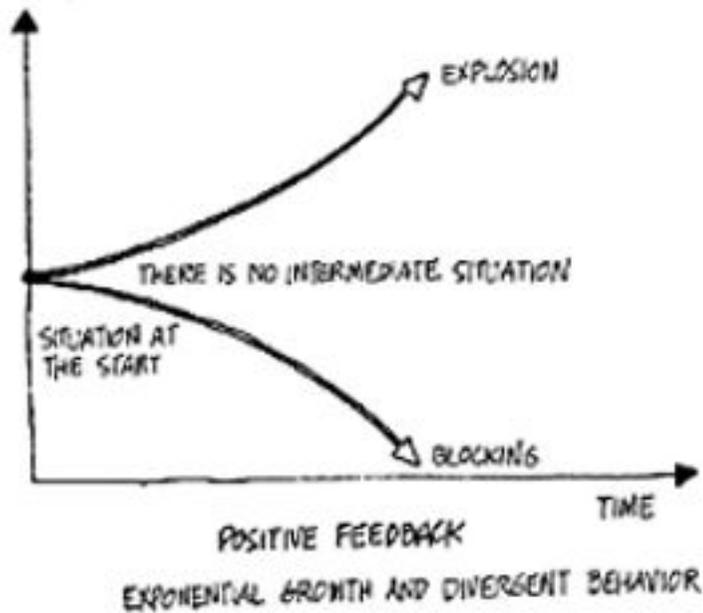
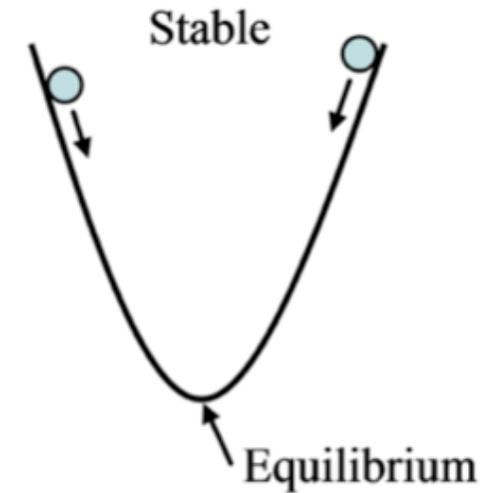
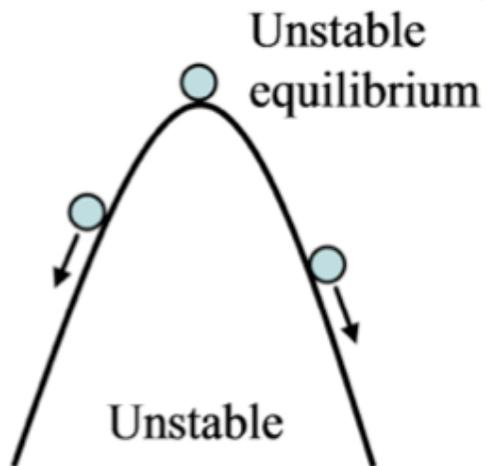
L'outil de l'hôte: le système immunitaire



# L'équilibre entre le microbiote intestinal et le système immunitaire



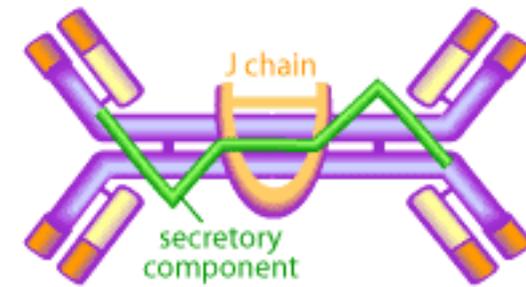
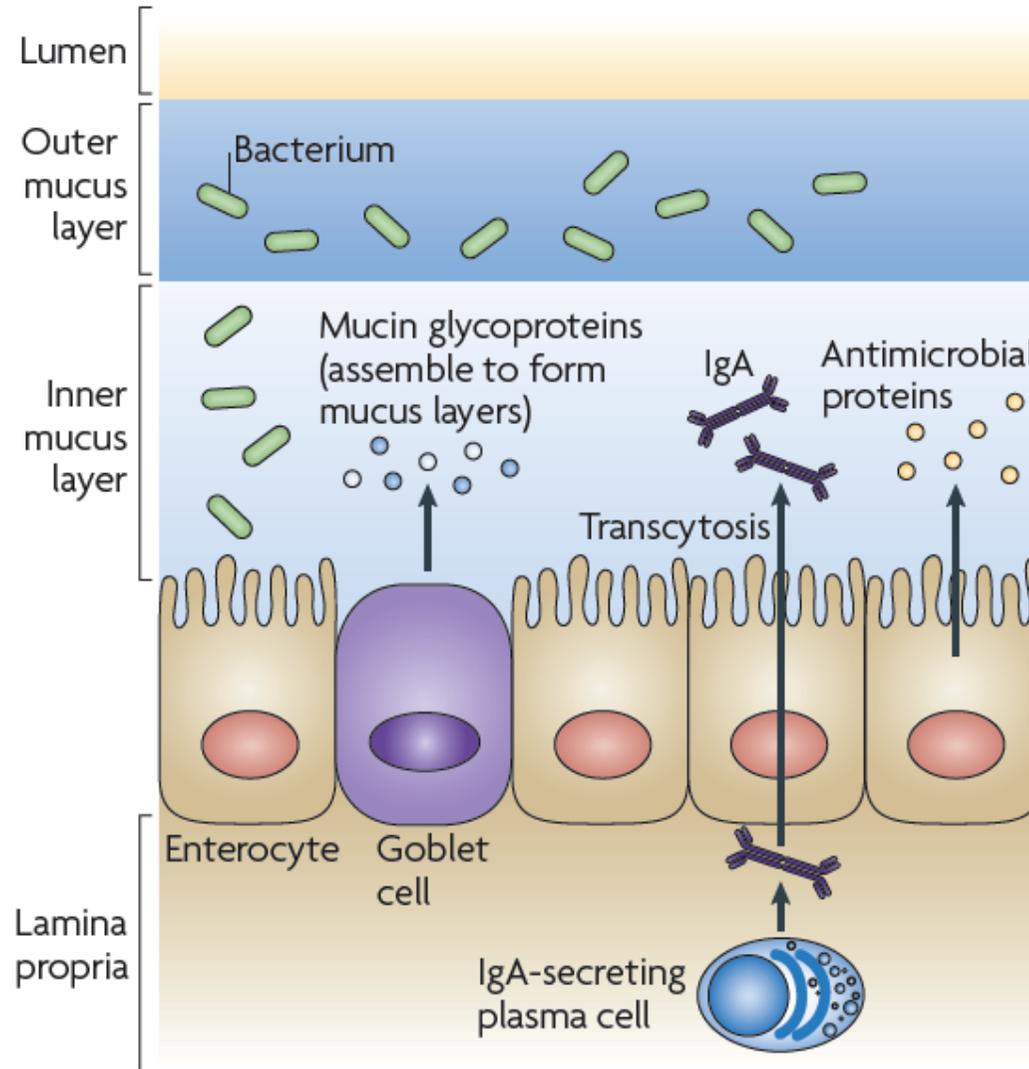
# L'équilibre hôte-microbe est en général robuste



