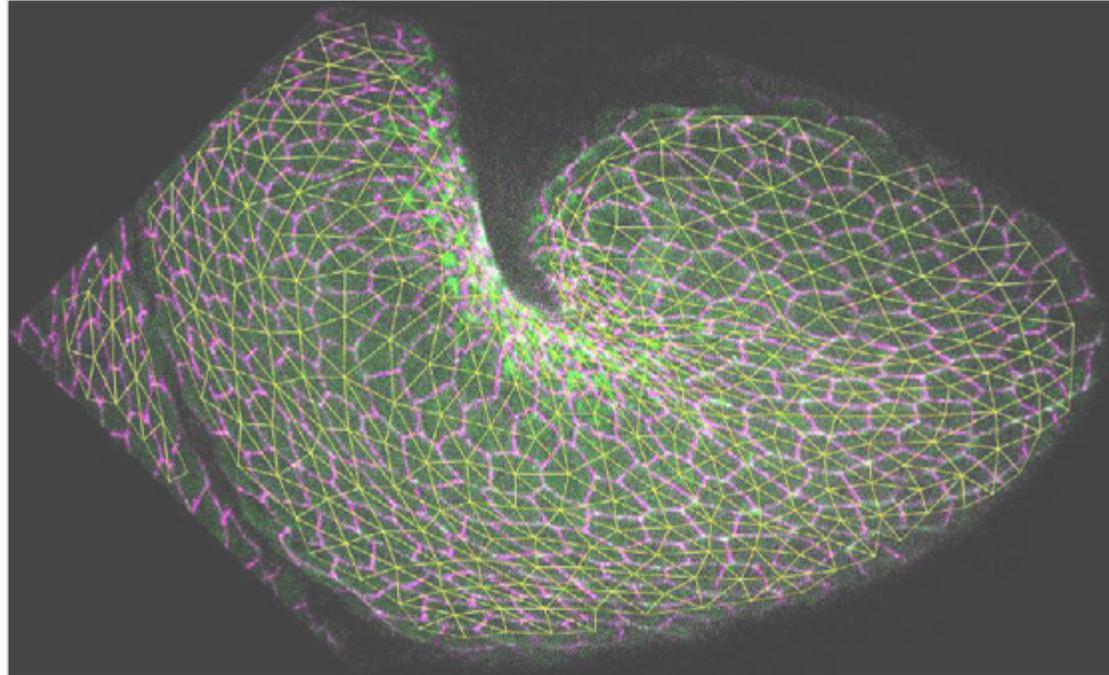


# Morphogenesis: space, time, information



Course 4: Folding, Invagination

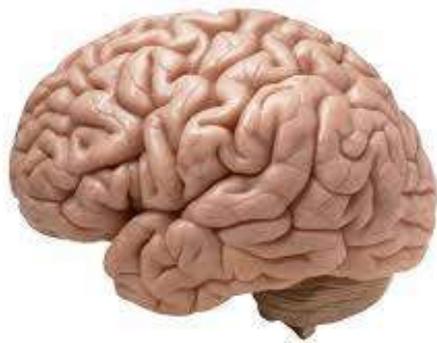
*Thomas Lecuit*  
*chaire: Dynamiques du vivant*



COLLÈGE  
DE FRANCE  
— 1530 —

# Tissue curvature: control and self-organisation

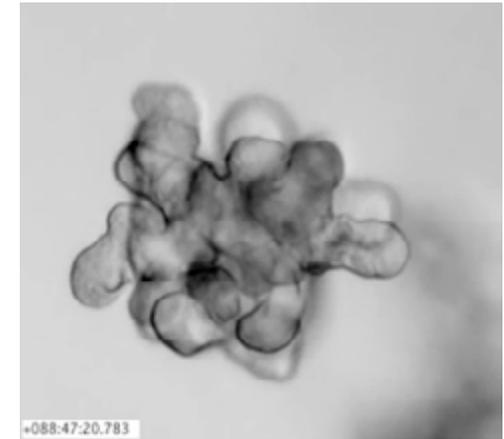
## I. Stress induced curvatures during tissue folding , looping, branching



Cortex convolution



Gut villi



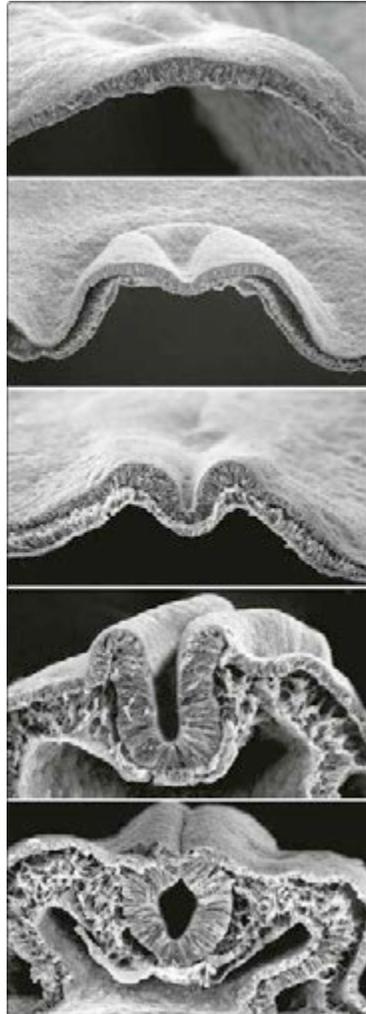
lung branching



Gut looping

# Tissue deformations: control and self-organisation

## 2. Tissue invagination

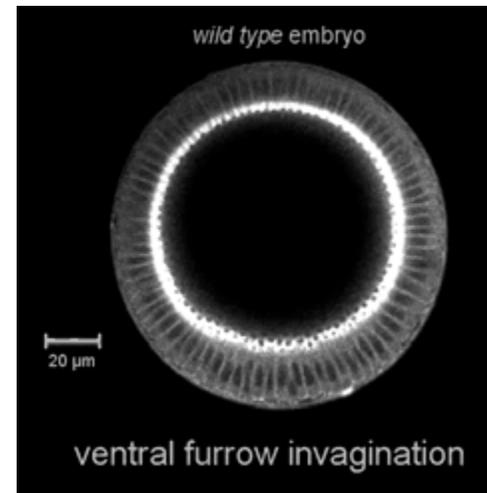


G. Schoenwolf U. Utah School of Medicine

Chick neural tube



Sea Urchin endoderm



*Drosophila* mesoderm

# Stress induced curvatures during tissue folding , looping etc

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## I. Self-Organised mechanical instabilities

### General Framework:

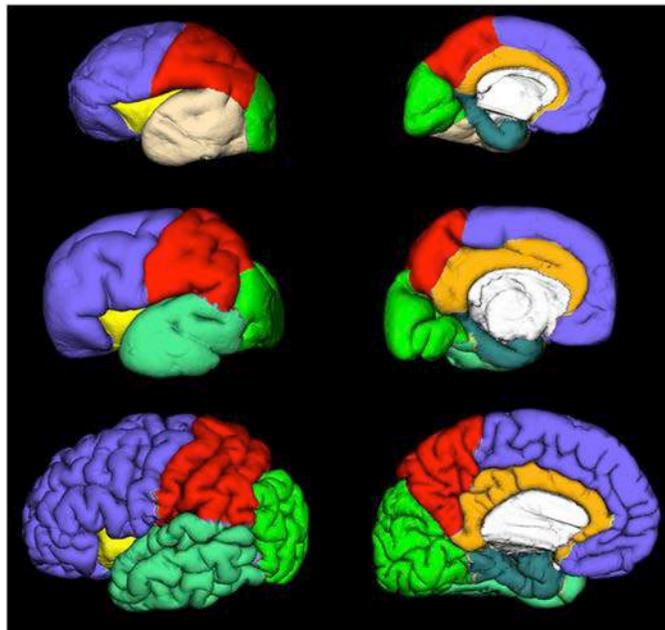
- Elastic material
- Internal stress: eg. growth
- External stress: boundary conditions
- Stress relaxation

Use of continuous/coarse grained models

# Growth induced mechanical instabilities: Cortex convolutions

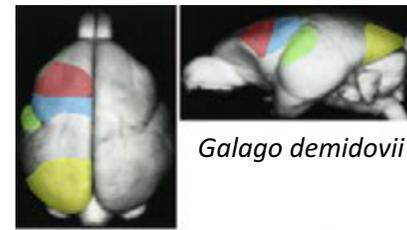
- Mechanics of Brain convolutions

- Development
- Evolution



*Homo sapiens*

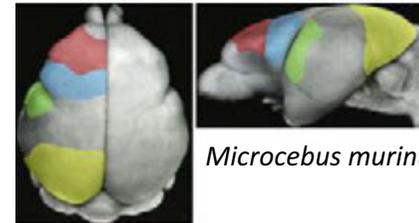
Vasung L, et al and Kostovic I (2016) *Front. Neuroanat.* 10:11. doi: 10.3389/fnana.2016.00011



*Galago demidovii*



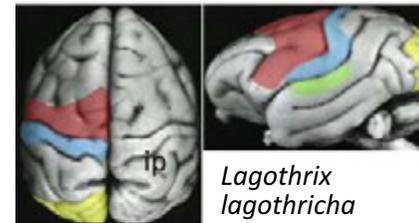
<https://fr.wikipedia.org/wiki/Galagoides>



*Microcebus murinus*



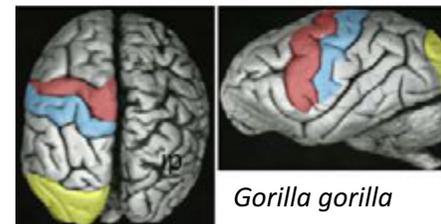
[https://en.wikipedia.org/wiki/Gray\\_mouse\\_lemur](https://en.wikipedia.org/wiki/Gray_mouse_lemur)



*Lagothrix lagotricha*



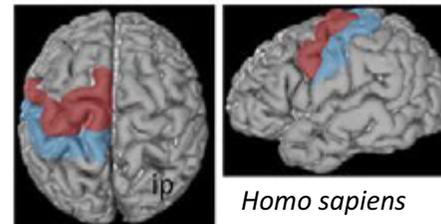
[https://fr.wikipedia.org/wiki/Lagothrix\\_lagotricha](https://fr.wikipedia.org/wiki/Lagothrix_lagotricha)



*Gorilla gorilla*



[https://fr.wikipedia.org/wiki/Gorilla\\_gorilla\\_gorilla](https://fr.wikipedia.org/wiki/Gorilla_gorilla_gorilla)



*Homo sapiens*



■ Motor cortex (primary motor and premotor cortex)  
■ Somatosensory cortex (BA 3, 1, 2)

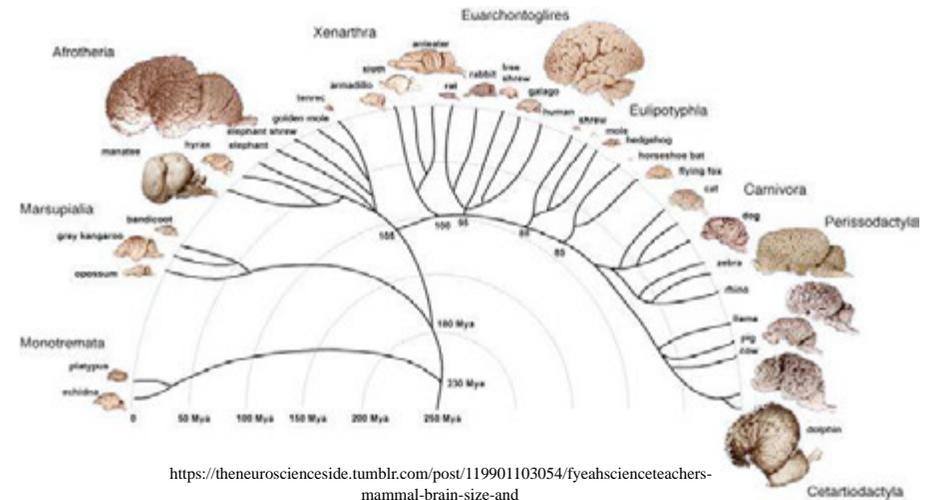
■ Auditory cortex (core and belt)  
■ Visual cortex (V1 and V2)



# Growth induced mechanical instabilities: Cortex convolutions

- Mechanics of Brain convolutions
- Cortical folding scaling laws

1. Folding index generally increases with brain mass.
2. But there are a number of outliers (cetacea, marsupials, humans)



3. Two different scaling laws for smooth and folded cortices

folded cortex

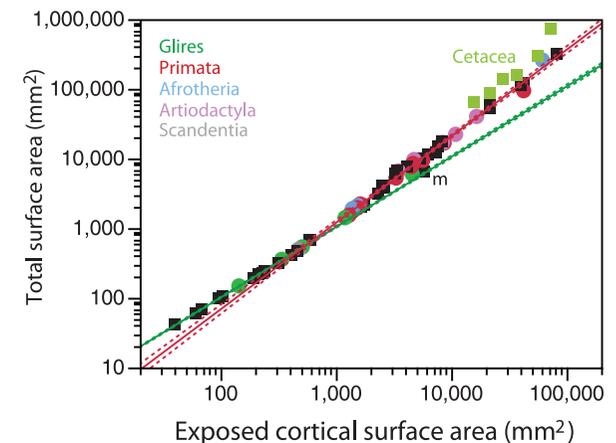
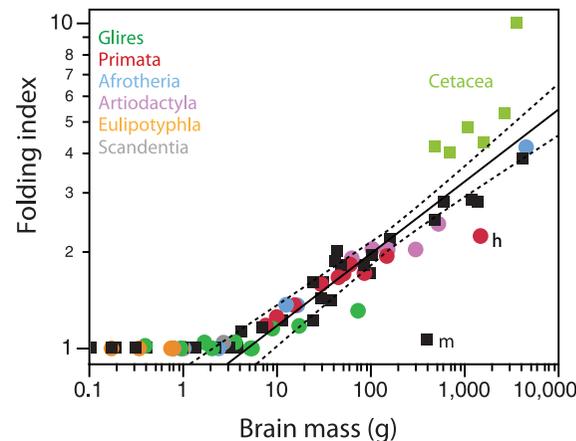
$$S_T = S_E^{1.24}$$

smooth cortex

$$S_T = S_E$$

Self-similar fractal configuration  
(fractal dimension  $d=2.48$ )

Similar to crumpled paper  
(fractal dimension  $d=2.5$ )



# Growth induced mechanical instabilities: Cortex convolutions

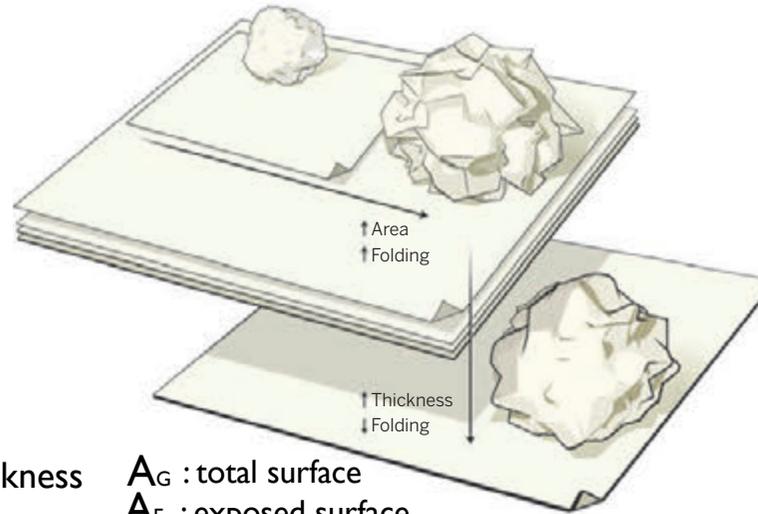
- A mechanical model of cortex folding: minimisation of stresses

1. Folding of a sheet of paper: Self-avoiding (non-intersecting) surface that minimises its free energy

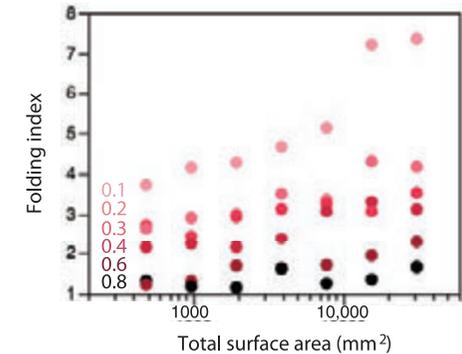
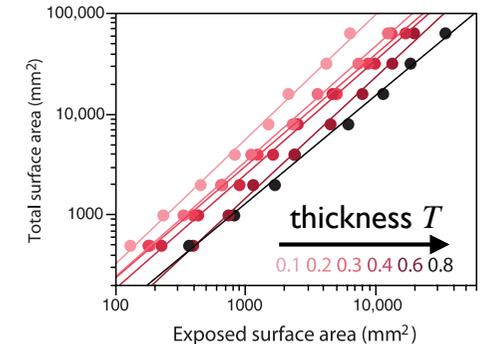
(minimisation of internal and external stresses)

$$T^{1/2} A_G = k A_E^{1.25}$$

(1.25 exponent so that  $k$  is dimensionless)



$A_G$  : total surface  
 $A_E$  : exposed surface  
 - a thinner sheet folds more



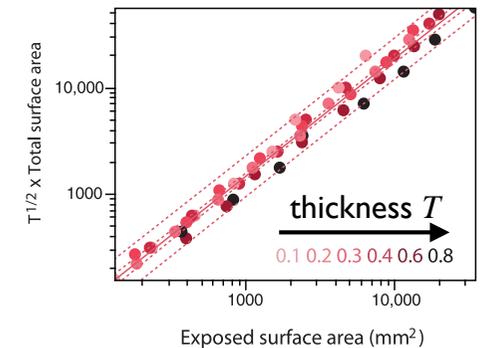
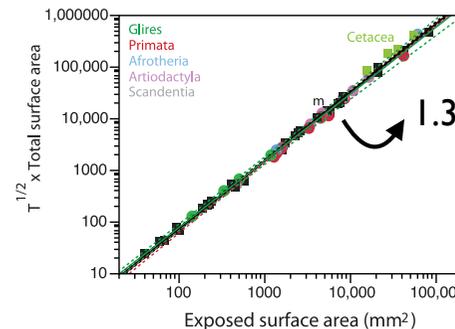
2. All cortices (gyrencephalic and lissencephalic) follow a unique power law where folding scales universally with surface area and thickness

3. The exponent is nearly the same as for folded paper

$$T^{1/2} A_G = A_E^{1.3}$$

lissencephaly if  $T = k^2 A_G^{1/2}$  ( $A_G = A_E$ )

gyrencephaly if  $T < k^2 A_G^{1/2}$  ( $A_G > A_E$ )



# Growth induced mechanical instabilities: Cortex convolutions

---

- A mechanical model of cortex folding: minimisation of stresses
  1. Universality of scaling law across folded and unfolded cortices calls for intrinsic property of cortex
  2. Follows minimisation of internal and external stresses
  3. Depends on relative rate of lateral expansion of total surface (progenitor expansion) relative to its thickness (radial neurogenesis, cell migration)
  4. Evolution of brain shape reflects evolution of mechanisms regulating  $T$  relative to  $A_G$
  5. What is the origin of internal stresses that emerge during development of brain?