# Le système immunitaire intestinal

1. Immunité innée
2. Immunité adaptative

# La barrière intestinale établie par le système immunitaire



Immune adaptations that maintain homeostasis with the intestinal microbiota

# PRIMATE DEFENSINS

Robert I. Lehrer

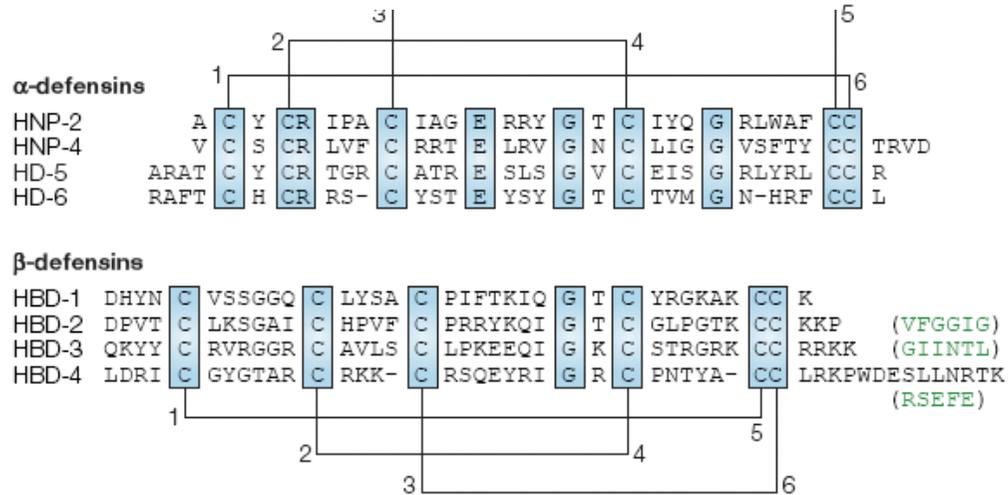
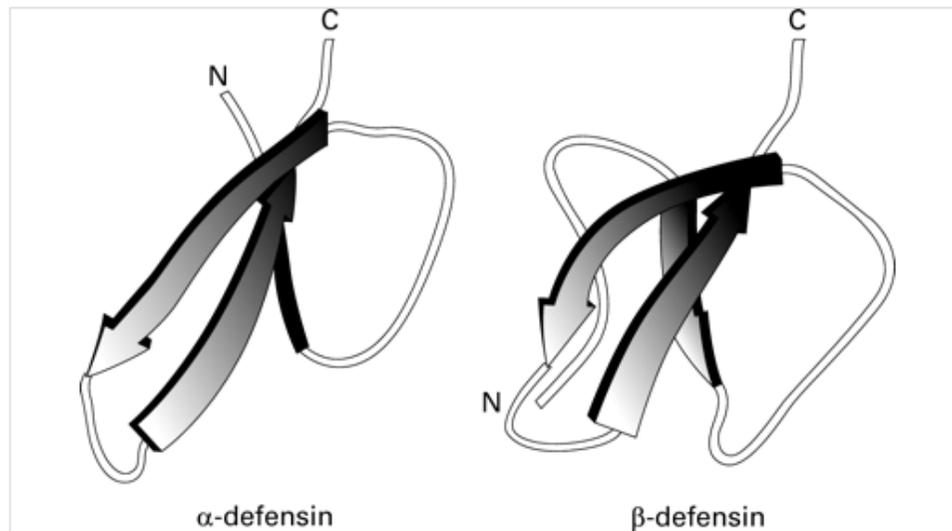
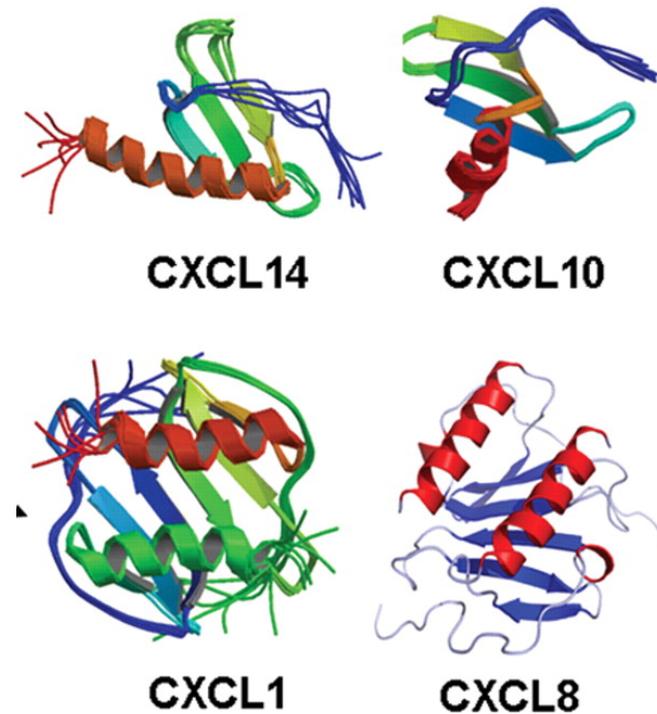
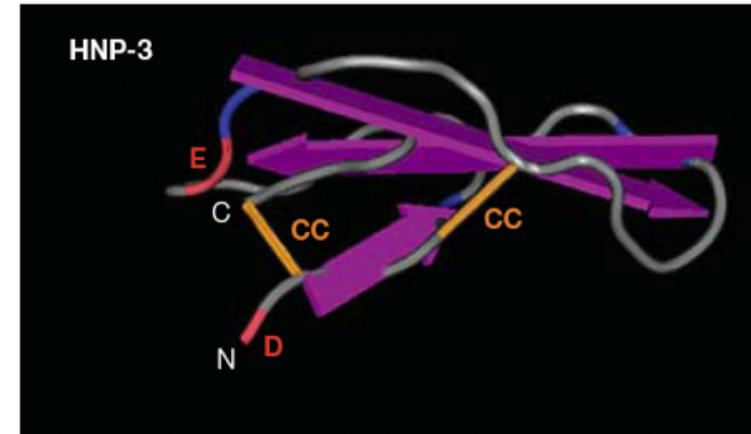


Figure 2 | The sequence and cysteine bonding of four human α-defensins (HNP-2, HNP-4, HD-5 and HD-6) and four human β-defensins (HBD-1–4). HNP-2 and -4 are expressed in



FOCUS ON ANTIMICROBIAL STRATEGIES

Nat Rev Med 2004



# La majorité des chimiokines sont des défensines

| Chemokine            |                           | Calculated<br>pI                | Antimicrobial activity (% Killed, mean±SD)* |                  |           |
|----------------------|---------------------------|---------------------------------|---|------------------|-----------|
| Family               | Member                    |                                 | <i>E. coli</i>                              | <i>S. aureus</i> |           |
| CXC                  | CXCL1/Gro $\alpha$        | 9.80                            | ++++ (87±13)                                | + (22±11)        |           |
|                      | CXCL2/Gro $\beta$         | 10.27                           | +++ (78±9)                                  | + (26±9)         |           |
|                      | CXCL3/Gro $\gamma$        | 9.85                            | +++ (64±11)                                 | + (39±10)        |           |
|                      | CXCL6/GCP-2               | 9.06                            | - (0)                                       | - (0)            |           |
|                      | CXCL8/IL-8                | 8.97                            | - (0)                                       | - (0)            |           |
|                      | CXCL9/MIG                 | 10.83                           | ++++ (100)                                  | ++++ (81±11)     |           |
|                      | CXCL10/IP-10              | 10.52                           | ++++ (100)                                  | +++ (77±12)      |           |
|                      | CXCL11/I-TAC              | 10.48                           | +++ (72±10)                                 | +++ (69±9)       |           |
|                      | CXCL12/SDF-1 <sup>†</sup> | 10.51                           | ++++ (86±12)                                | ++ (59±7)        |           |
|                      | CXCL13/BCA-1              | 10.91                           | ++++ (83±9)                                 | ++ (50±10)       |           |
|                      | CXCL14/BRAK               | 10.28                           | ++ (50±10)                                  | + (35±8)         |           |
|                      | CX3C                      | CX3CL1/fractalkine <sup>‡</sup> | 9.78  | - (0)            | - (0)     |
|                      | C                         | XCL1/lymphotactin               | 11.35                                       | ++++ (100)       | ++ (47±9) |
|                      | CC                        | CCL1/I-309                      | 10.10                                       | ++++ (88±12)     | + (37±5)  |
| CCL2/MCP-1           |                           | 9.58                            | - (0)                                       | - (0)            |           |
| CCL3/MIP-1 $\alpha$  |                           | 4.60                            | - (0)                                       | - (0)            |           |
| CCL5/RANTES          |                           | 9.25                            | - (0)                                       | - (0)            |           |
| CCL7/MCP-3           |                           | 10.10                           | - (0)                                       | - (0)            |           |
| CCL8/MCP-2           |                           | 10.38                           | + (26±12)                                   | - (0)            |           |
| CCL11/eotaxin        |                           | 10.35                           | +++ (66±7)                                  | ++ (49±10)       |           |
| CCL13/MCP-4          |                           | 10.40                           | +++ (76±9)                                  | - (0)            |           |
| CCL16/LEC            |                           | 8.71                            | - (0)                                       | - (0)            |           |
| CCL17/TARC           |                           | 9.29                            | +++ (59±10)                                 | ++ (46±8)        |           |
| CCL18/PARC           |                           | 9.39                            | ++++ (100)                                  | +++ (69±9)       |           |
| CCL19/MIP-3 $\beta$  |                           | 10.16                           | ++++ (82±10)                                | - (0)            |           |
| CCL20/MIP-3 $\alpha$ |                           | 10.08                           | ++++ (100)                                  | ++ (51±7)        |           |
| CCL21/SLC            |                           | 10.46                           | ++ (45±9)                                   | +++ (75±8)       |           |
| CCL22/MDC            |                           | 9.04                            | ++++ (100)                                  | +++ (74±11)      |           |
| CCL25/TECK           | 10.69                     | +++ (68±13)                     | ++ (48±12)                                  |                  |           |
| CCL27/CTAK           | 9.11                      | - (0)                           | - (0)                                       |                  |           |

**Microbe associated molecular patterns  
(MAMPs)**



**Pattern recognition proteins  
(PRRs)**

# Détecteurs innés (Prix Nobel de Médecine 2011)

*Jules Hofmann, Bruno Lemaître (Strasbourg)*

*Bruce Beutler (Scripps, CA)*



Figure 5. Germinating Hyphae of *A. fumigatus* on a Dead *Drosophila*  
Scanning electron micrograph of a *Drosophila* adult that succumbed  
to infection by *A. fumigatus* and is covered with germinating hyphae  
(200× magnification).

# A quoi sert l'immunité adaptative?

NATURE|Vol 445|11 January 2007

**ESSAY**

**ADAPTIVE IMMUNITY**

## Care for the community

A memory-based immune system may have evolved in vertebrates because of the need to recognize and manage complex communities of beneficial microbes.

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**Margaret McFall-Ngai**

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# **Les microbes et le développement du système immunitaire**

# Microbial Factor-Mediated Development in a Host-Bacterial Mutualism

Tanya A. Koropatnick,<sup>1</sup> Jacquelyn T. Engle,<sup>2</sup> Michael A. Apicella,<sup>3</sup> Eric V. Stabb,<sup>4</sup> William E. Goldman,<sup>2</sup> Margaret J. McFall-Ngai<sup>1,5\*</sup>

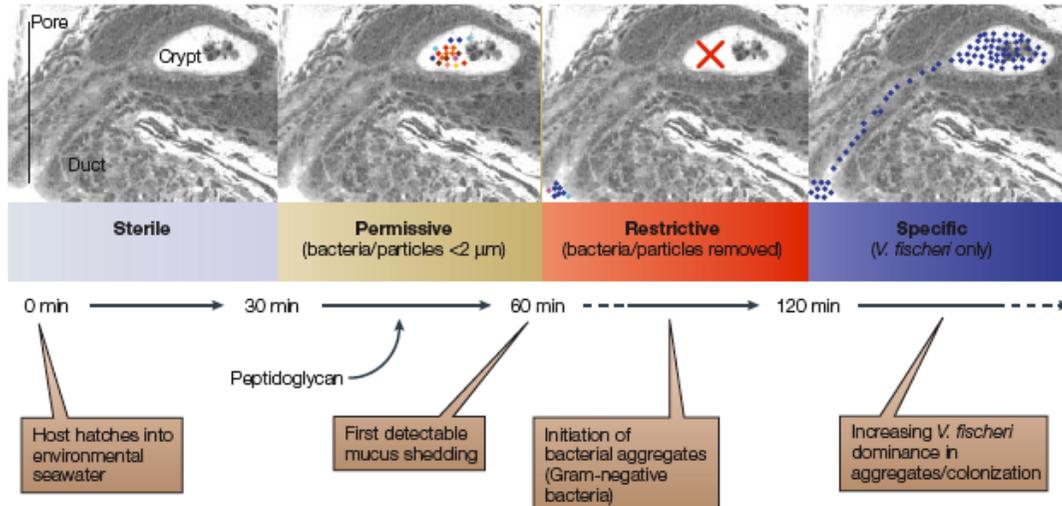
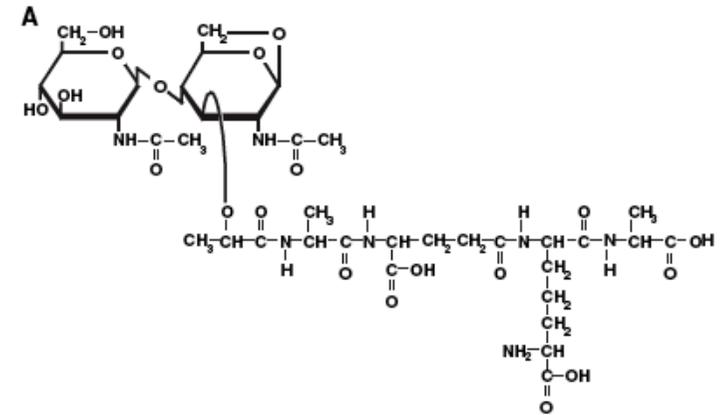
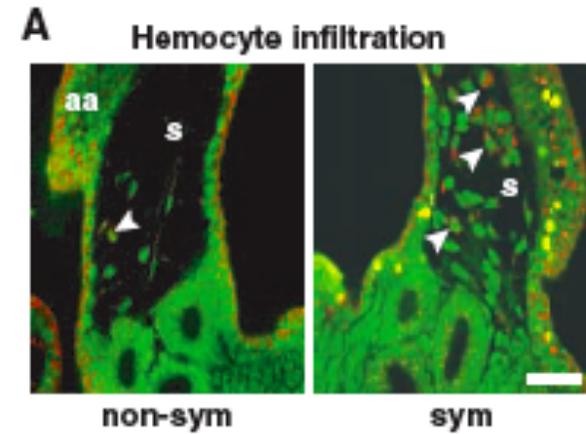