# Growth induced mechanical instabilities: Cortex convolutions

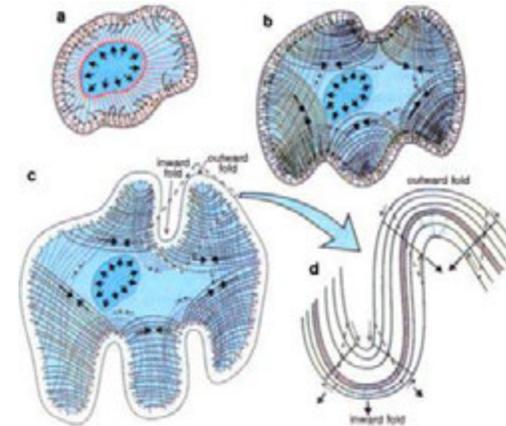
- Models of brain convolutions: internal stresses

1. Tensile forces of axons (white matter) onto cortex (grey matter)

**A tension-based theory of morphogenesis and compact wiring in the central nervous system**

David C. Van Essen  
Department of Neurobiology, University of California, San Diego, La Jolla, California 92037

DC Van Essen *Nature* (1997) 385:313

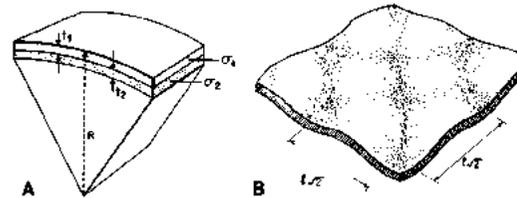


2. Differential growth rate in grey matter compared to white matter: compressive stress

**Mechanical Model of Brain Convolutional Development**

Pathologic and experimental data support a model based on differential growth within the cerebral cortex.

David P. Richman, R. Markus, Steven John W. Hutchinson, Vincenzo C. Lippman, Jr.



DP. Richman et al. and V. Caviness *Science* (1975) 189:18

3. Chemical Turing pattern pre-pattern the cortex and induce growth heterogeneities

Lefèvre & Mangin *Plos Comp. Biol.* 2010



# Growth induced mechanical instabilities: Cortex convolutions

- Models of brain convolutions: internal stresses

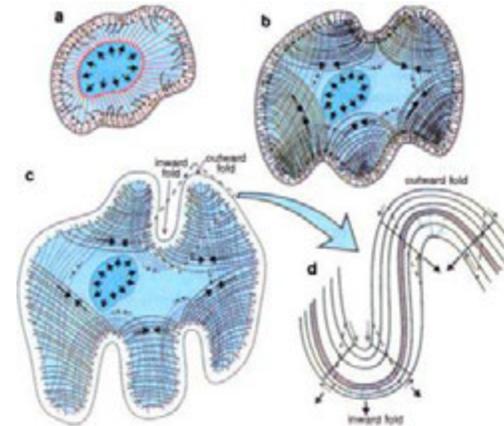
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Experiments mapping distribution of tensile stress is not consistent  
 Xu et al, L. Taber *J. Biomech. Eng.* (2010) 132: 0710131



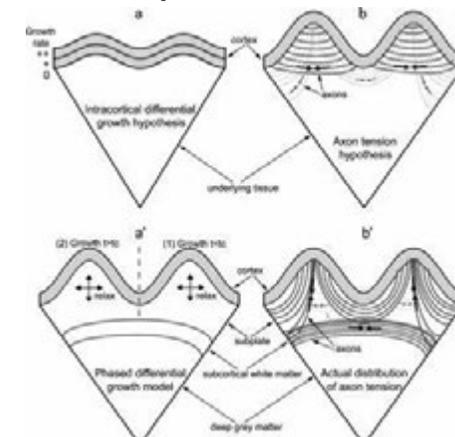
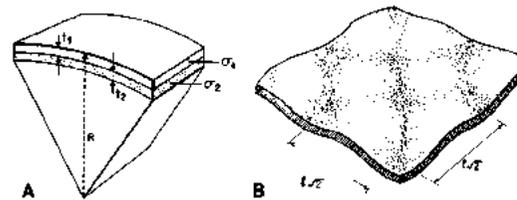
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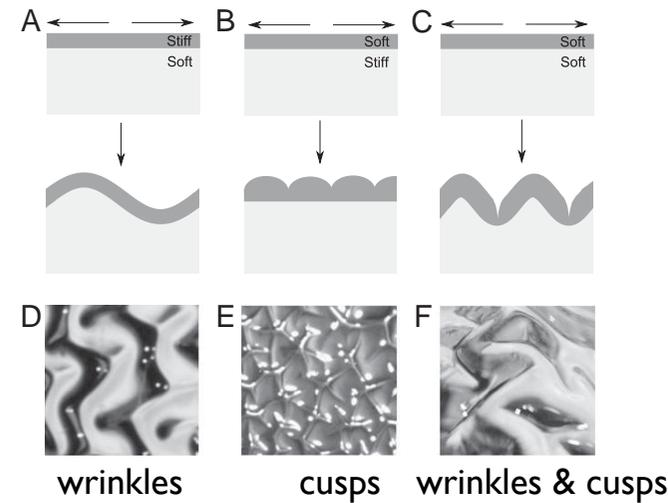
- Chemical Turing pattern pre-pattern the cortex and induce growth heterogeneities

no evidence: Lefèvre & Mangin *Plos Comp. Biol.* 2010

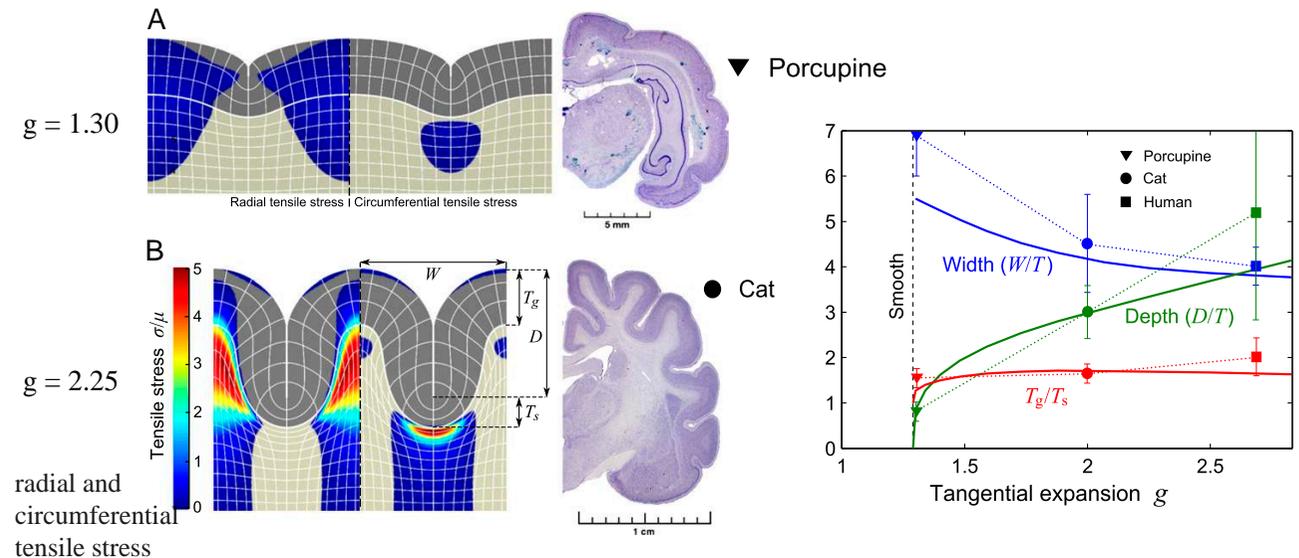


# Growth induced mechanical instabilities: Cortex convolutions

- Cortical folding (gyrification) arises as a mechanical instability due to heterogeneous growth of a soft tissue

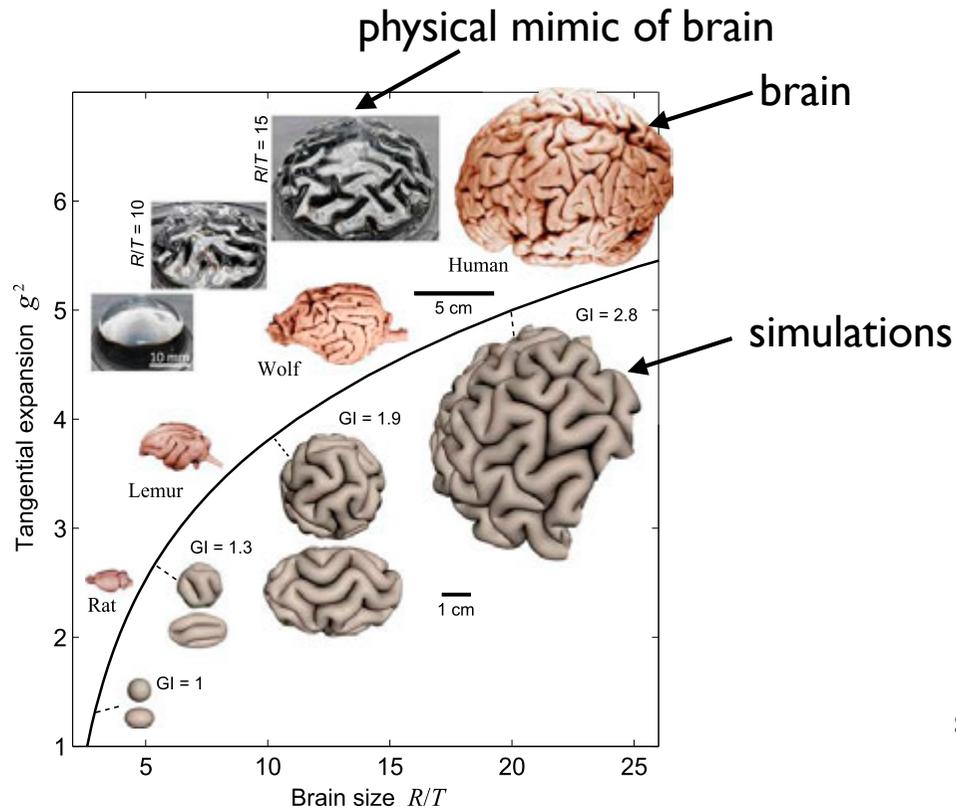


- Brain best modelled as soft tissue (both in white and grey matter)
- Elastic solid
- Differential growth of white ( $g=1$ ) and grey matter ( $g=1+x$ )

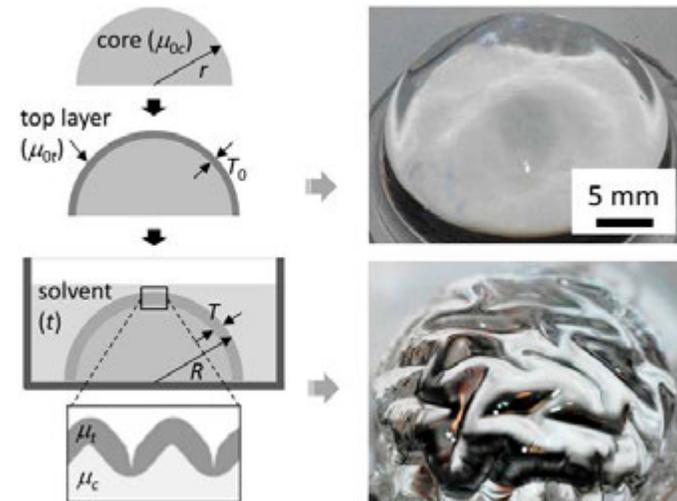


# Growth induced mechanical instabilities: Cortex convolutions

- Cortical folding (gyrification) arises as a mechanical instability due to heterogeneous growth of a soft tissue



physical model of brain like instability



swelling of outer layer (growth)

GI: gyration index (total area/exposed area)



# Growth induced mechanical instabilities: Cortex convolutions

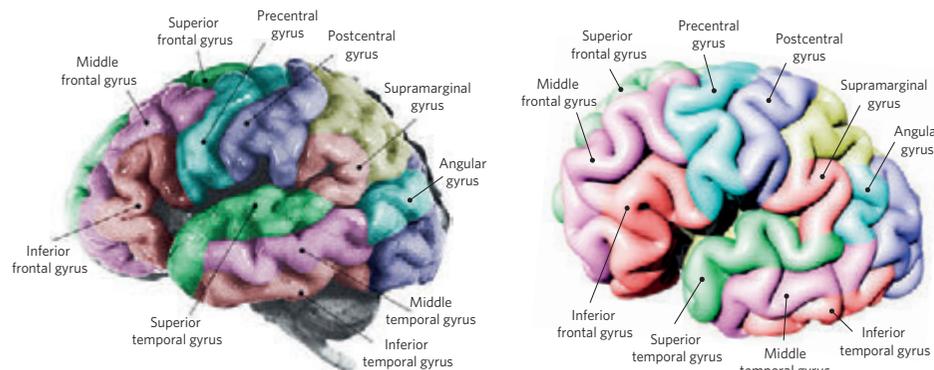
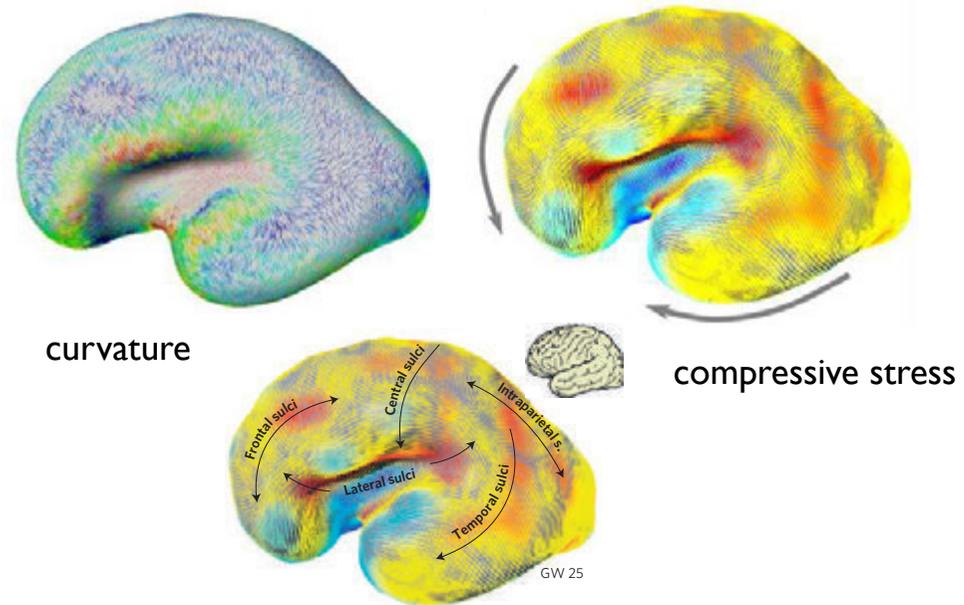
- 3D model of brain folding: impact of initial geometry

Initial geometry characterised by curvature map

High (Low) curvature associated with Low (High) compressive stress

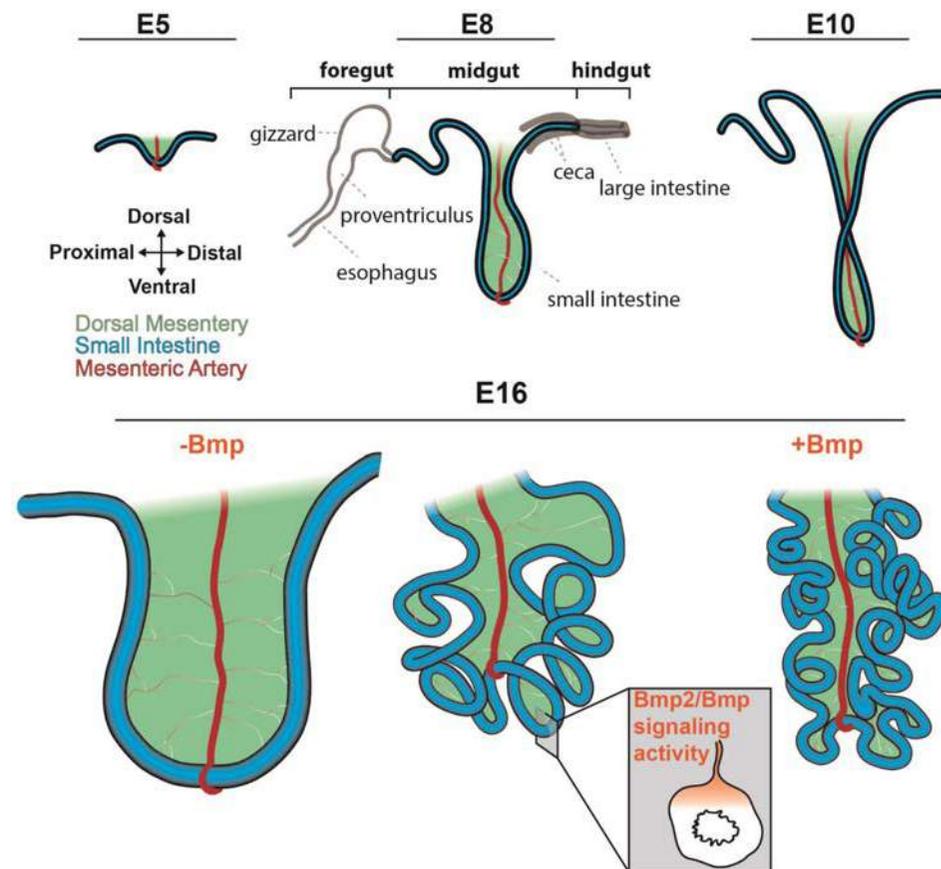
Main gyri result from compressive stress

Other smaller gyri are less reproducible in terms of their position.