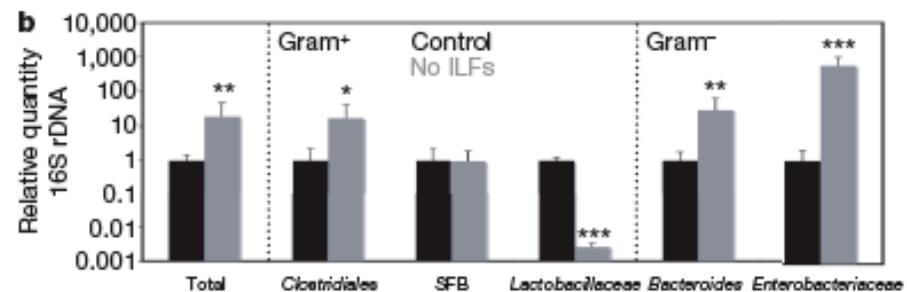
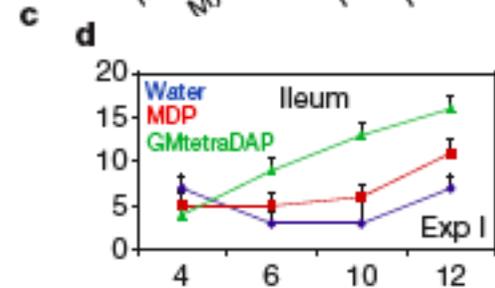
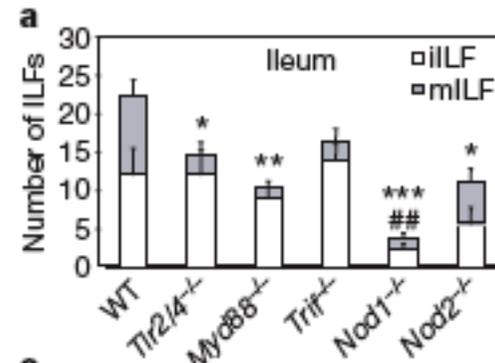
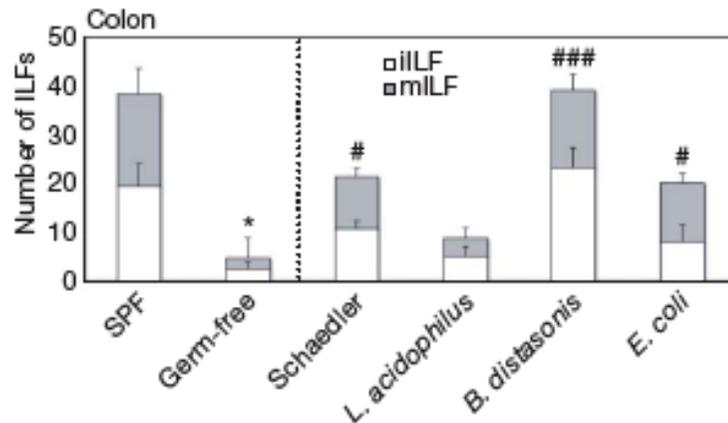
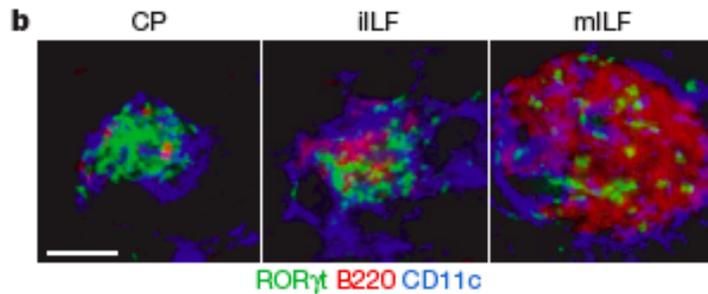
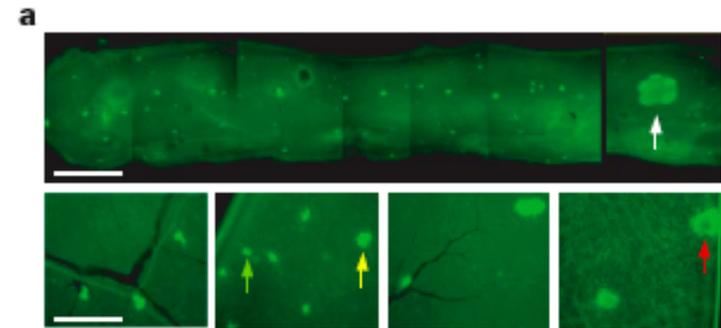
Figure 3 | **The initial interactions of the juvenile host with the environment.** Between 30 and 60 minutes after hatching, the light-organ crypts are open to small numbers (1–3) of bacterial cells or particles less than  $2 \mu\text{m}$  in diameter. These initial entrants are later removed by an as-yet-unknown mechanism. After 1 hour, the host sheds mucus in response to bacterial peptidoglycan and by 2 hours, *V. fischeri* ( $\sim 1 \mu\text{m}$  in diameter) begin to aggregate above the pore and then migrate through the duct before colonizing the crypt epithelium. At this point, the light organ transforms from a ‘permissive’ environment to an environment that is exclusive to the symbiont.



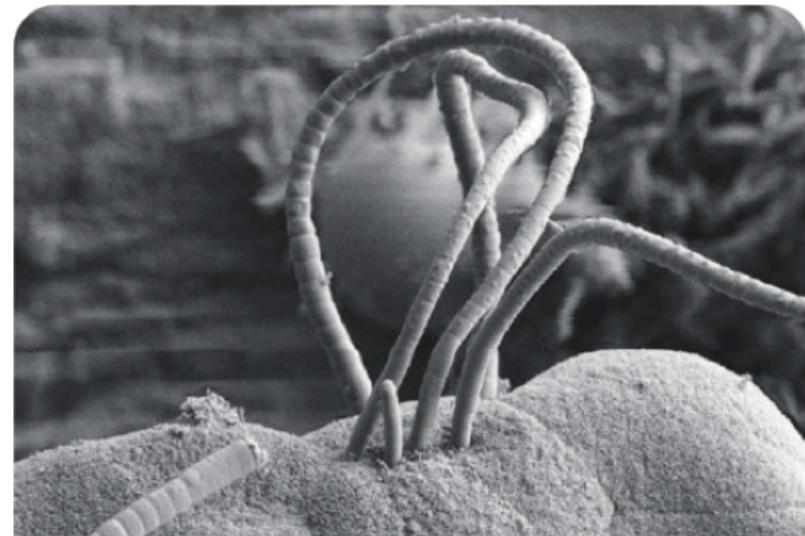
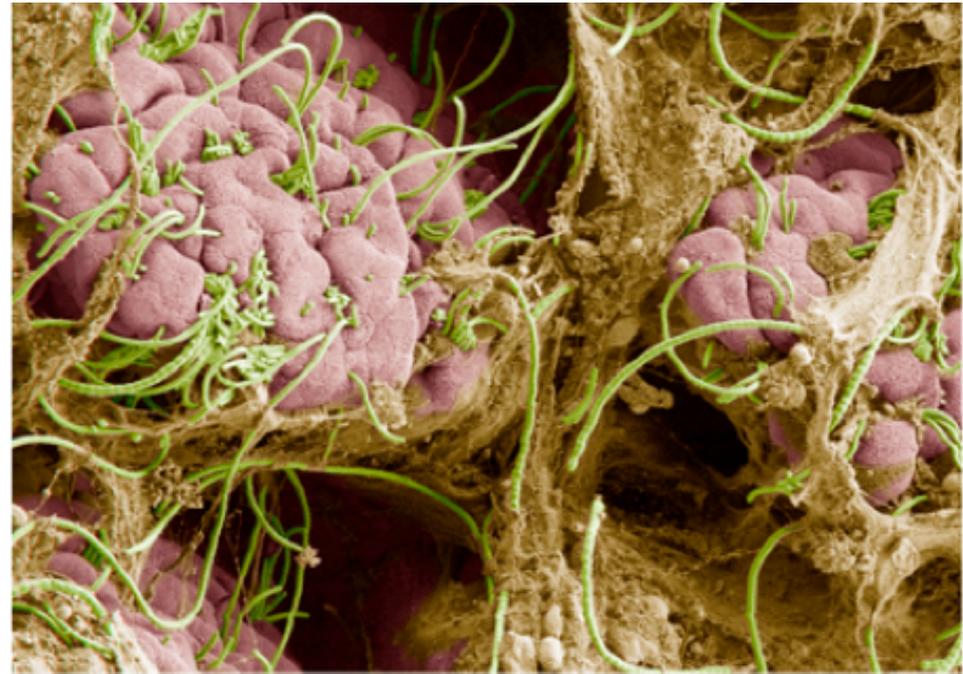
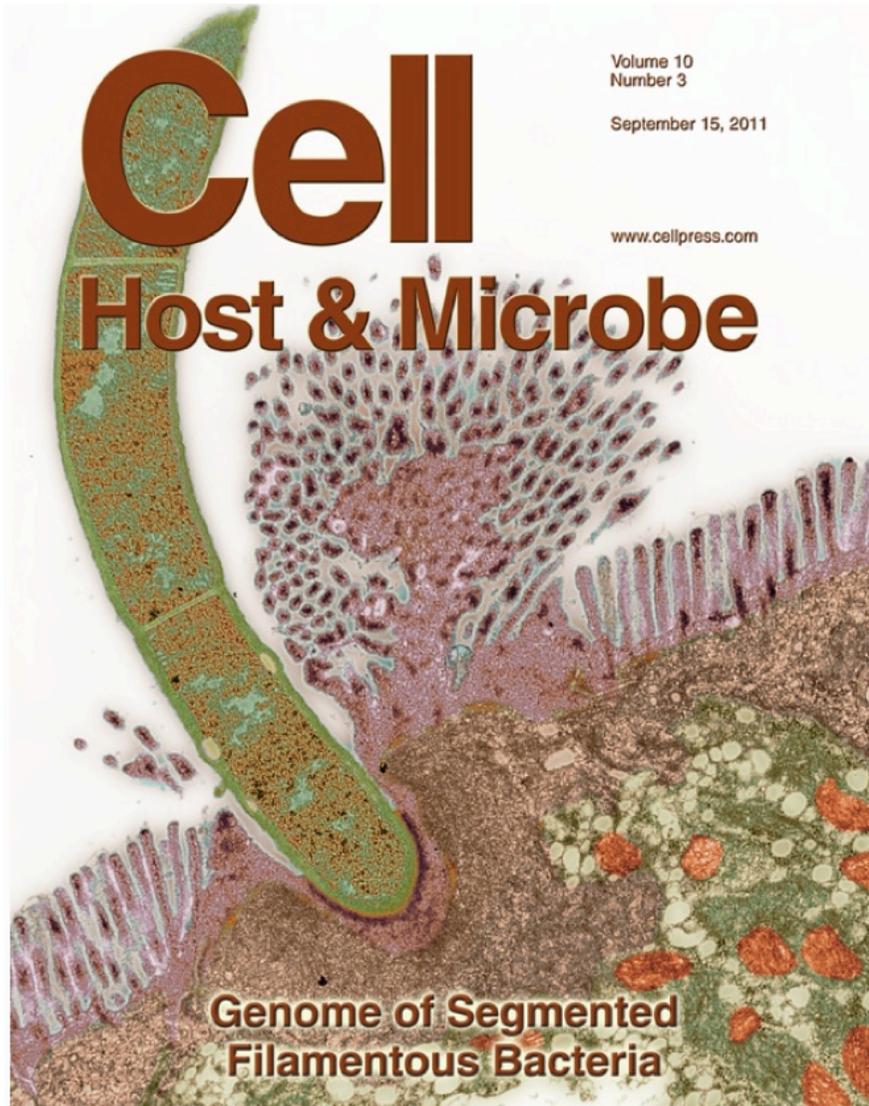
# Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis

Nature 2008

Djahida Bouskra<sup>1</sup>, Christophe Brézillon<sup>2</sup>, Marion Béard<sup>3</sup>, Catherine Werts<sup>4,6</sup>, Rosa Varona<sup>5</sup>, Ivo Gomperts Boneca<sup>4,6</sup> & Gérard Eberl<sup>1</sup>



# Bactéries Segmentées Filamenteuses (SFB)



## Segmented Filamentous Bacteria Are Potent Stimuli of a Physiologically Normal State of the Murine Gut Mucosal Immune System

GWEN L. TALHAM,<sup>1†</sup> HAN-QING JIANG,<sup>1</sup> NICOLAAS A. BOS,<sup>2</sup> AND JOHN J. CEBRA<sup>1\*</sup>

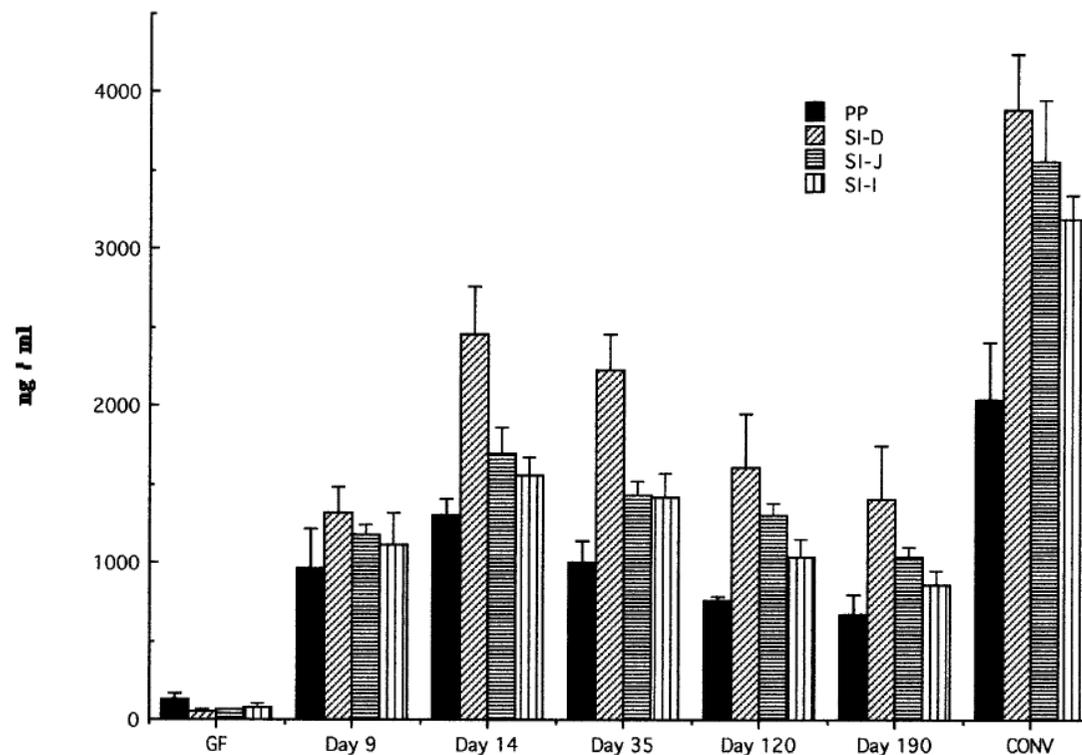
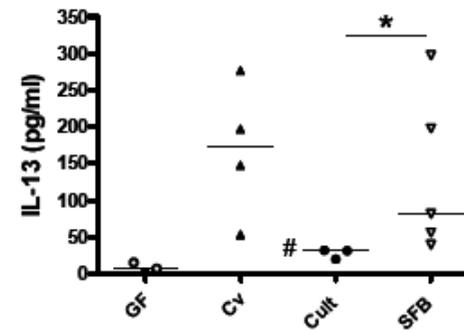
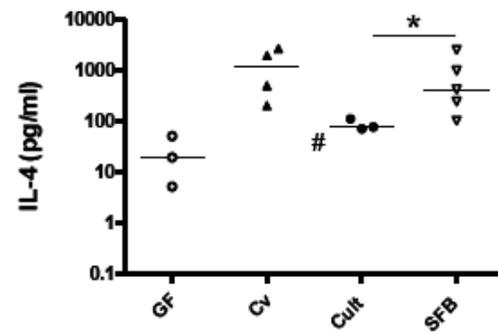
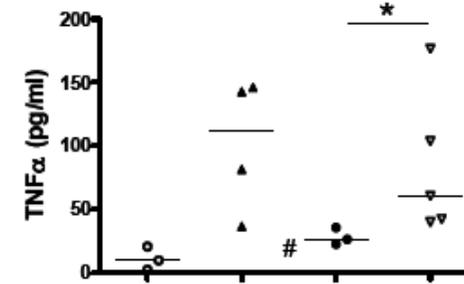
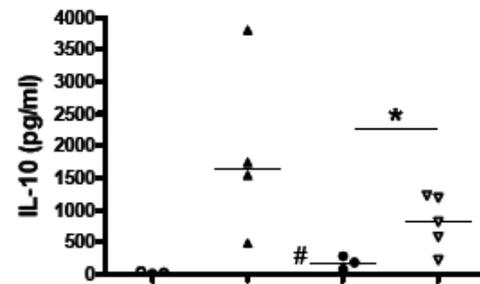
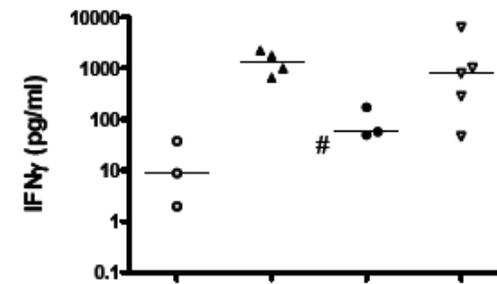
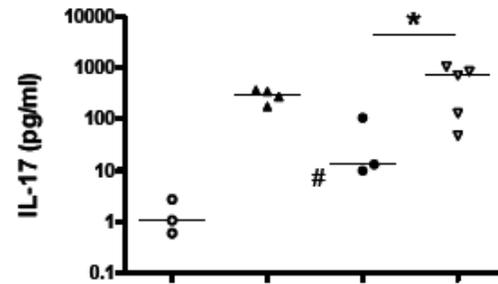
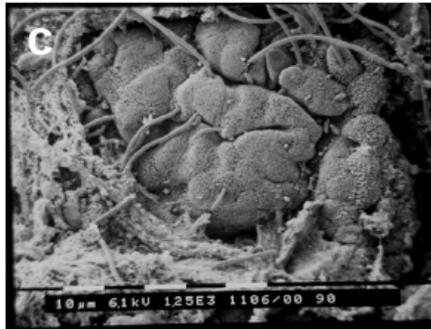
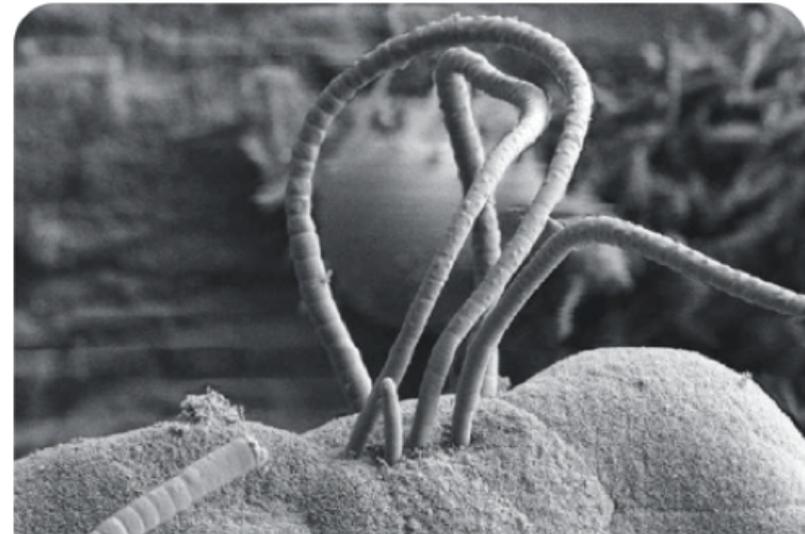
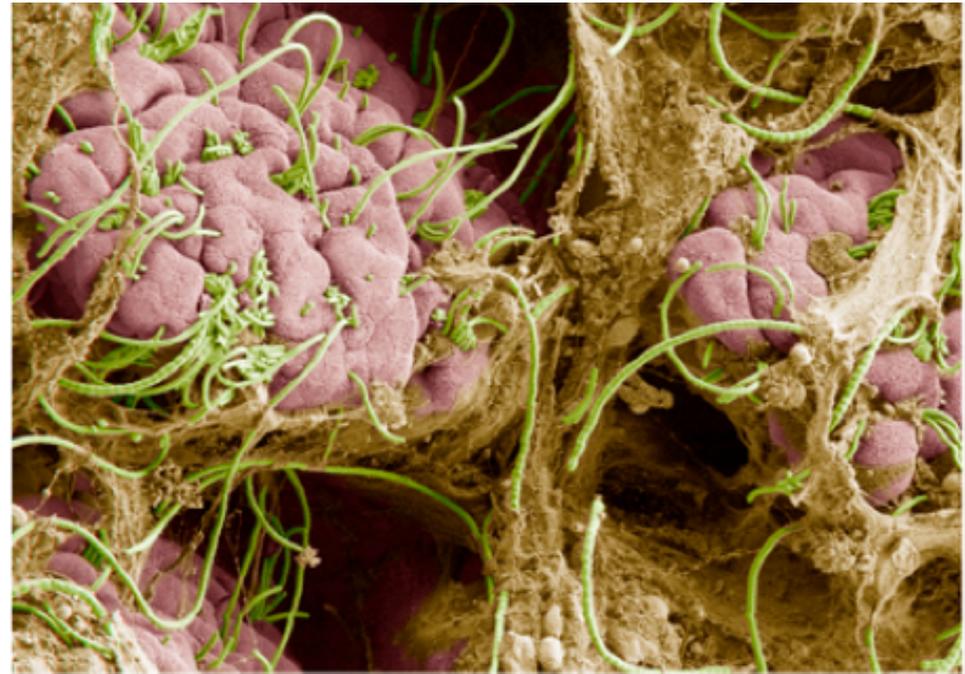
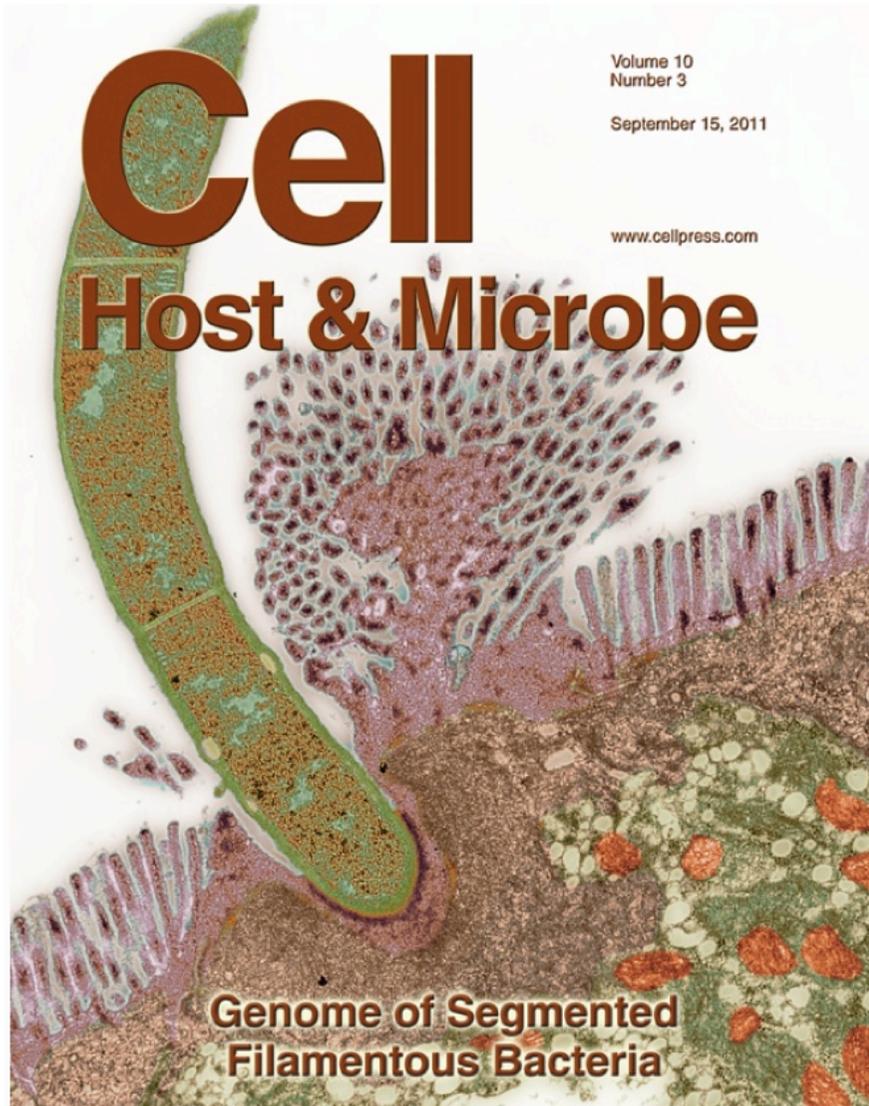


FIG. 2. Production of total IgA in supernatant of organ fragment cultures of PP and small intestine of GF, conventionally reared (CONV), and SFB-monoassociated C3H/HeN mice at various time points. RIA was used to detect IgA. Data are means  $\pm$  standard errors of the mean. SI, small intestine; D, duodenum; J, jejunum; I, ileum. The number of fragment cultures per tissue per time point ranged from 4 to 14.

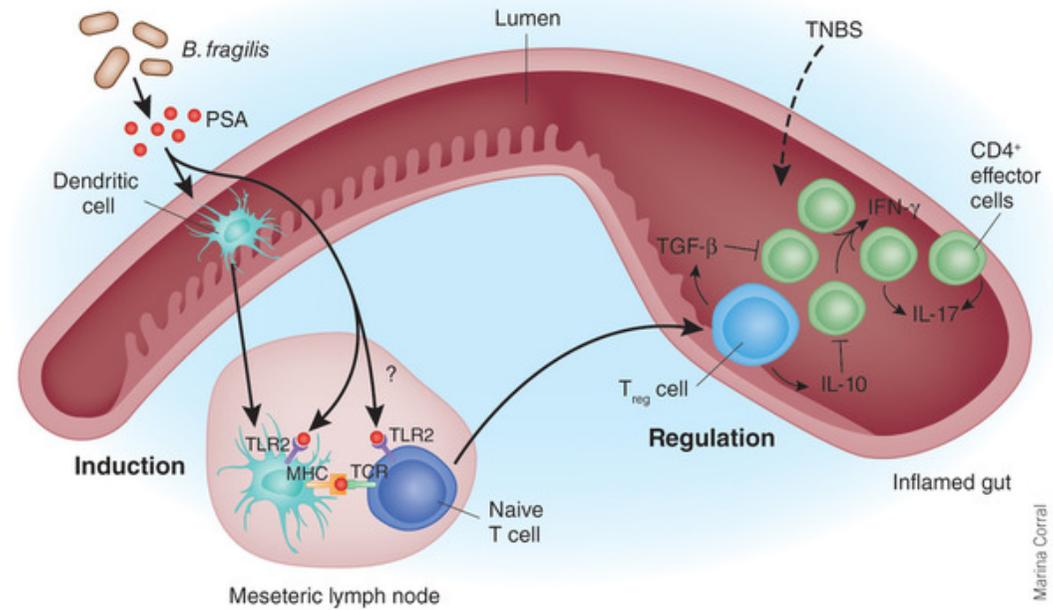
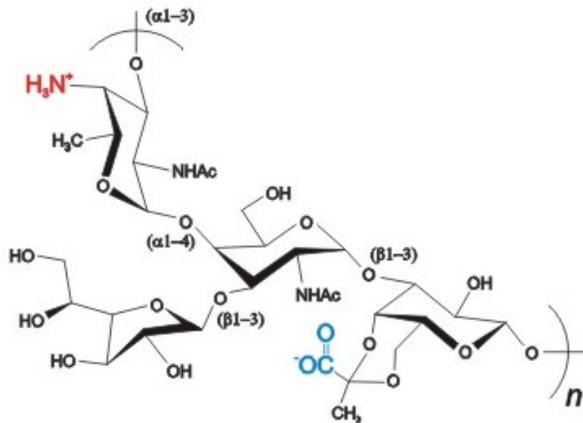
# Les SFB induisent fortement les réponses adaptatives