# Growth induced mechanical instabilities: Gut looping

- Gut morphogenesis involves looping and coiling

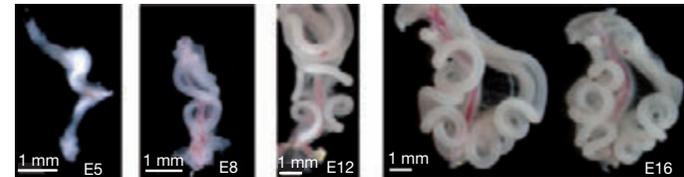


T. Huycke and C. Tabin. *Int. J. Dev. Biol.* 62: 109-119 (2018)

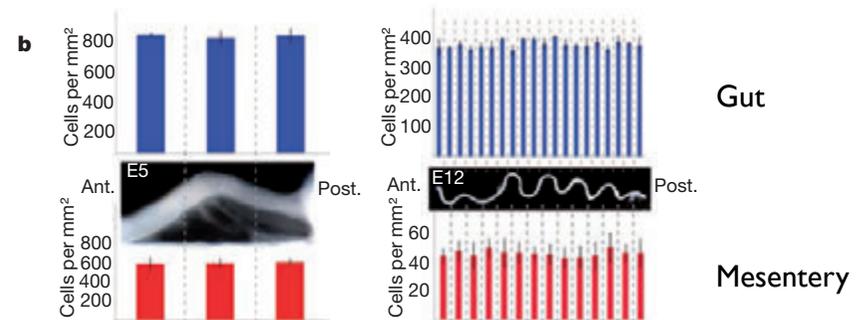
<https://doi.org/10.1387/ijdb.170325ct>

# Growth induced mechanical instabilities: Gut looping

- Gut looping arises as a mechanical instability due to differential tissue growth

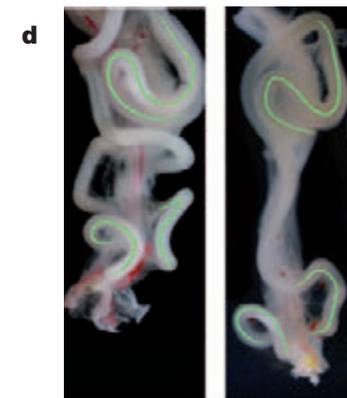


- Formation of stereotypical looping patterns
- The gut shows uniform proliferation
- Mechanical dissociation of mesentery and gut leads to gut relaxation by decoiling and mesentery relaxation: gut is normally compressed and mesentery is stretched.
- So the gut and mesentery are elastic, isotropic tissues.
- Hypothesis: Differential uniform growth underlies differential strain.



Mechanical dissociation of mesentery

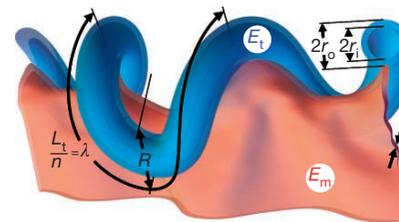
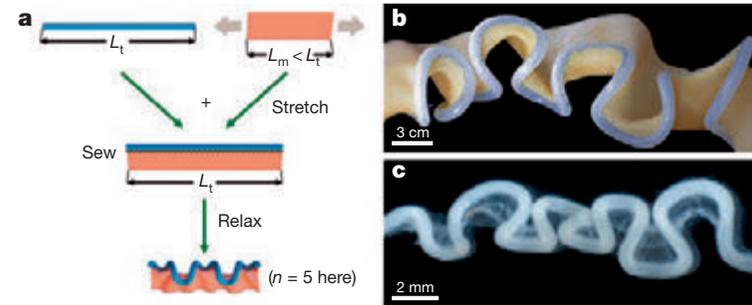
Mesentery removed *in vitro*



With mesentery Partial mesentery removal *in ovo*

# Growth induced mechanical instabilities: Gut looping

- A physical simulacrum of the gut and mesentery is similar qualitatively to gut looping in vivo
- An elastic theory predicts tissue geometry given elastic moduli of tube (gut) and sheet (mesentery), strain and force balance.
- When strain  $\varepsilon$  (due to differential growth) is above critical value  $\varepsilon_*$ , the gut buckles and loops.
- Geometry/Elasticity relations:



onset of looping:

$$\lambda \propto \left( \frac{E_t I_t}{E_m h} \right)^{1/3}$$

with  $\varepsilon_* \propto A/\lambda$

A: amplitude  
 $\lambda$ : length

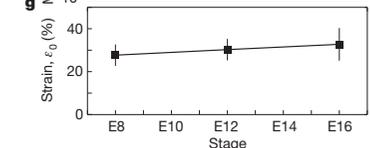
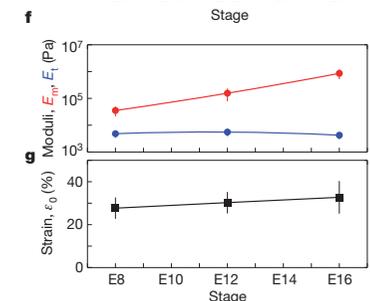
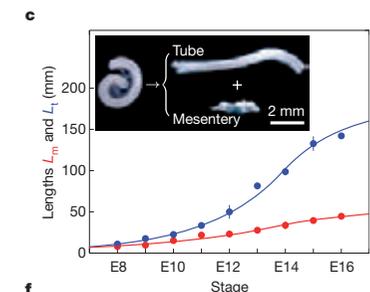
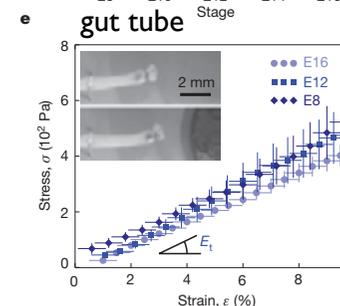
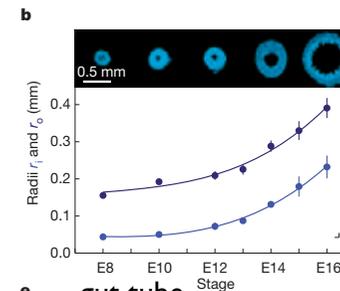
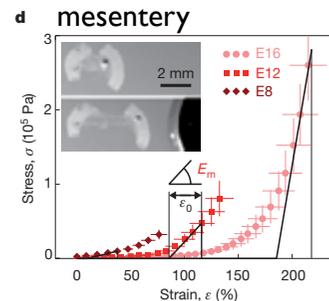
complete looping:

$$R \propto \left( \frac{E_t I_t}{E_m h \varepsilon_0^2} \right)^{1/3}$$

$\varepsilon = \varepsilon_0 \gg \varepsilon_*$

$E_t, E_m$ : Young's moduli  
I: quadratic moment

- Geometric measurements
- Mechanical measurements (stress/strain relations) of mesentery and gut (using magnetic force on steel ball)

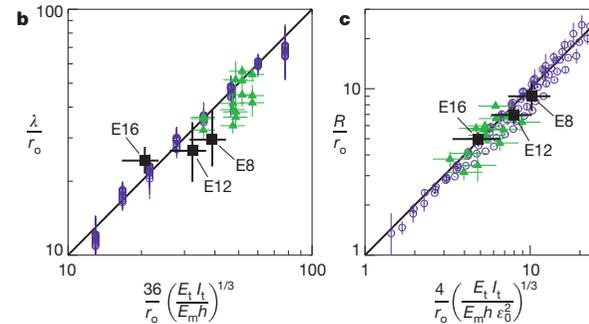


# Growth induced mechanical instabilities: Gut looping

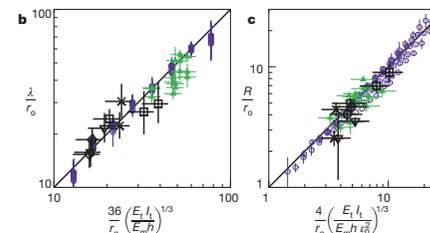
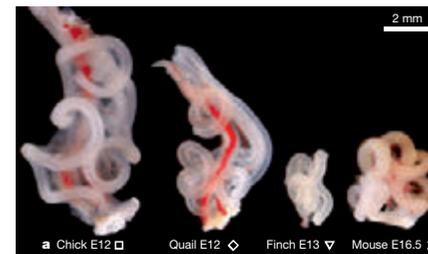
- No free parameter
- The elastic model predicts relationship between geometry and stiffness parameters.

$$\lambda \propto \left( \frac{E_t I_t}{E_m h} \right)^{1/3} \quad R \propto \left( \frac{E_t I_t}{E_m h \varepsilon_0^2} \right)^{1/3}$$

- The same elastic model explains the gut looping in different bird species and the mouse.
- Tuning parameters (from evolutionary perspective) are relative stiffness of tube and mesentery, differential growth rate (strain).



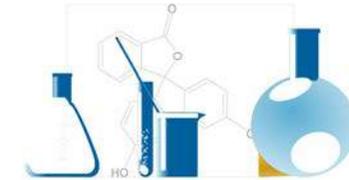
Chick  
Rubber model  
Simulations



# Growth induced mechanical instabilities: Gut vilification

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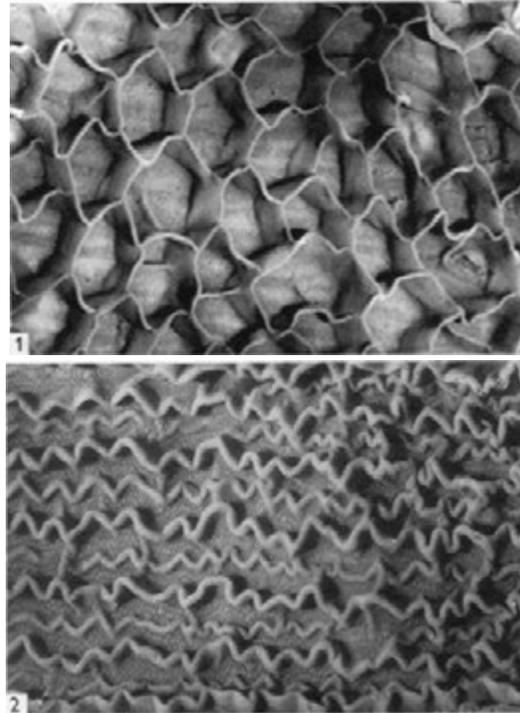
- Self-organised mechanics: chick gut
- Self-organised biochemistry: Turing field in mouse gut



# Growth induced mechanical instabilities: Gut vilification



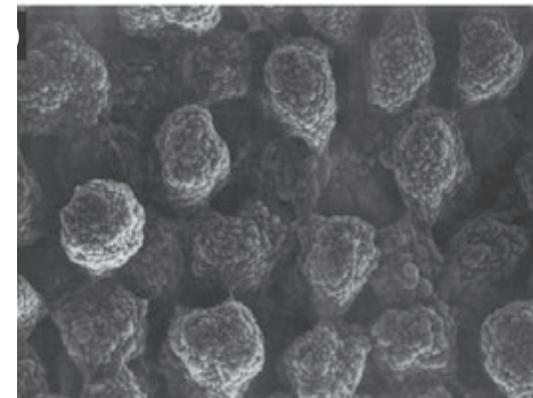
snake



snake: gut



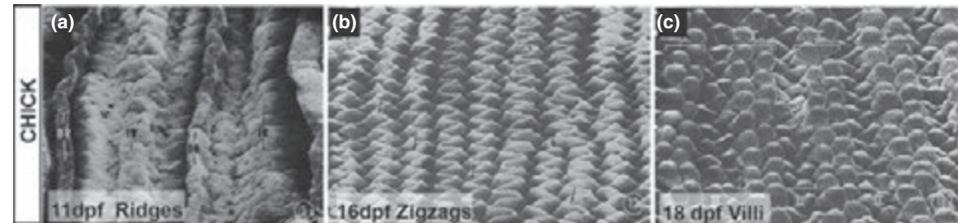
chick: duodenum



rat: gut

# Growth induced mechanical instabilities: Gut vilification

- Gradual development of folding patterns in the gut of birds

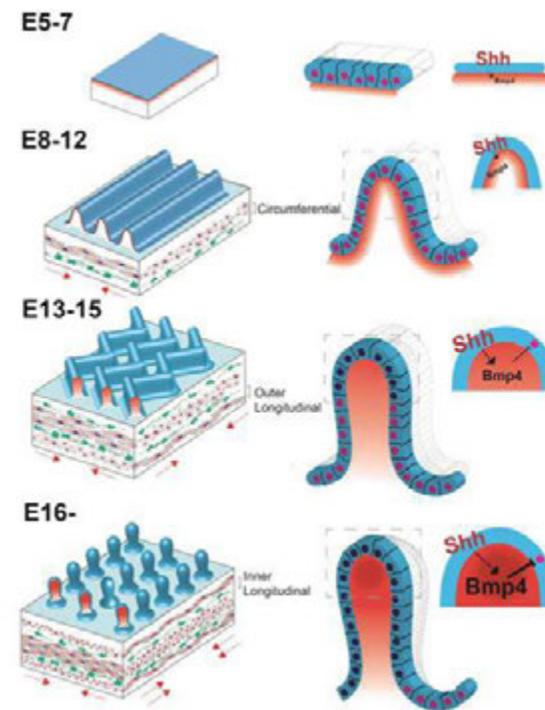


Epithelium and mesenchyme form annealed surfaces

Formation of ridges in epithelium and mesenchyme parallel formation of circumferential smooth muscles

Formation of Zigzags parallels formation of longitudinal muscles

Formation of Villi

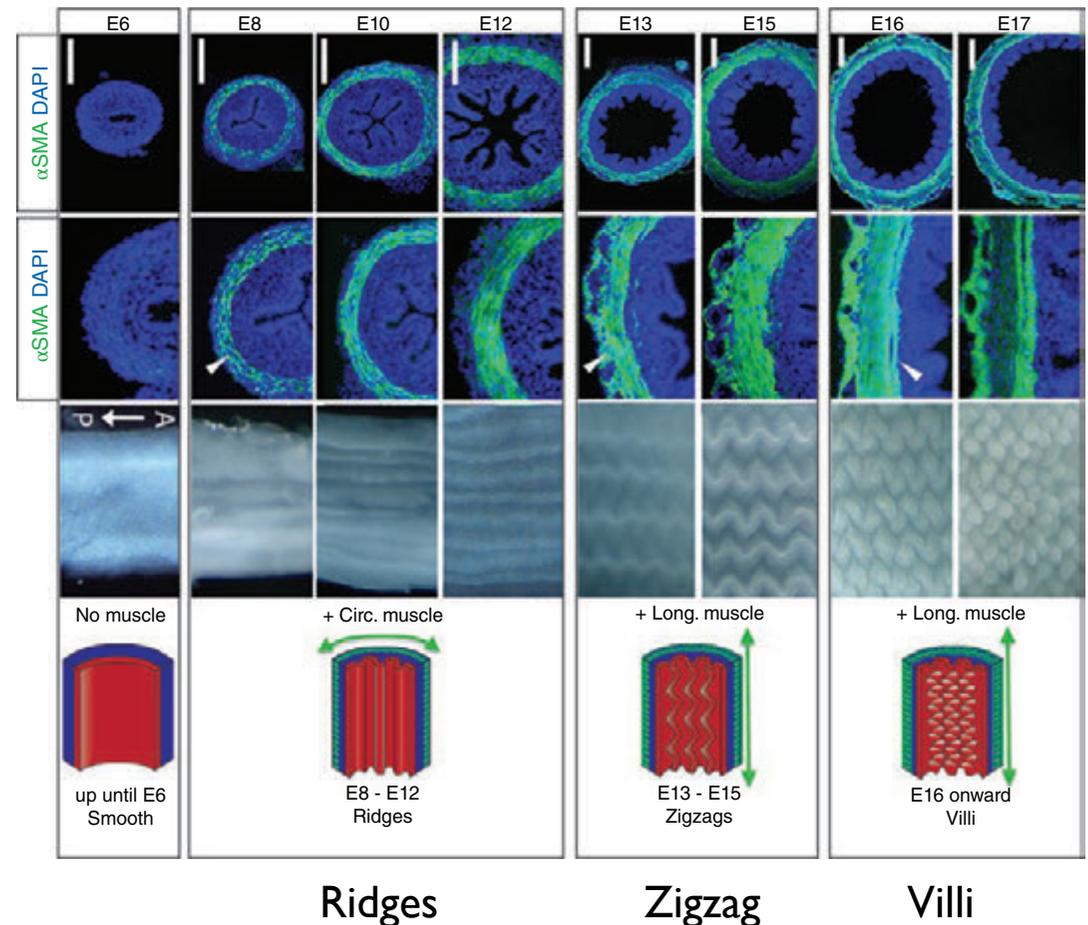


K Walton et al., C. Tabin and D. Gumucio *WIREs Dev Biol.* 2018;e317.

# Growth induced mechanical instabilities: Gut vilification

- Gradual development of folding patterns in the gut

- Epithelium and mesenchyme form annealed surfaces ensheathed by sequential layers of smooth muscles
- Formation of **Ridges** in epithelium and mesenchyme parallel formation of circumferential smooth muscles
- Formation of **Zigzags** parallels formation of longitudinal muscles
- Formation of **Villi** formation of longitudinal muscles



# Growth induced mechanical instabilities: Gut vilification

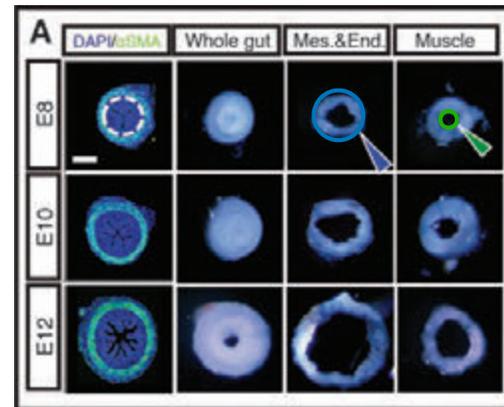
- Tissue ridges arise from mechanical instability caused by differential growth and constraints

- Mesenchyme and Epithelium grow more than surrounding smooth muscles and are consequently constrained and compressed.