# Les SFB élèvent le niveau d'alerte du système immunitaire

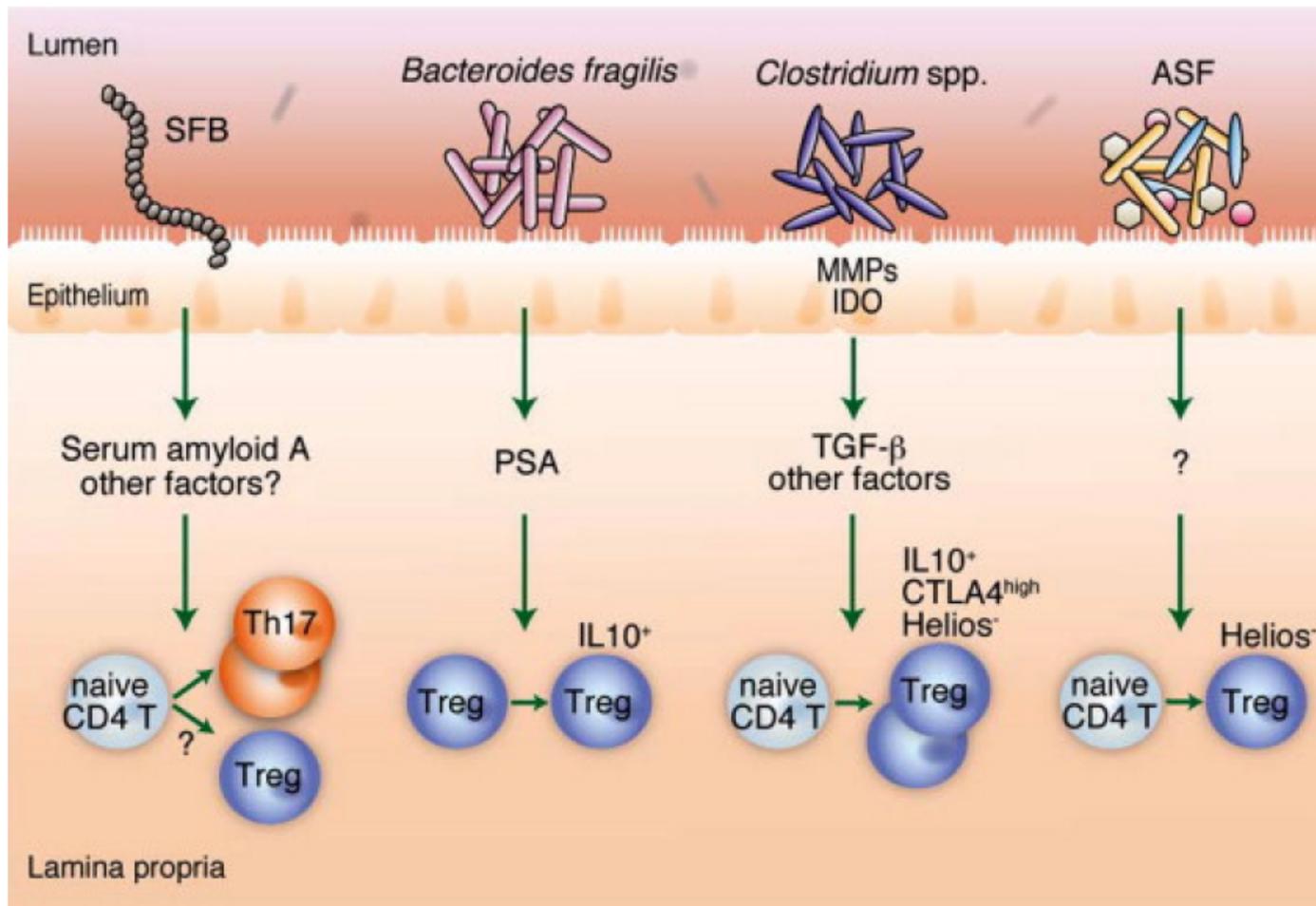


# Bacteroides fragilis et le PSA

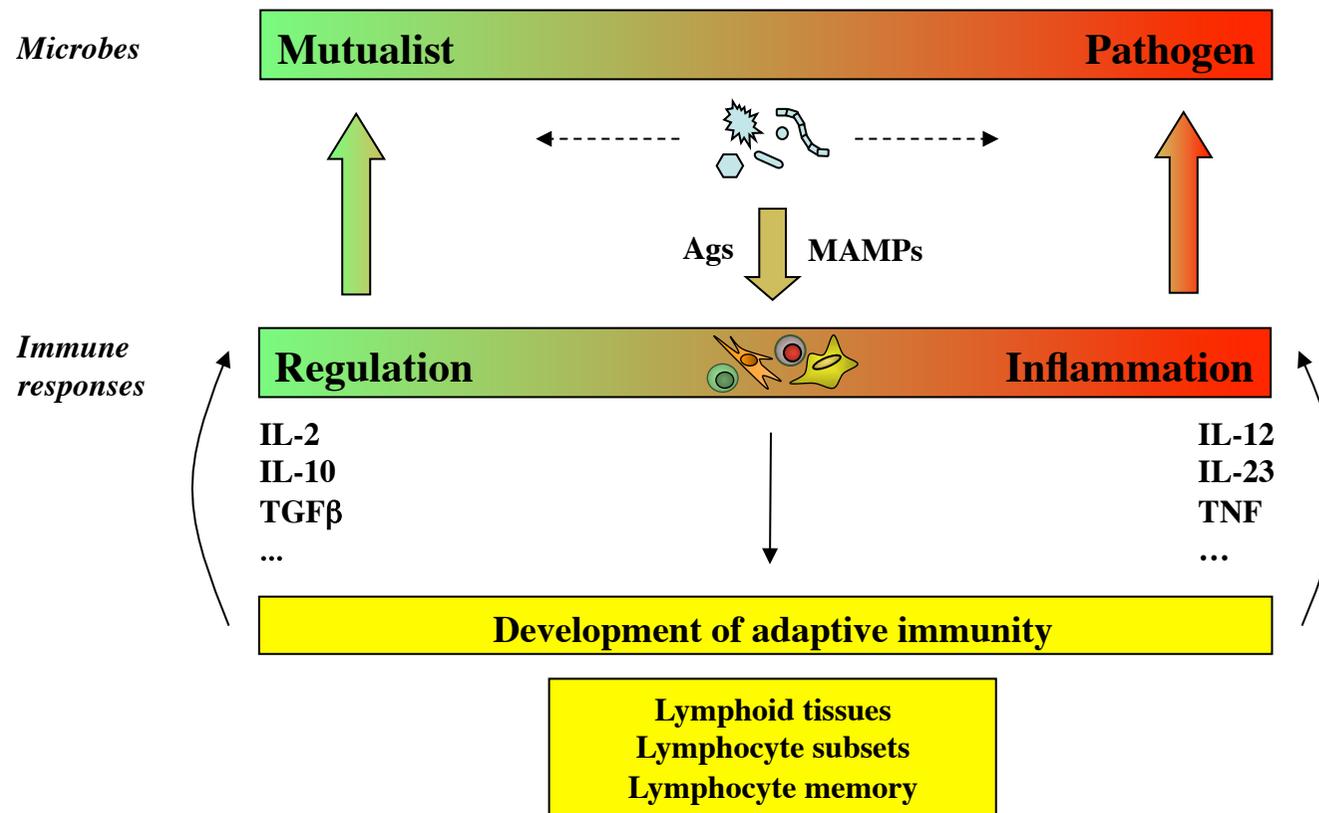


# Induction of Treg cells in the mouse colonic mucosa: A central mechanism to maintain host–microbiota homeostasis

Takeshi Tanoue, Kenya Honda 🌱 . 📧



# La construction du système immunitaire



# Redefining Chronic Viral Infection

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Cell 2009

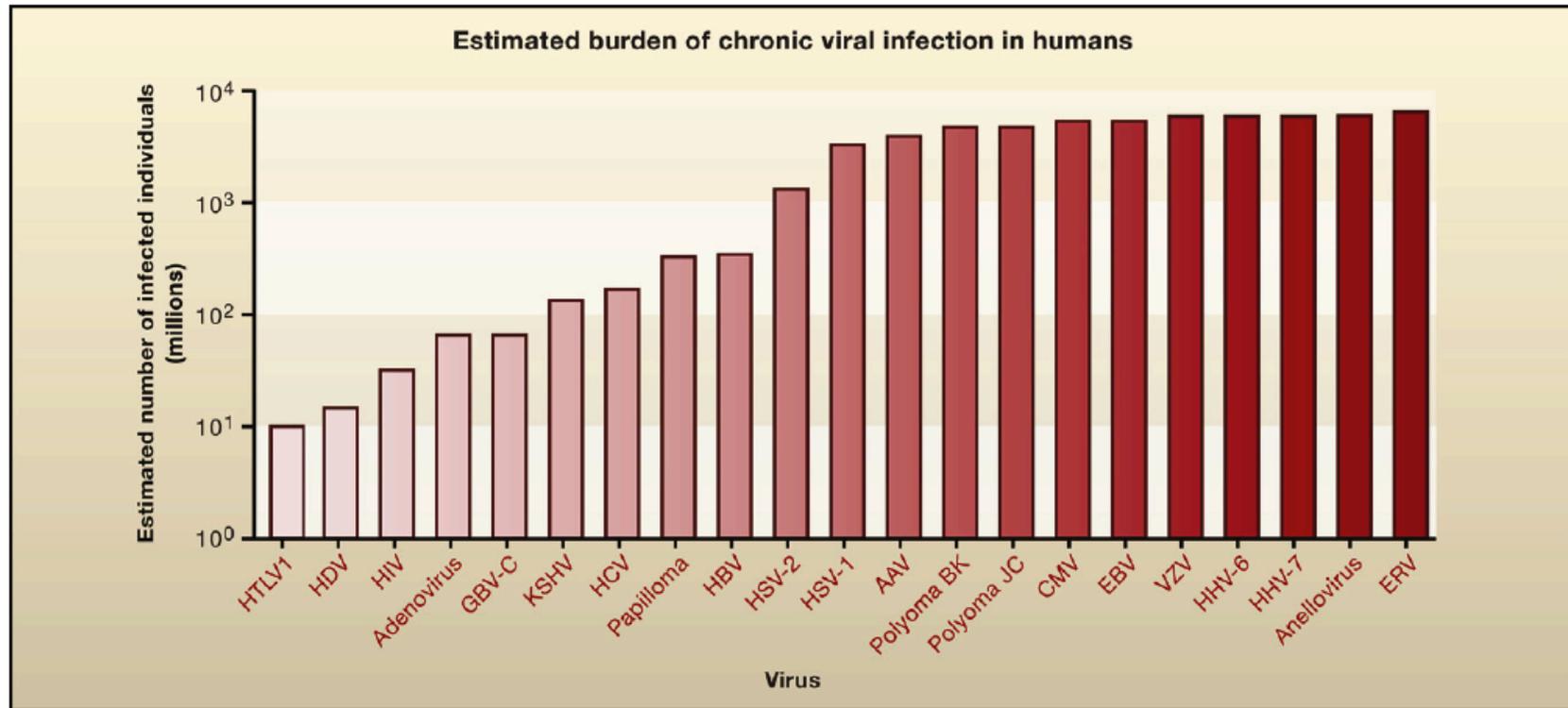