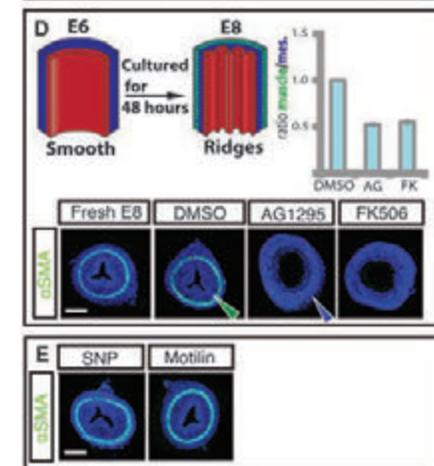
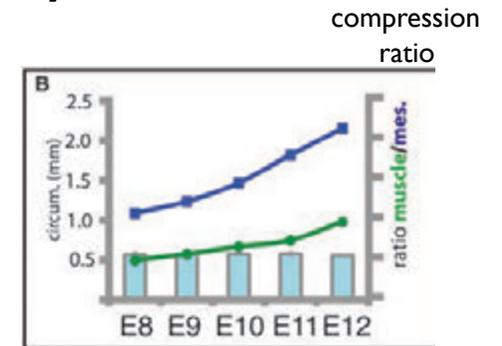
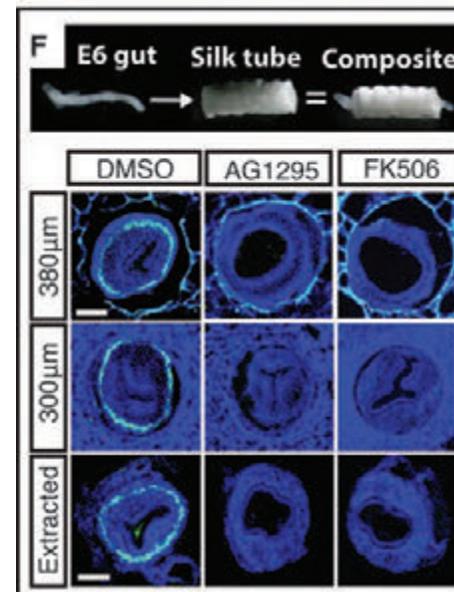
- Removal of smooth muscles leads to elastic unfolding of epithelium and mesenchyme

- Addition of artificial mechanical constraint rescues the need for surrounding smooth muscles

- **Role of circumferential constraints**



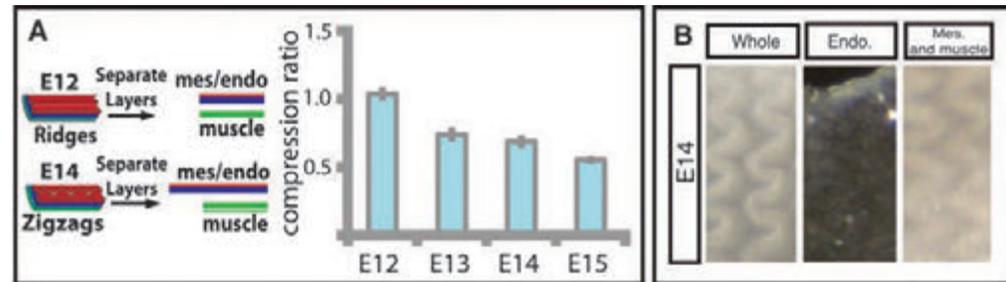
Mechanical removal of constraint



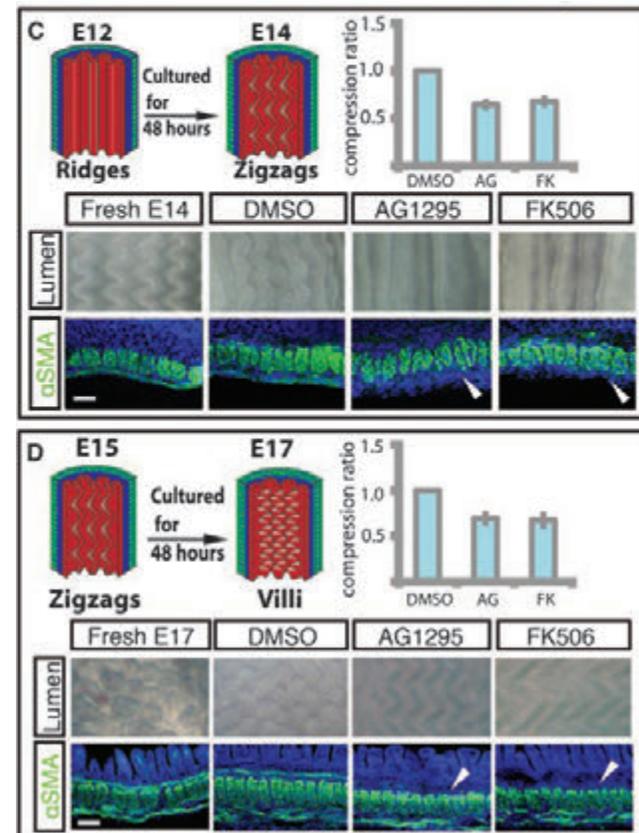
Pharmacological removal of constraint (inhibition of smooth muscle differentiation)

# Growth induced mechanical instabilities: Gut vilification

- Tissue zigzags and villi arise from mechanical instability caused by differential growth and constraints



- Removal of smooth muscles leads to elastic lengthening of endothelium and mesenchyme
- Longitudinal muscles are required for zigzag and villi formation
- Role of longitudinal constraints

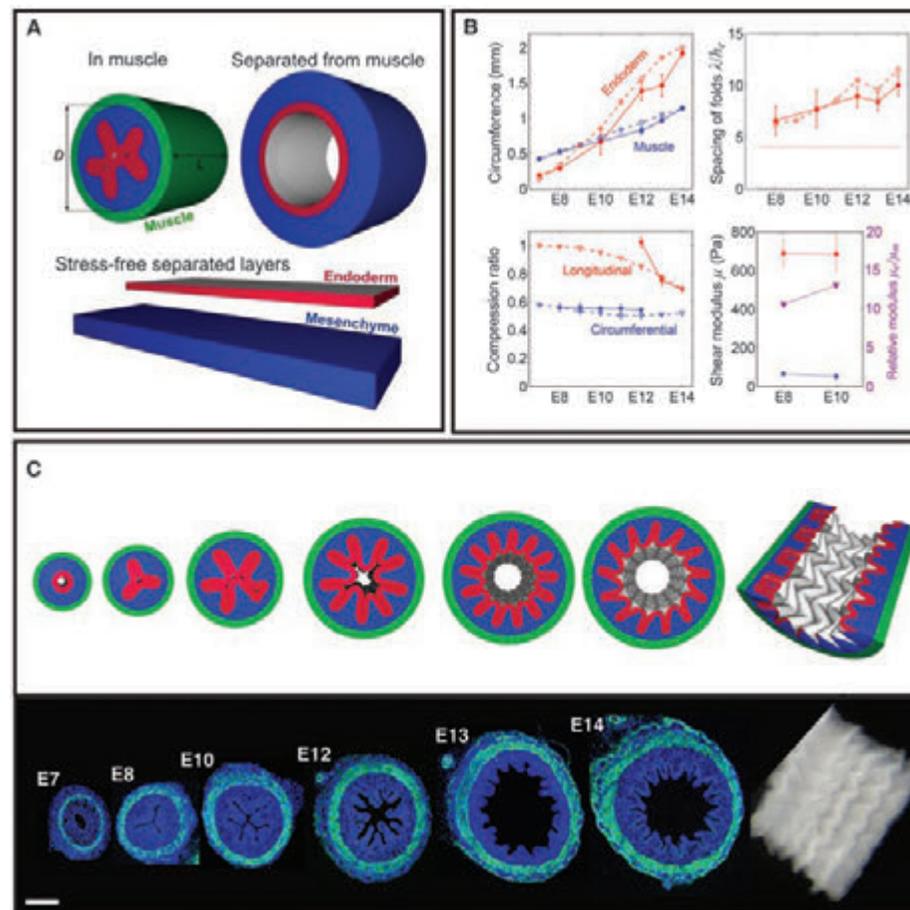


# Growth induced mechanical instabilities: Gut vilification

- Tissue zigzags and villi arise from mechanical instability caused by differential growth and constraints

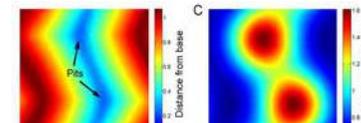
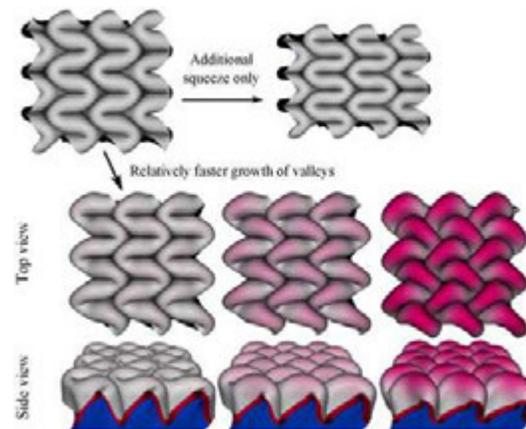
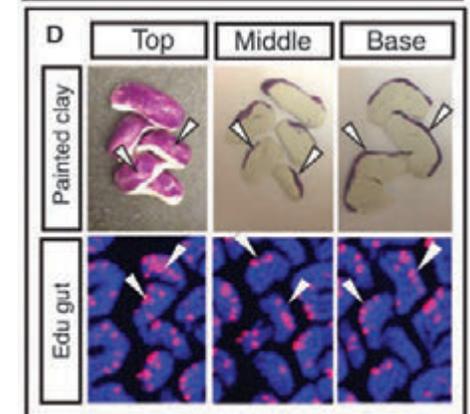
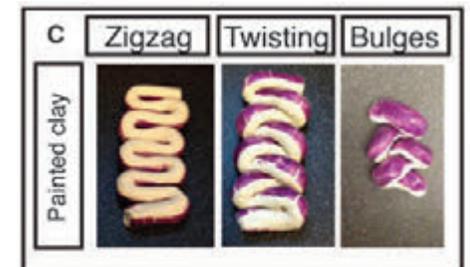
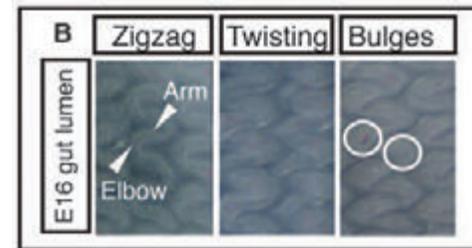
E. Hannezo J. Prost and J-F. Joanny *PRL* 107, 078104 (2011)

- Computation model      Similar earlier theoretical studies: M. Ben Amar and F. Jia. *PNAS* 110: 10525–10530 (2013)



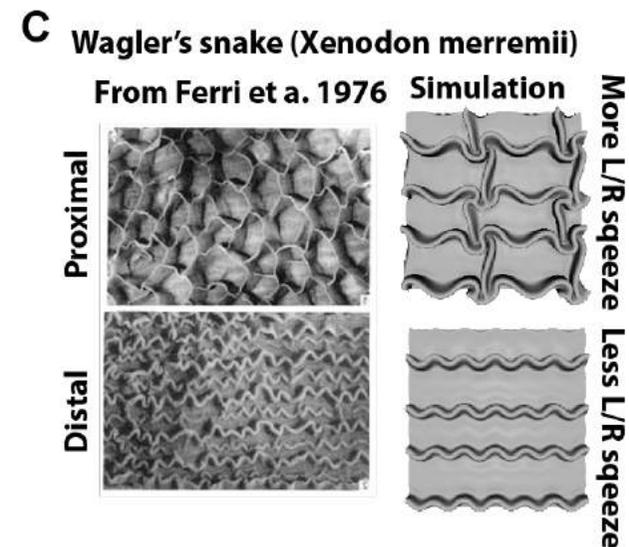
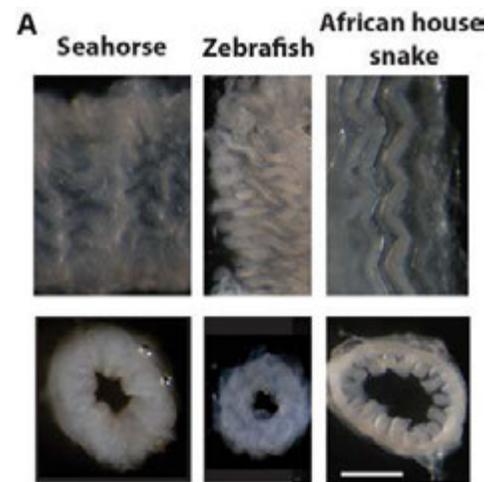
# Growth induced mechanical instabilities: Gut vilification

- Non-uniform growth also underlies formation of villi
  - Proliferation concentrated in furrows
  - Proliferating cells cause twisting and upward movement of proliferating cells
  - Leads to formation of villi
  - Computer simulations recapitulate this transition



# Growth induced mechanical instabilities: Gut vilification

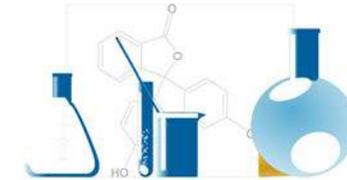
- Exploring the morphospace of vilification with mechanical instabilities



# Gut vilification with Turing instabilities

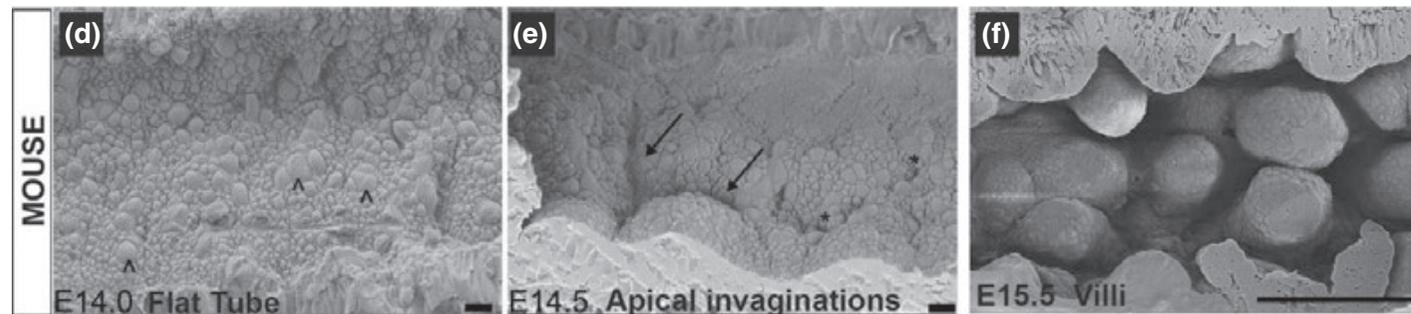
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- Self-organised mechanics: chick gut
- Self-organised biochemistry: Turing field in mouse gut



# Gut vilification with Turing instabilities

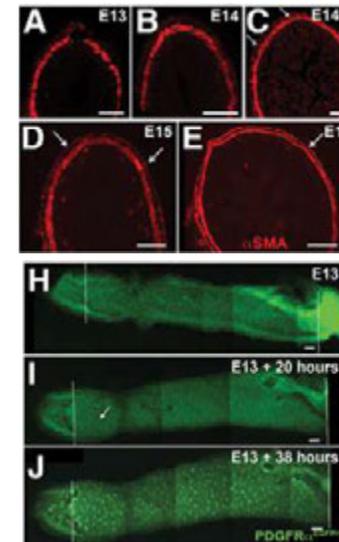
- Direct villus emergence from a flat epithelium in mammals



# Gut vilification with Turing instabilities

- No evidence of mechanical constraints governing vilification in the mouse

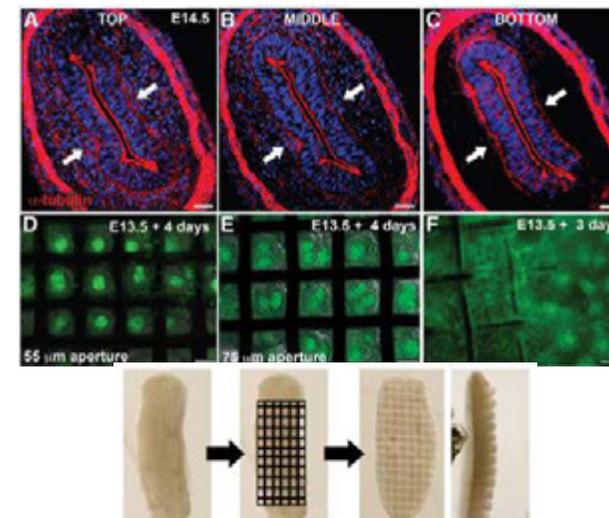
- Villus emergence precedes (by 24h) formation of longitudinal muscles
- Mechanical dissection of the gut does not affect timing of villi formation (circularity of gut affected by longitudinal cut)



SMA: marker of smooth muscles

PDGFRα: marker of mesenchymal condensation

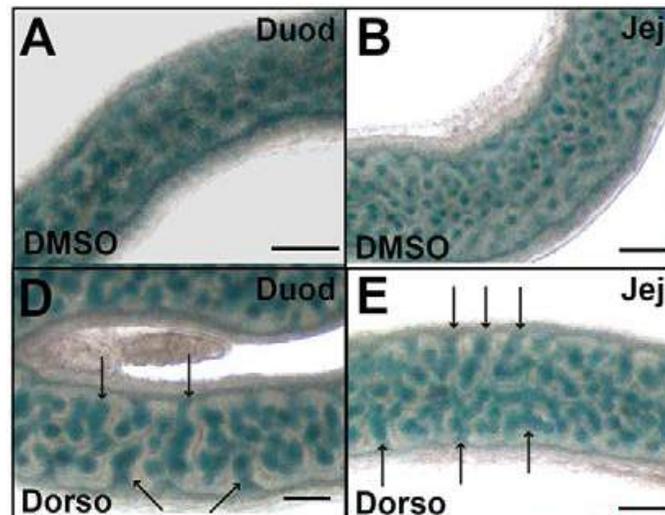
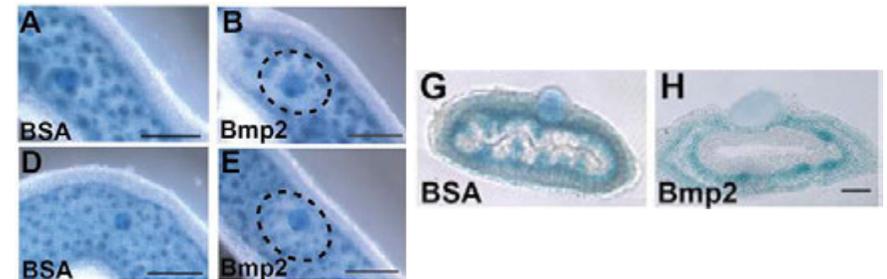
- Mesenchymal condensation at the base of epithelium are visible before any deformation of epithelial layer.
- Thus, mesenchymal condensation is an inducer of epithelial vilification
- Mechanical constraints on epithelial layers do not affect villi size.
- Must be controlled by mesenchymal signal



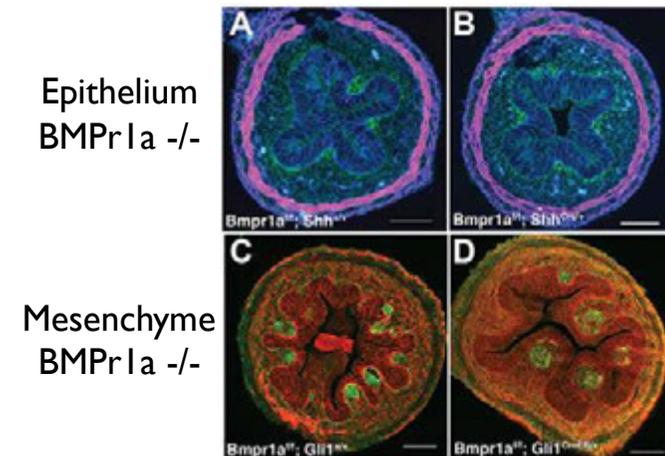
# Gut vilification with Turing instabilities

- Spatial patterns of vilification by developmental signalling (BMP)
  - BMP signals control vilification:
    - BMP2 beads inhibit mesenchymal condensation and vilification
    - BMP inhibitor causes enlargement of mesenchymal condensates (spots become stripes).
    - BMP signalling affects vilification in the mesenchyme but not in epithelium

Ptc marks mesenchymal condensates

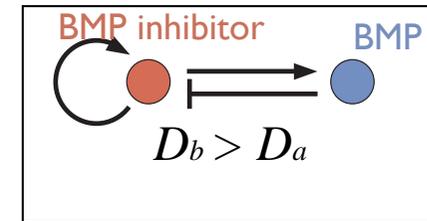


Dorsomorphin: BMP inhibitor

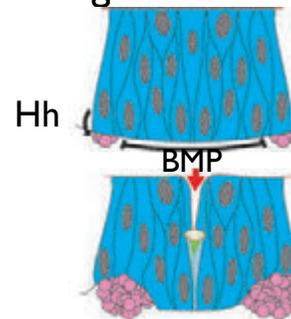
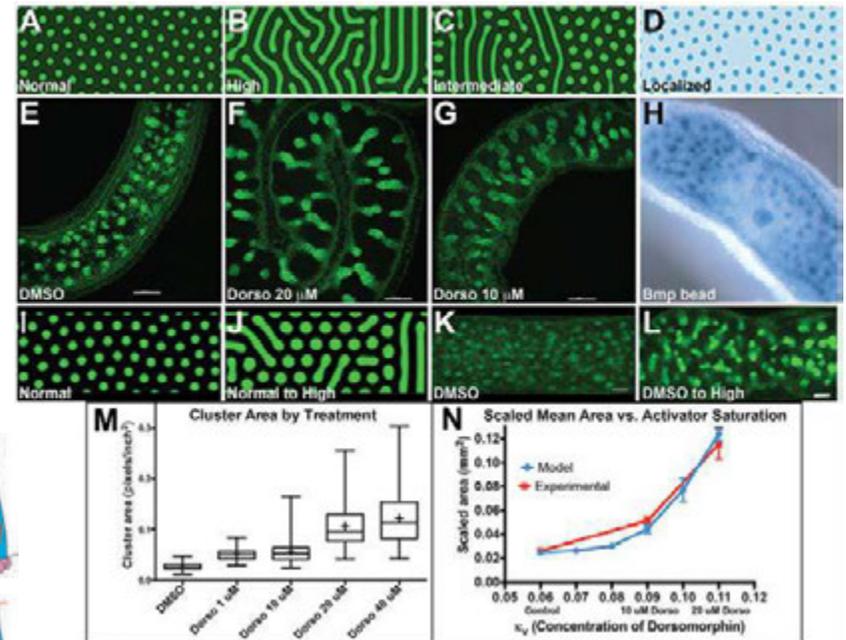


# Gut vilification with Turing instabilities

- A Turing reaction-diffusion model of vilification



- Turing activator: BMP inhibitor
- Turing inhibitor: BMP signals
- BMP regulators are co-expressed as required for Turing model
- Saturation of activator produces stripes in simulations and experiments
- Dynamic adaptation of pattern to dose of Turing activator (BMP inhibitor).



- However, mesenchymal condensation also emerge from Turing-like mechanical instabilities.
- Possibility of mechanical amplification of chemical instability (or vice versa).

Course 27 November

See: G.F. Oster, J.D. Murray, and A.K. Harris. *J. Embryol. esp. Morph.* 1983. 78:83-125  
 A.K Harris, D. Stopak and P. Warner. *J. Embryol exp. Morph.* 1984. 80:1-20  
 A. Shyer et al, R. Harland. *Science* 357: 811-817 (2017)



# How is the position of curvatures specified?

---

- Curvatures are not positioned deterministically (by upstream pre-pattern)
- They are self-organised due to mechanical or mechano-chemical instabilities
- Stereotypical pattern specified by geometry (shape of brain) and elastic properties.

## ✓ Organ morphogenesis

- But:
- Curvatures also arise in precisely defined position, following a highly reproducible pattern.

## ✓ Embryonic gastrulation and neurulation



# Tissue invagination

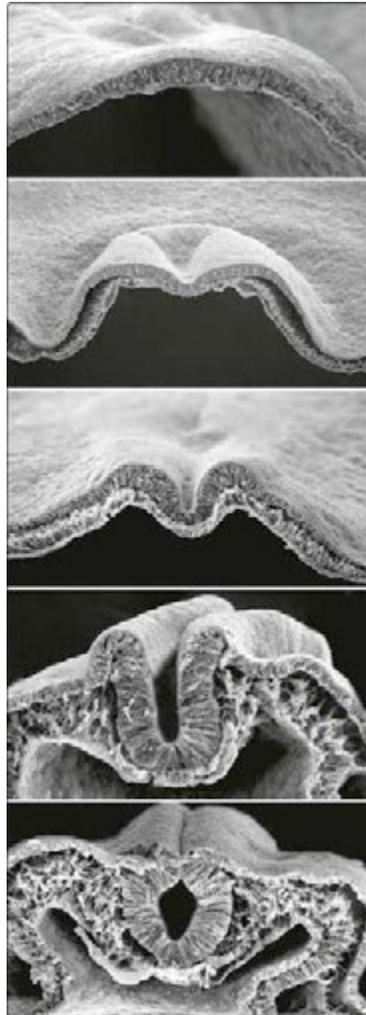
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## I. Controlled and Self-Organised mechanical instabilities

- Internal stress: eg. contractility
- External stress: boundary conditions (e.g. constraints associated with tissue geometry)
  
- Goal: Predict tissue morphogenesis (invagination) from cellular behaviours (cell contractility).

# Tissue deformations: control and self-organisation

## 2. Tissue invagination

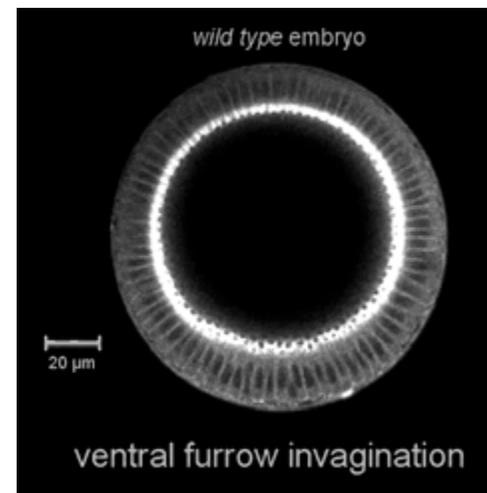


G. Schoenwolf U. Utah School of Medicine

Chick neural tube



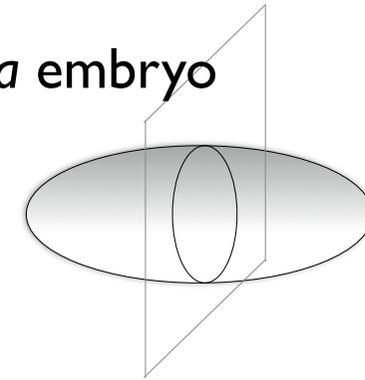
Sea Urchin endoderm



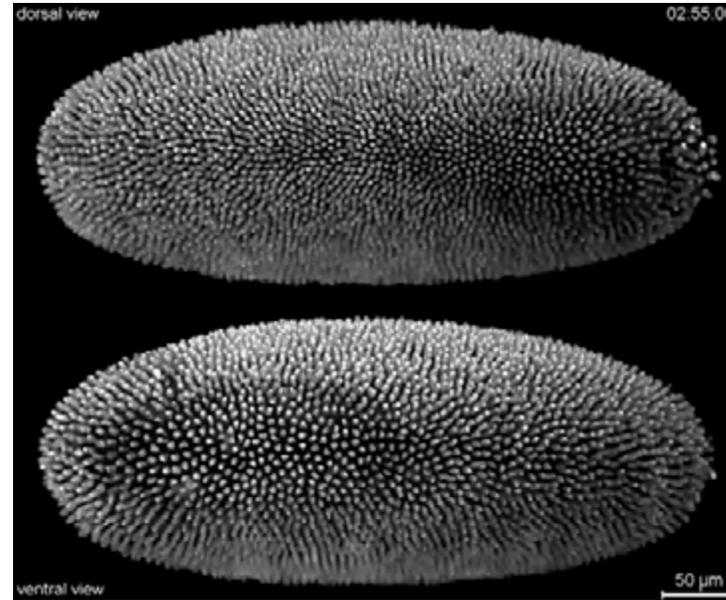
*Drosophila* mesoderm

# Programmed tissue invagination: Gastrulation

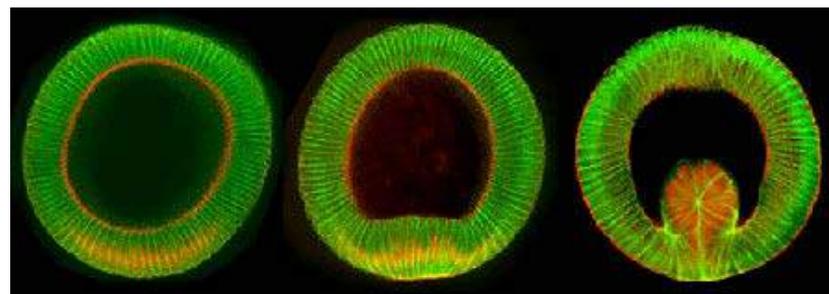
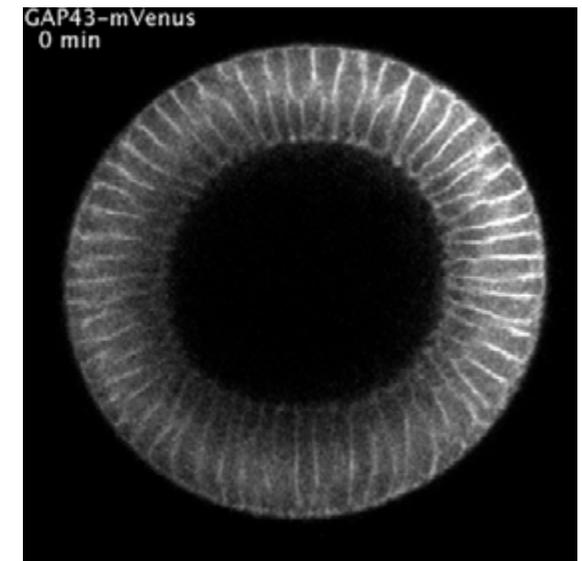
- Tissue invagination during gastrulation: the *Drosophila* embryo



- Gastrulation: separation of tissue layers (ectoderm, mesoderm and endoderm)
- The ventral epithelium (presumptive mesoderm) forms a furrow and invaginates
- Leads to the formation of two distinct tissues



Keller lab Janelia Research Campus



Twist

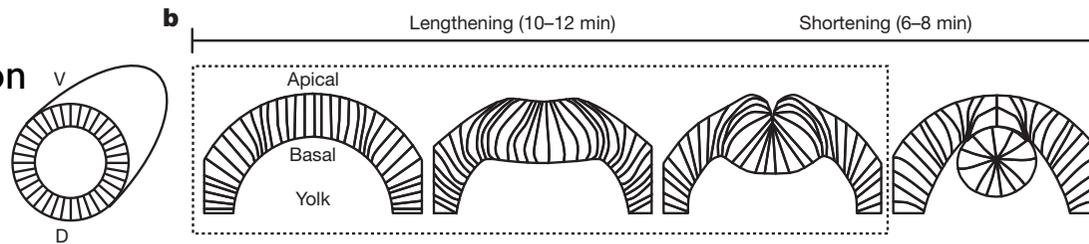


# Programmed tissue invagination: Gastrulation

## 3D cellular model

Cell shape changes:

- Cell apex constriction
- Cell lengthening
- Basal expansion

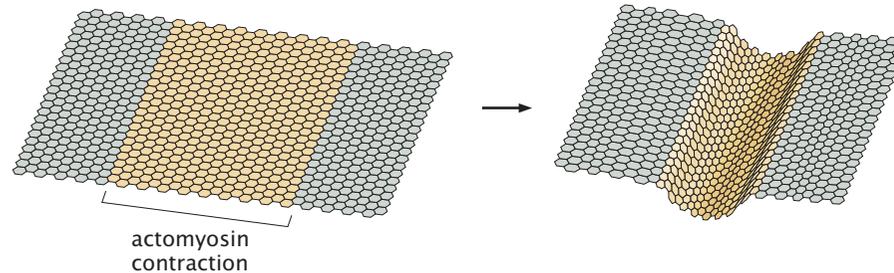


## 2D cellular model

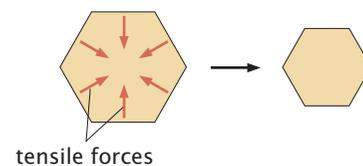
Cell shape changes:

- Apical cell constriction

(A) tissue invagination



(B) apical constriction

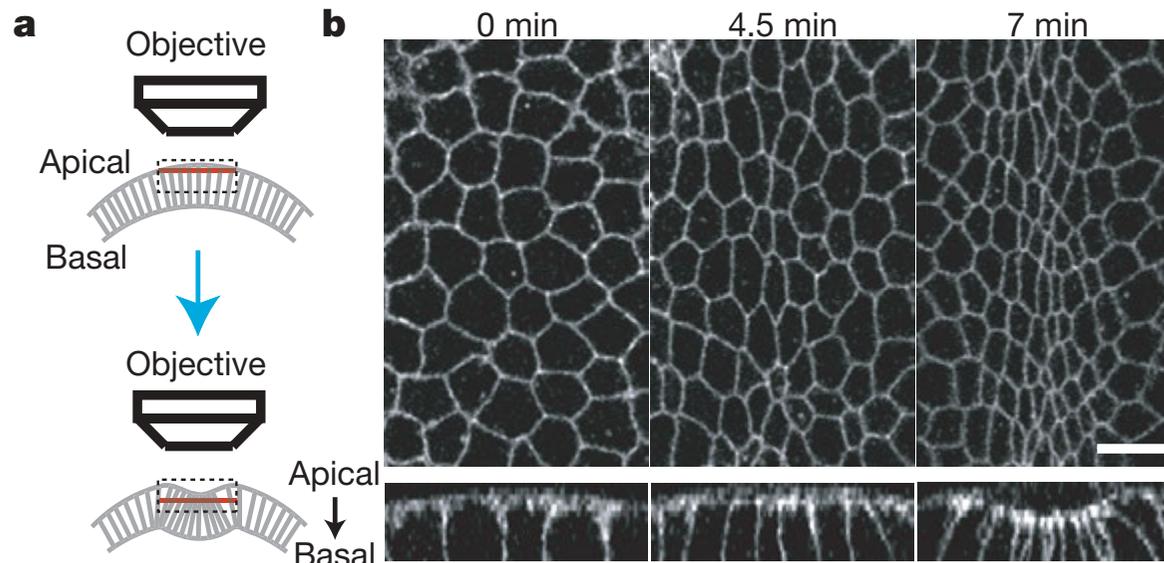


ill. Nigel Orme

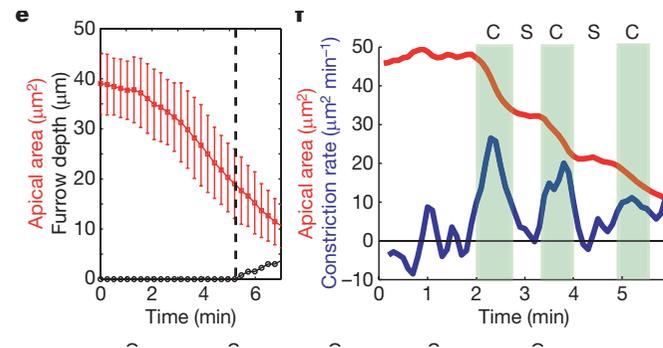


# Programmed tissue invagination: Gastrulation

2D cellular model — Tissue furrowing correlates with apical cell constriction



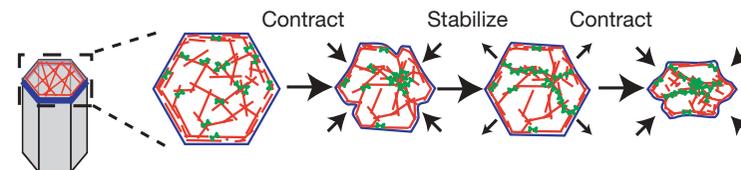
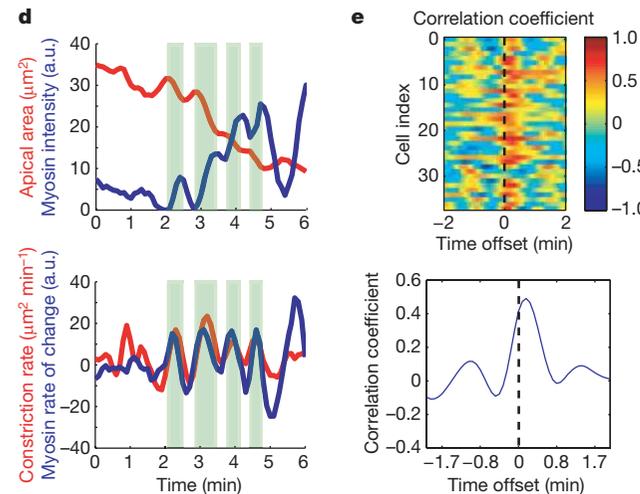
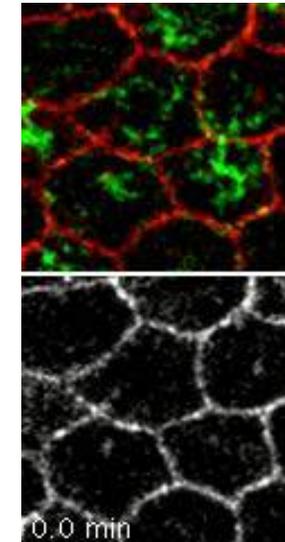
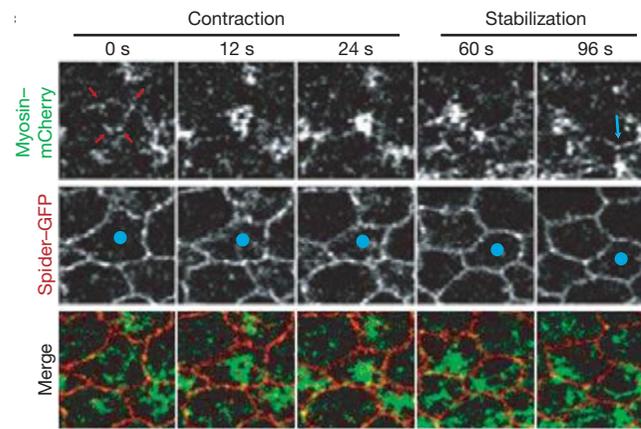
- Constriction is pulsatile