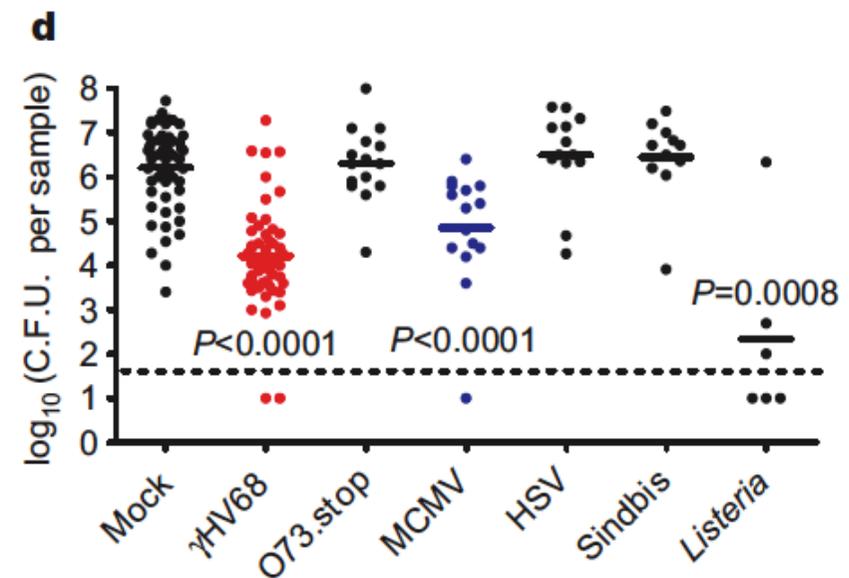
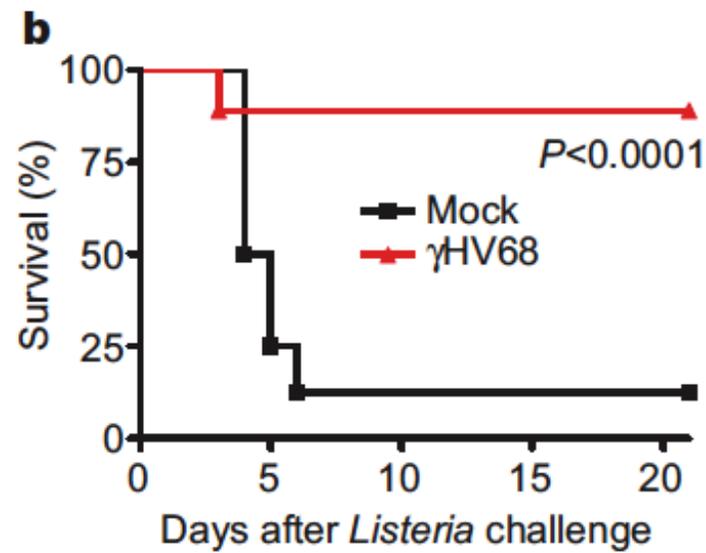


Figure 2. Chronic Viral Infections in Humans

# Herpesvirus latency confers symbiotic protection from bacterial infection

Erik S. Barton<sup>1†</sup>, Douglas W. White<sup>1,5</sup>, Jason S. Cathelyn<sup>2</sup>, Kelly A. Brett-McClellan<sup>1</sup>, Michael Engle<sup>3</sup>, Michael S. Diamond<sup>1,2,3</sup>, Virginia L. Miller<sup>2,4</sup> & Herbert W. Virgin IV<sup>1,2</sup>

Nature 2007



# Ilya Illitch Metchnikof (1845-1916) et les probiotiques