# Programmed tissue invagination: Gastrulation

## 2D cellular model — Apical cell constriction is driven by contractile actomyosin pulses

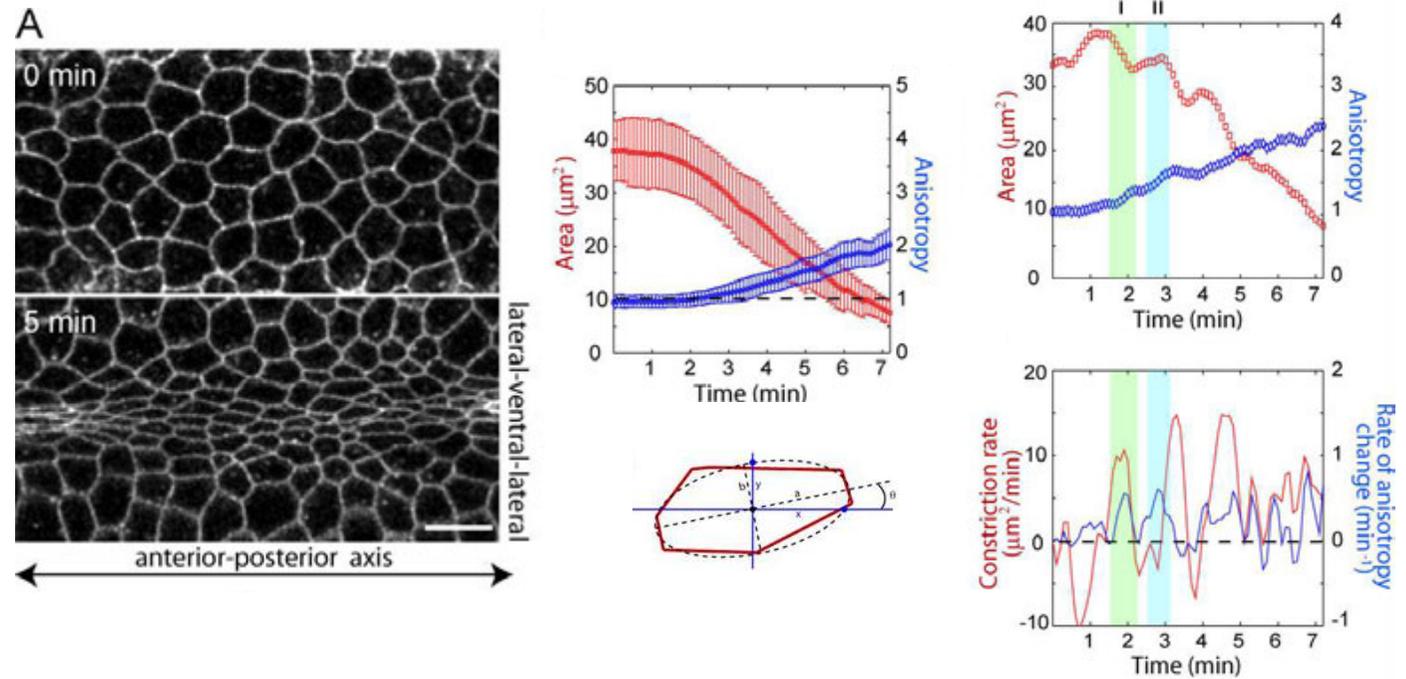
- Myosin-II contractility is pulsatile
- Cell apical constriction correlates with Myosin-II pulsation
- Phases of constriction alternate with phases of stabilisation of cell shape
- A « mechanical ratchet » ensures irreversible cell and tissue deformations



# Programmed tissue invagination: Gastrulation

## 2D cellular model — Cell-Cell mechanical coupling

- Cells are strained along the anteroposterior axis
- Anisotropic cell constriction

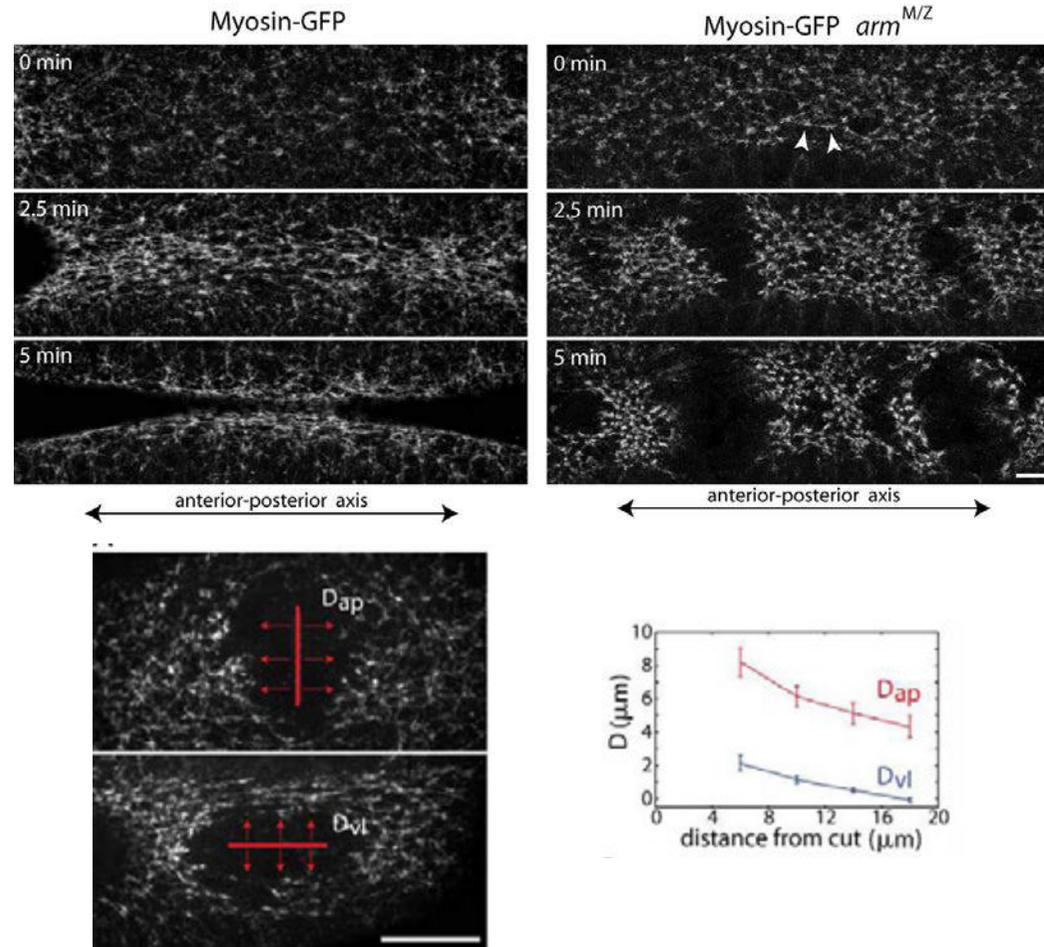
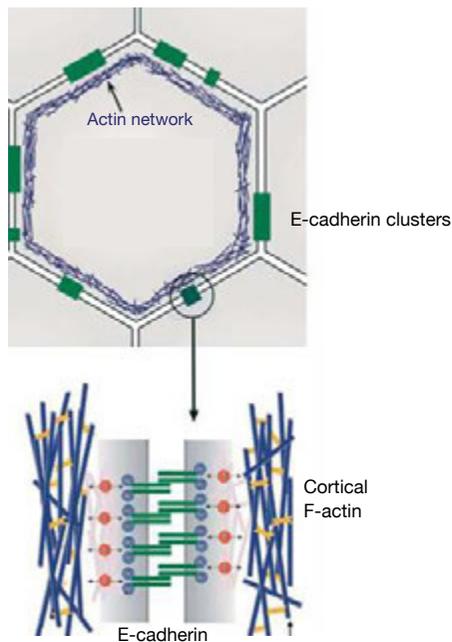


A. Martin et al. and E. Wieschaus. *J. Cell Biol.* (2010) 188 (5):735–749

# Programmed tissue invagination: Gastrulation

2D cellular model — Anisotropic apical cell constriction is due to increased tension along the anteroposterior axis

- Integrity of cell-cell junctions underlies tension build up via cell mechanical coupling

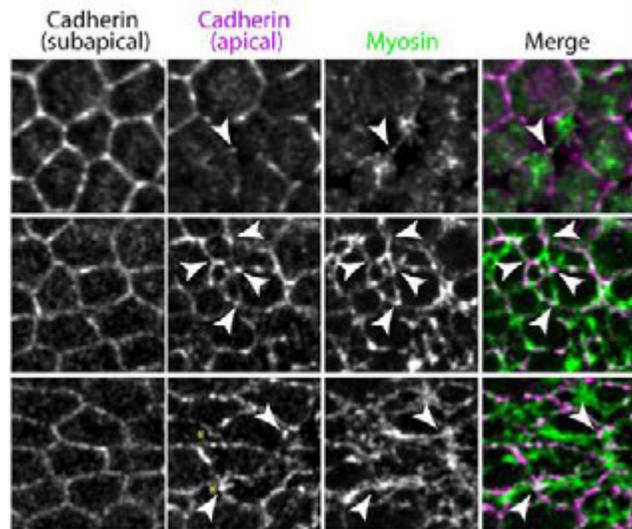


Laser ablation experiments: relaxation kinetics is related to total mechanical stress

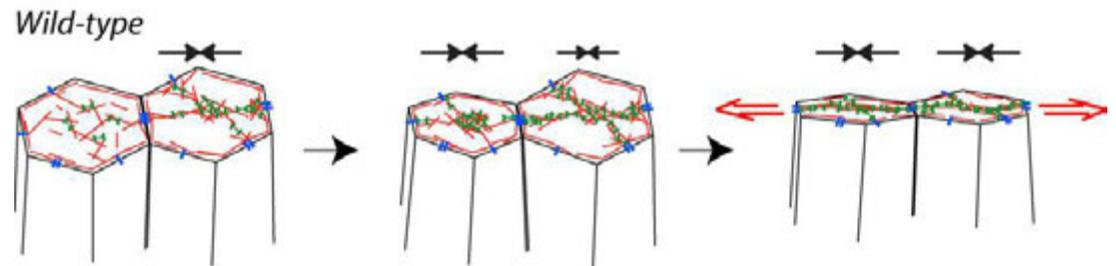
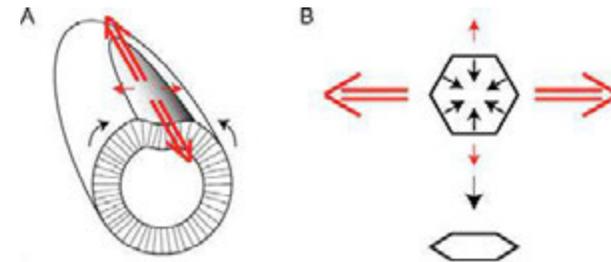


# Programmed tissue invagination: Gastrulation

2D cellular model — Cell-cell mechanical coupling requires actomyosin coupling to cell junctions



- Multicellular actomyosin network underlies cell mechanical coupling



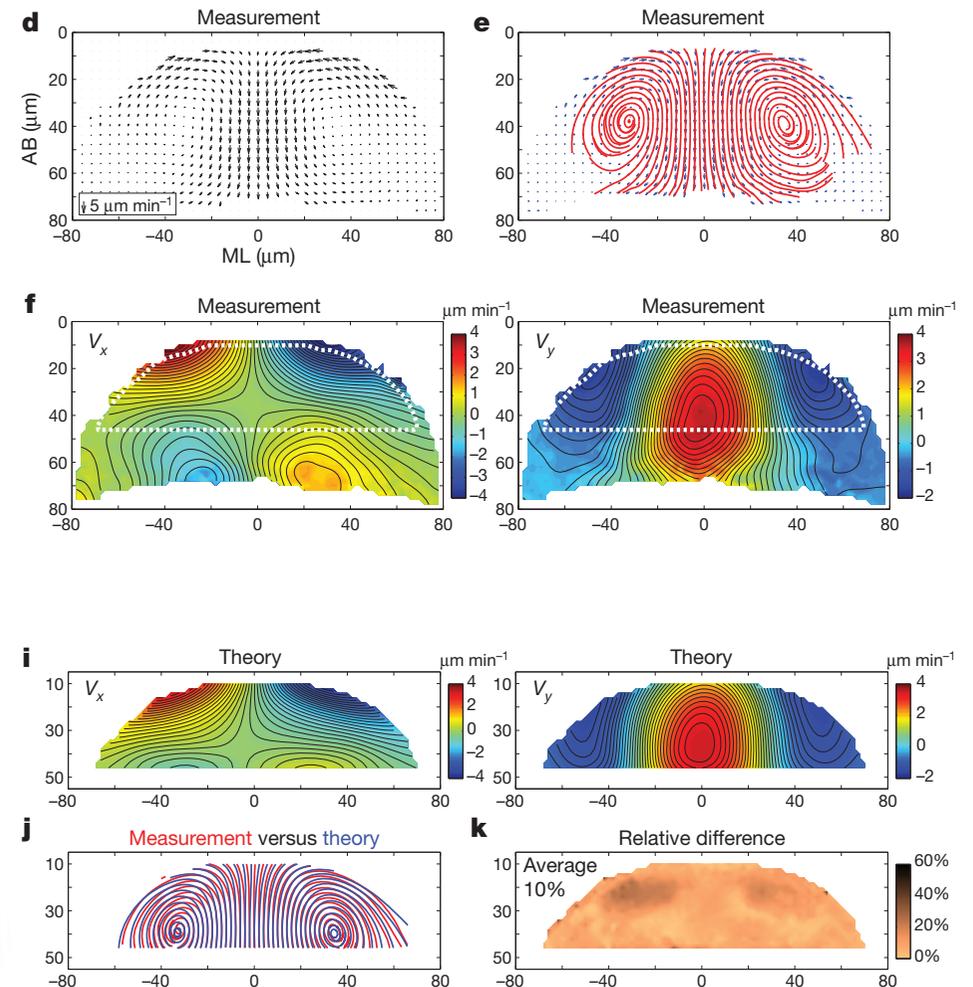
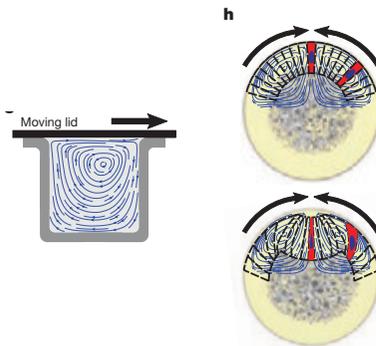
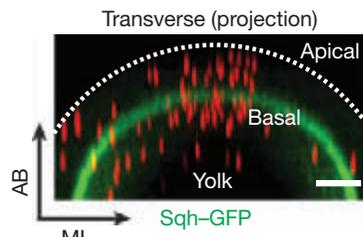
A. Martin et al. and E. Wieschaus. *J. Cell Biol.* (2010) 188 (5):735–749



# Programmed tissue invagination: Gastrulation

## 2D cellular model: apical constriction drives tissue scale hydrodynamic flow

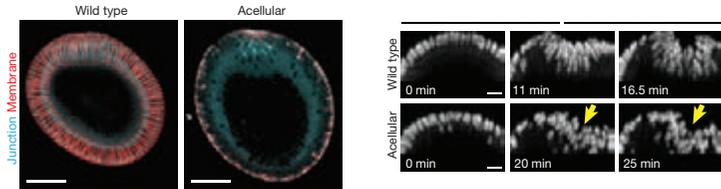
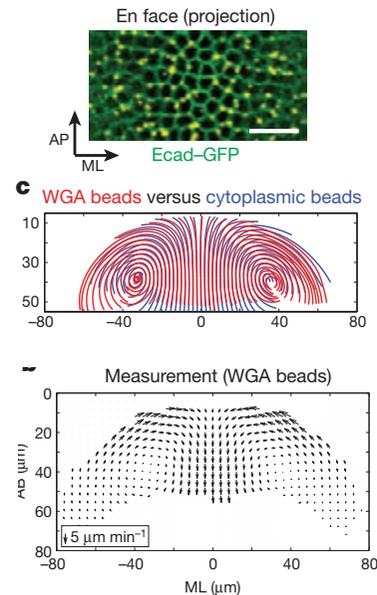
- Accounts for tissue flow at the onset of tissue invagination (furling)
- Stokes equations predict viscous flow at low Reynolds number regime:
- flow velocity at the interior of a domain is uniquely determined by velocities at the domain boundary.
- Suggests that tissue flow arises from shear forces associated with apical contractility at tissue scale



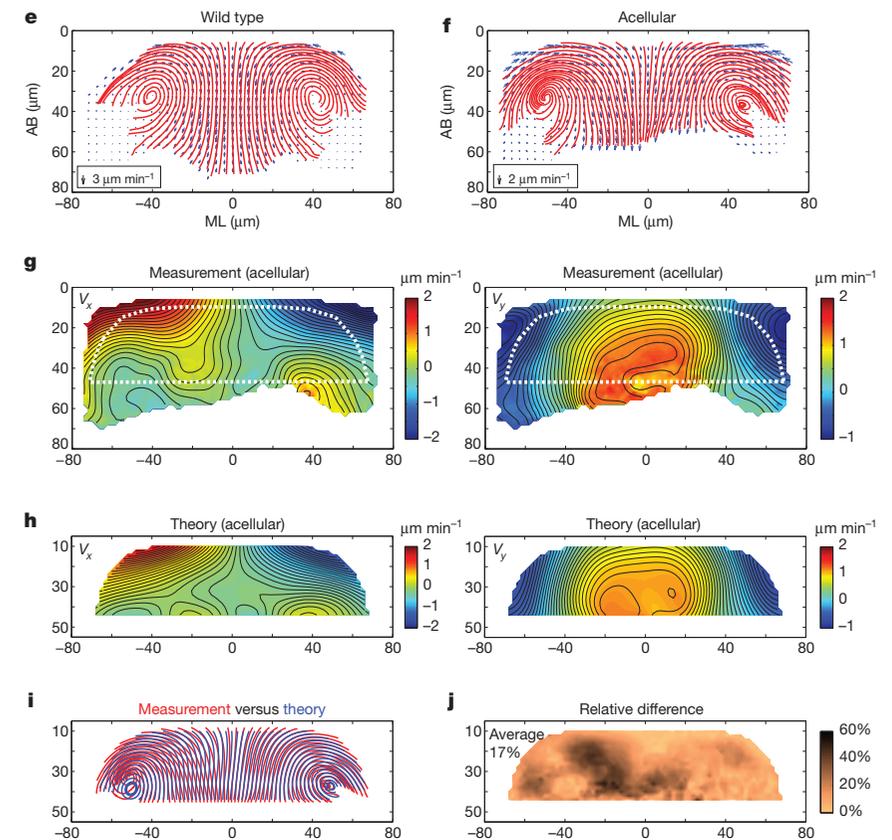
# Programmed tissue invagination: Gastrulation

## 2D cellular model: apical constriction drives tissue scale hydrodynamic flow

- Flows of cytoplasmic beads and membrane bound beads are similar suggesting that cells passively respond to cytoplasmic flow
- The individuality of cells is not necessary for tissue hydrodynamic flow: flow in acellular embryo is similar to normal embryos

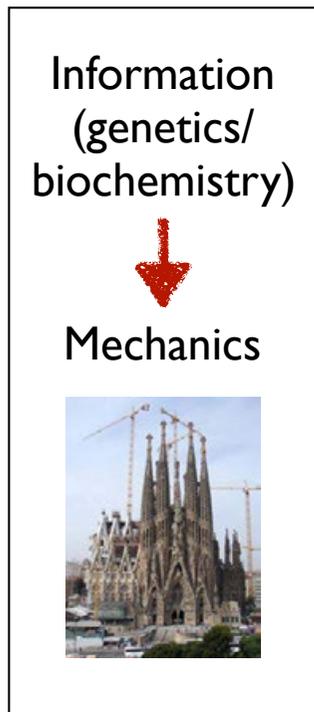


Acellular embryos

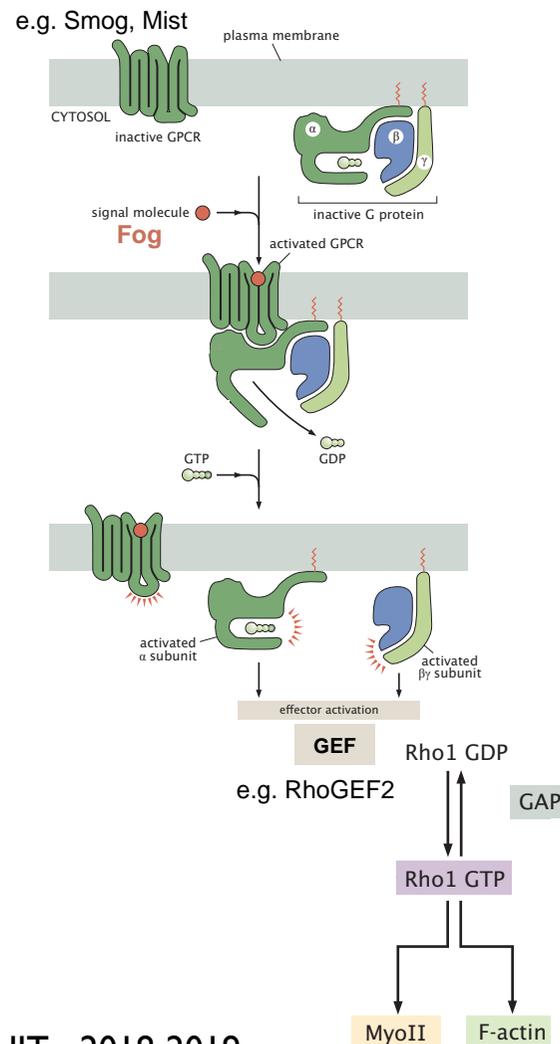


# Programmed tissue invagination: Gastrulation

## 2D cellular model: The spatial program controlling apical cortical mechanics

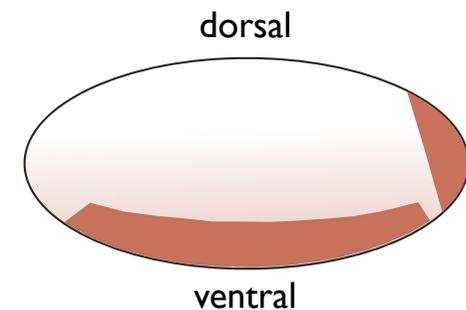


**The pathway:**  
GPCR and G protein signalling underlies  
Myosin2 activation

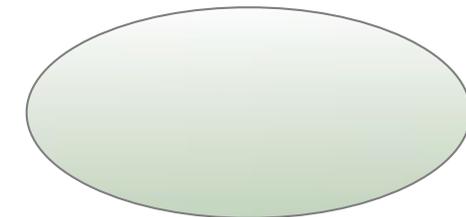


**The program:**  
Embryonic prepattern spatially  
controls signalling

**Switch  
component**



**Ubiquitous  
factors**

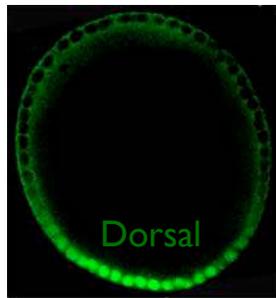


Smog,  
G proteins,  
GEFs,  
Rho1, ...

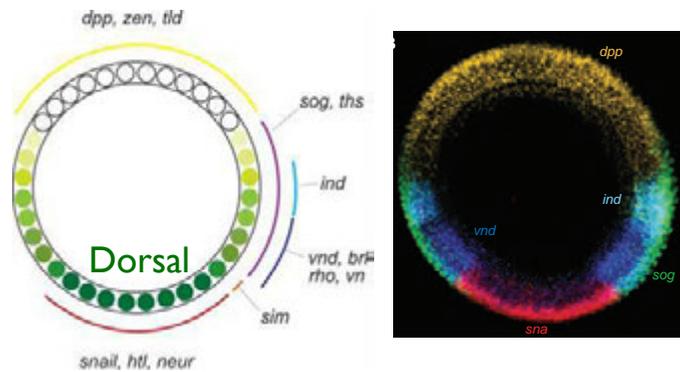
# Programmed tissue invagination: Gastrulation

## 2D cellular model: The spatial program controlling apical cortical mechanics

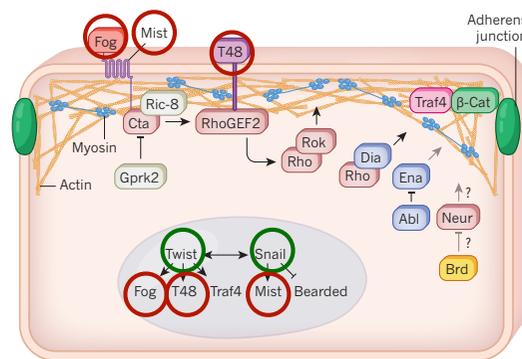
The pathway:  
Positional information and spatial activation of Myosin2



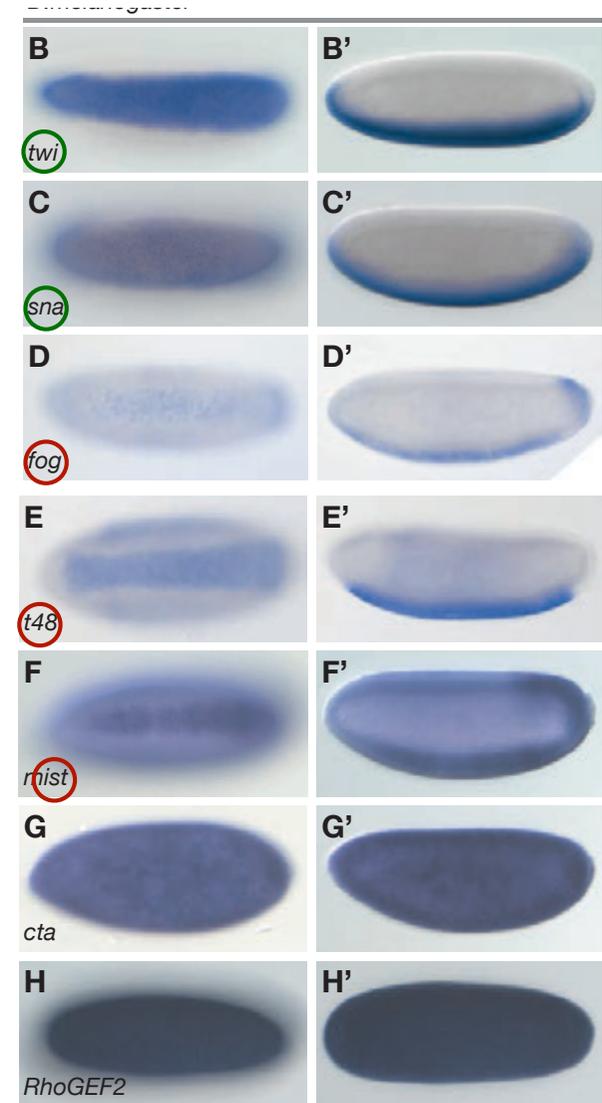
B. Shilo lab



M. Levine. PNAS, 2008 105 (51) 20072-20076;



D. Gilmour, M. Rembold and M. Leptin. *Nature* 541:311-320 (2017)

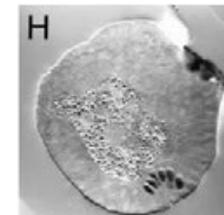
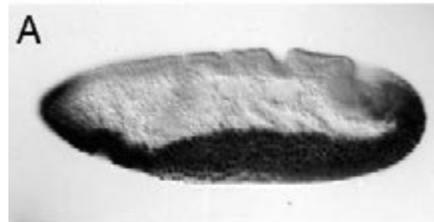


Urbansky et al. S. Lemke. *eLife* 2016;5:e18318

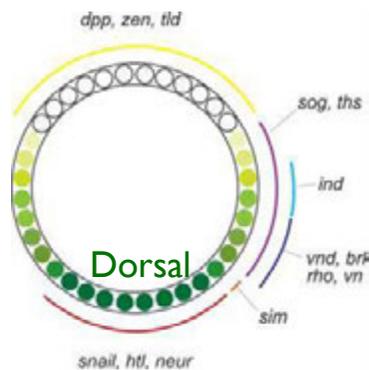
# Programmed tissue invagination: Gastrulation

## 2D cellular model: The spatial program controlling apical cortical mechanics and invagination

Snail positive cells can invaginate on a cell-autonomous basis. Indicates that cell autonomous activation of the « invagination program » is sufficient to induce proper invagination.



host	snail/twist -/-
donor	wt



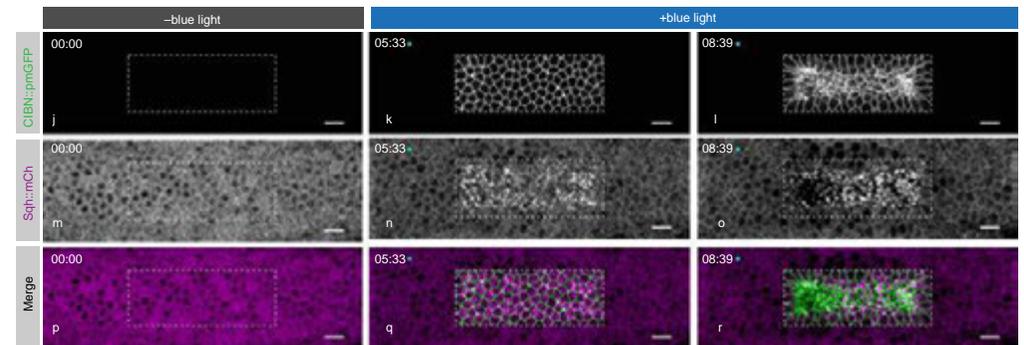
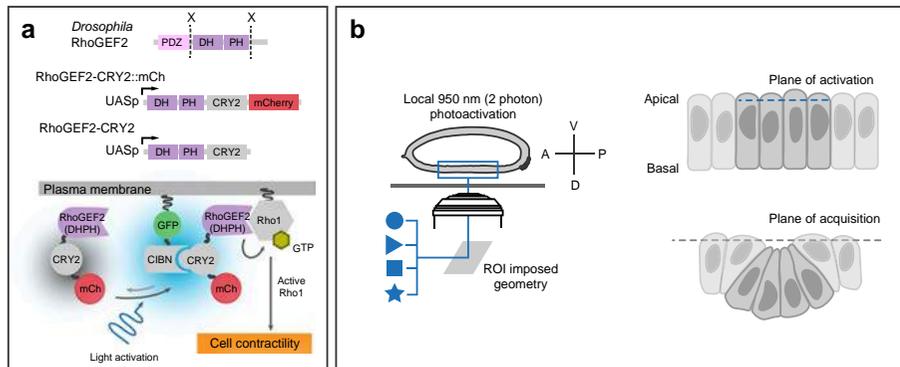
host	Toll -/-
donor	wt

M. Leptin and S. Roth *Development* 120, 853-859 (1994)

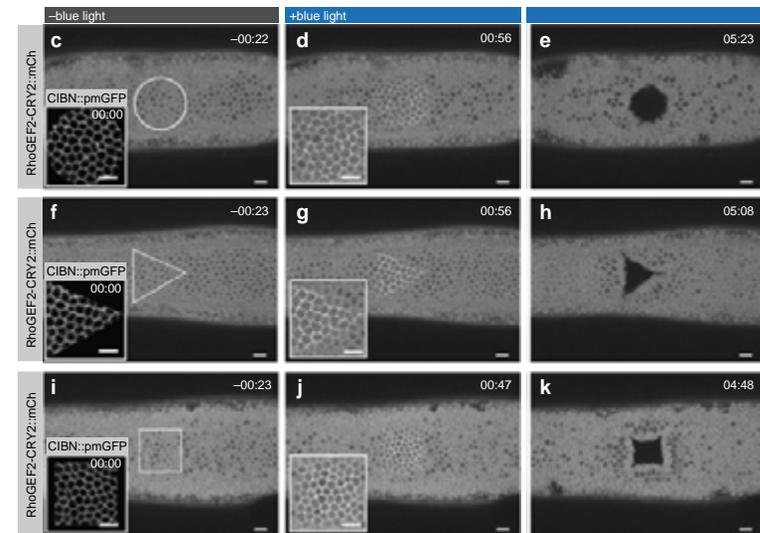


# Programmed tissue invagination: Gastrulation

## 2D cellular model: The spatial program controlling apical cortical mechanics and invagination



- Optogenetic activation of the Rho|GTP pathway
- Rho1 activation causes MyosinII activation cell autonomously
- Rho1 activation is sufficient to induce tissue invagination. The geometry of activation directs the geometry of invagination

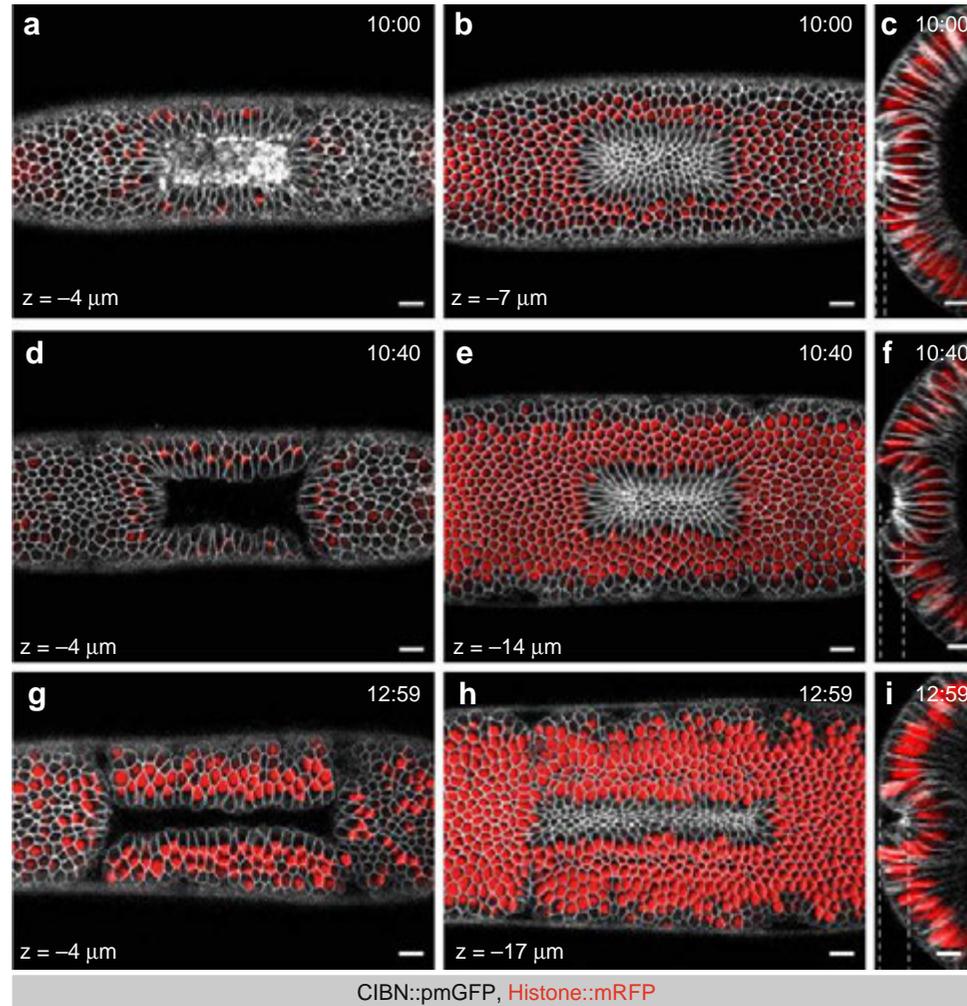


E Izquierdo, T. Quikler and S. de Renzis *Nature Communications*. (2018)9:2366 | DOI: 10.1038/s41467-018-04754-z

# Programmed tissue invagination: Gastrulation

## 2D cellular model: The spatial program controlling apical cortical mechanics and invagination

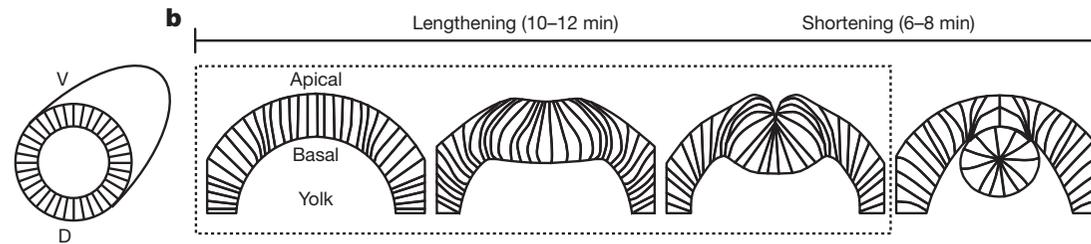
- Optogenetic activation of the Rho1GTP pathway causes *complete* tissue invagination, suggesting that apical contractility is sufficient for complete invagination



E Izquierdo, T. Quikler and S. de Renzis *Nature Communications*. (2018)9:2366 | DOI: 10.1038/s41467-018-04754-z

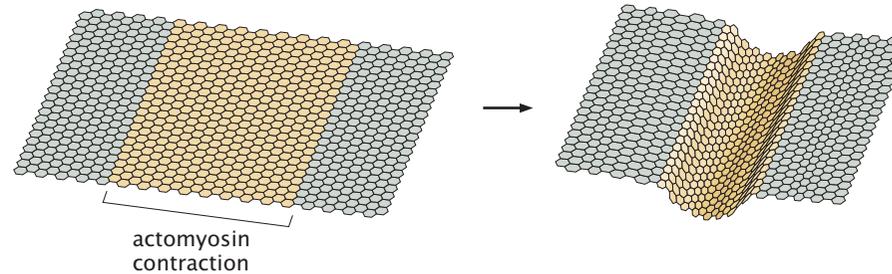
# Programmed tissue invagination: Gastrulation

## 3D cellular model

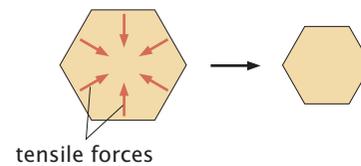


## 2D cellular model

(A) tissue invagination

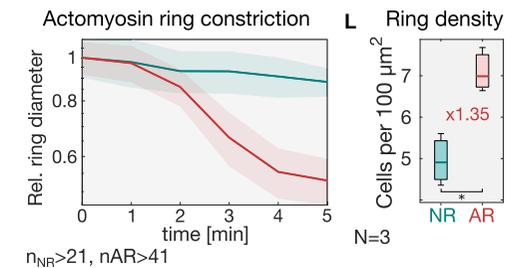
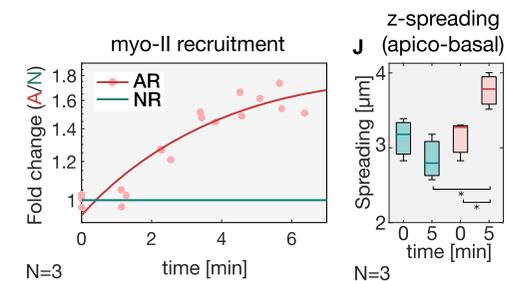
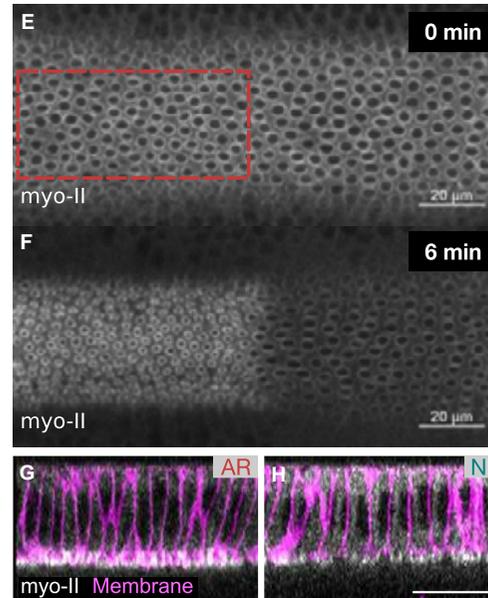
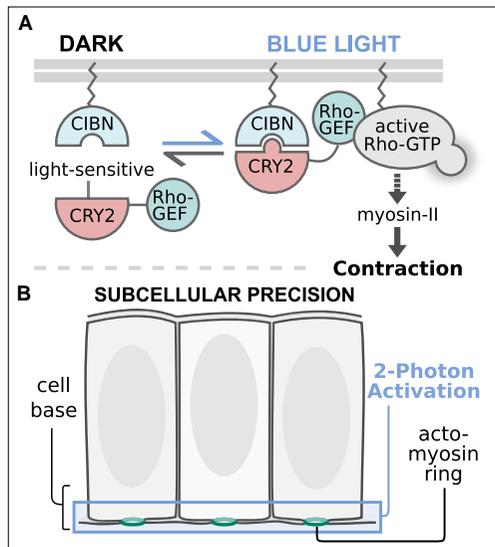
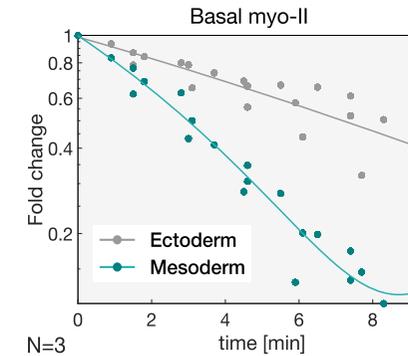
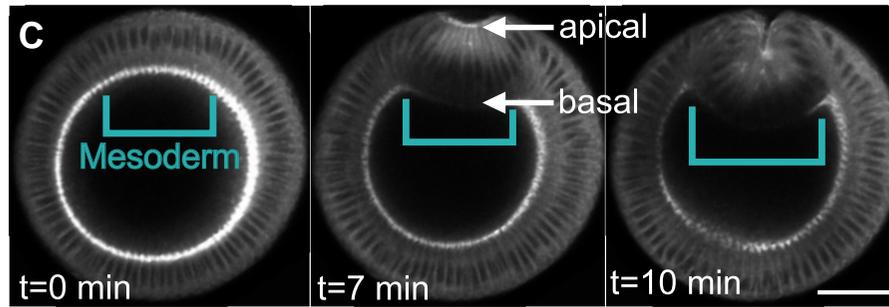


(B) apical constriction



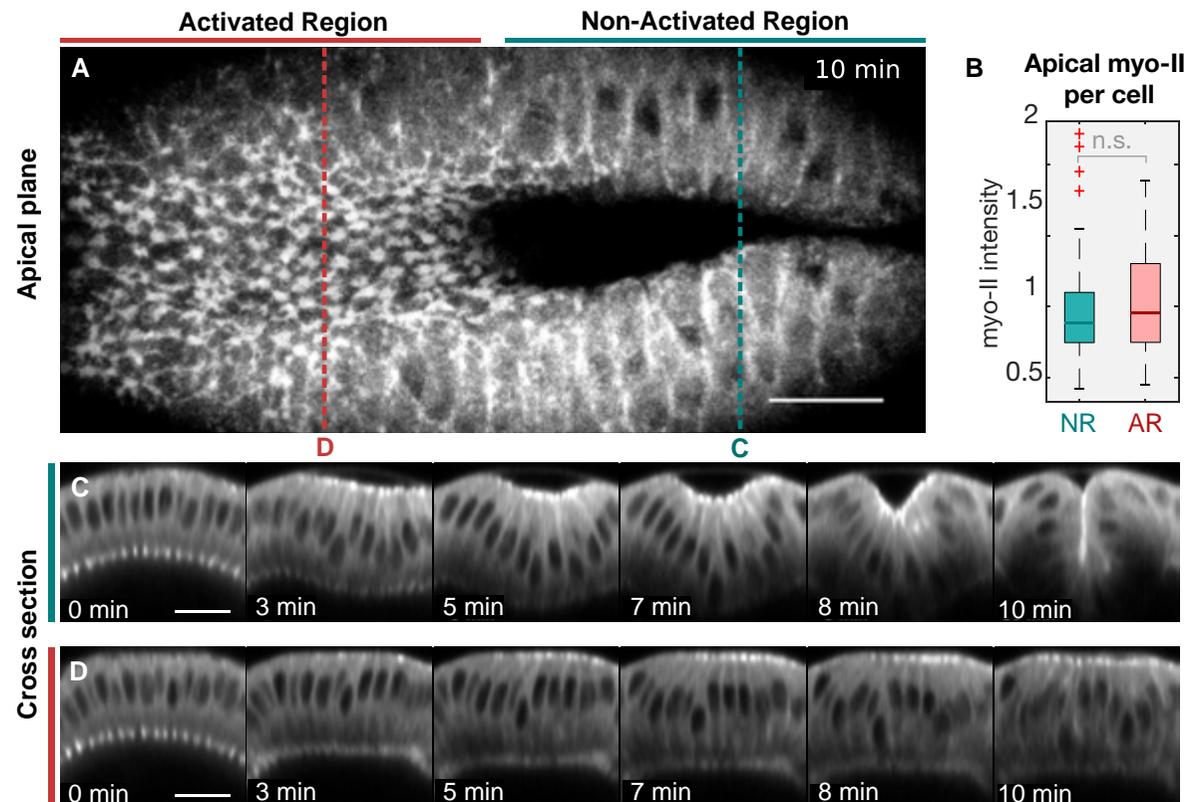
# Programmed tissue invagination: Gastrulation

## 3D cellular model: basal inhibition of Myosin-II is required for invagination



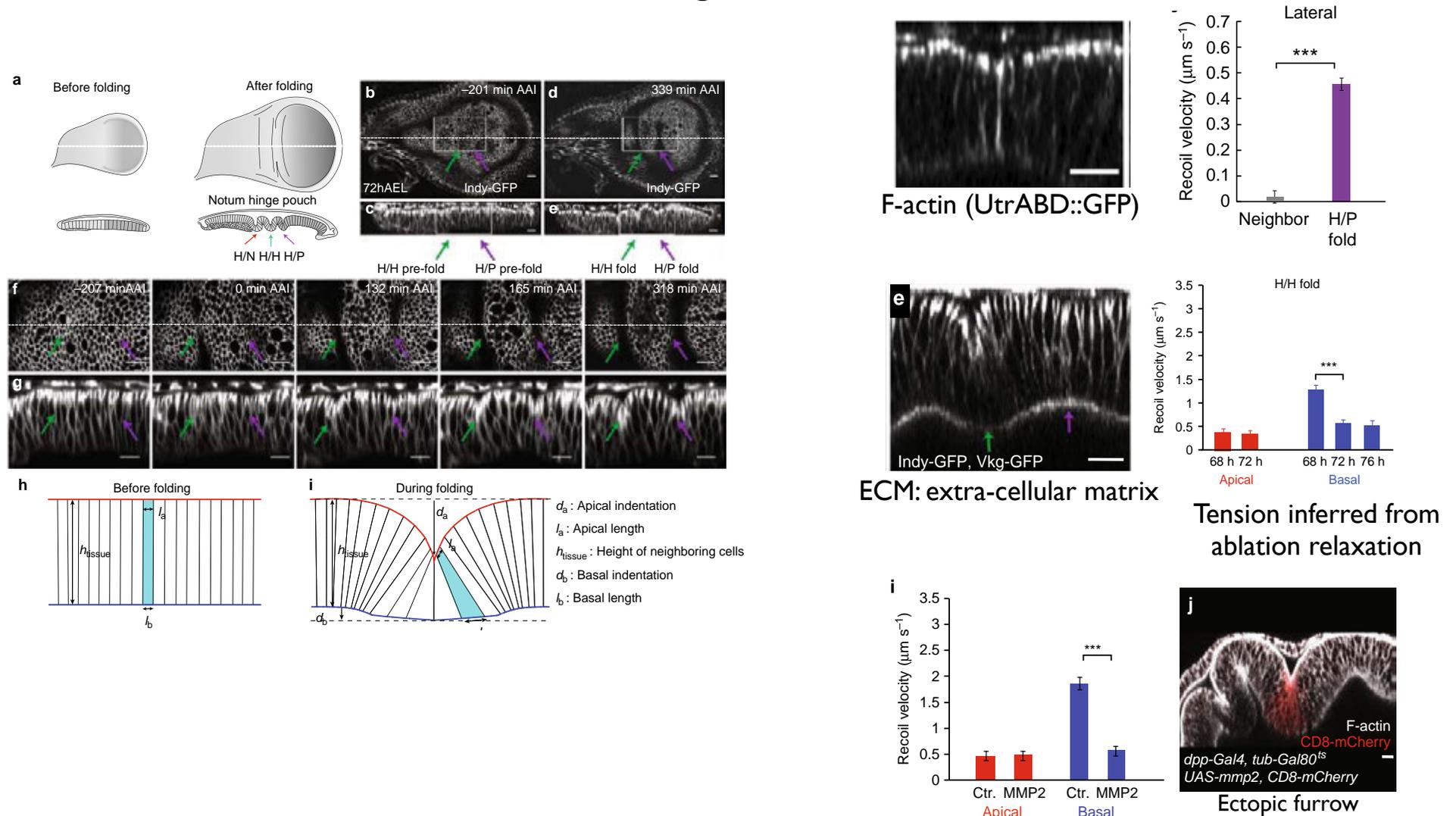
# Programmed tissue invagination: Gastrulation

3D cellular model: basal inhibition of Myosin-II is required for invagination



# Programmed tissue invagination: Gastrulation

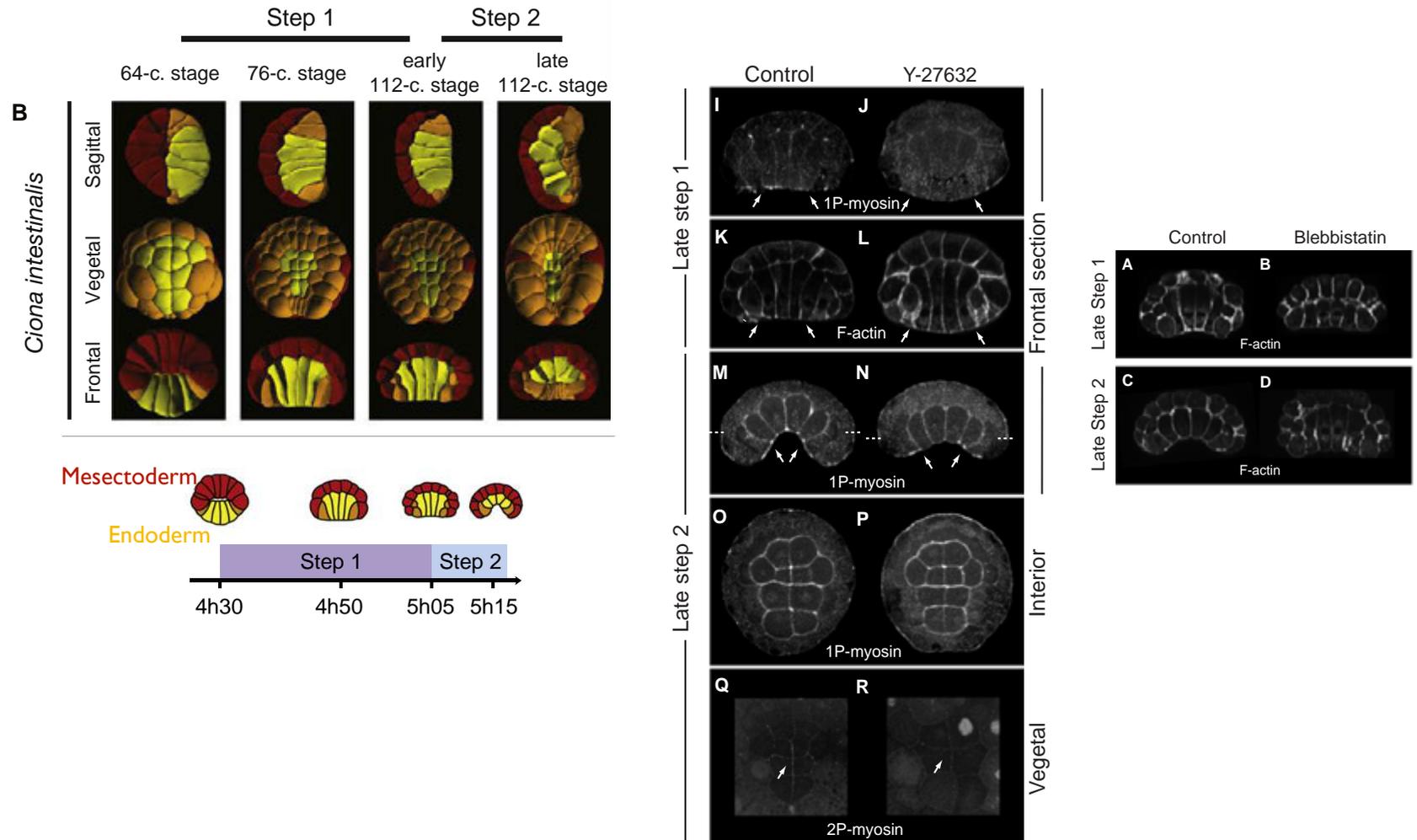
## 3D cellular model: increased lateral tension and basal tension relaxation induce tissue invagination



# Programmed tissue invagination: Gastrulation

## 3D cellular model: increased apical and lateral tension induce tissue invagination

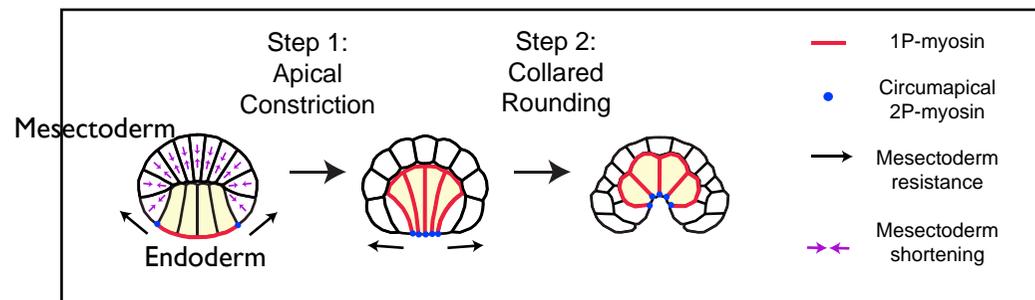
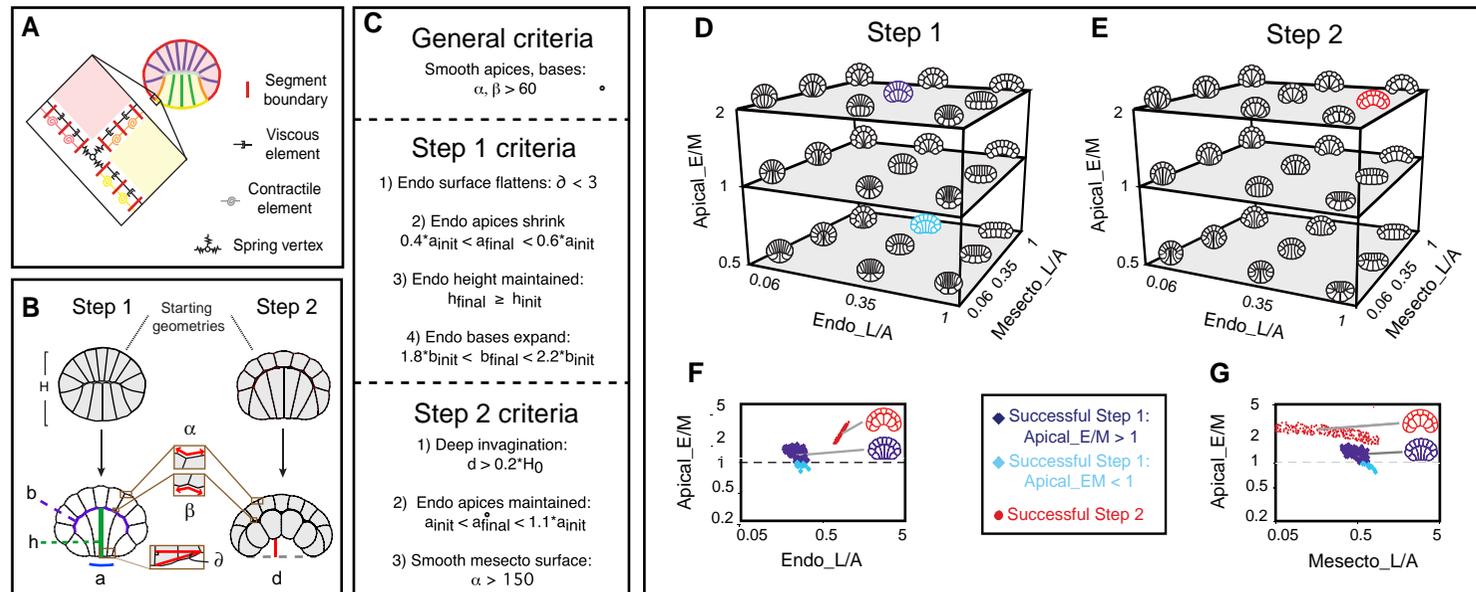
- Sequential apical and lateral recruitment of Myosin-II to cell cortex



K. Sherrard, F. Robin, P. Lemaire and E. Munro. *Current Biology* 20, 1499–1510 (2010)

# Programmed tissue invagination: Gastrulation

## 3D cellular model: increased apical and lateral tension induce tissue invagination



K. Sherrard, F. Robin, P. Lemaire and E. Munro. *Current Biology* 20, 1499–1510 (2010)

See also: M. Rauzi et al, and M Leptin. *Nature Communications* | 6:8677 | DOI: 10.1038/ncomms9677 |

# Programmed tissue invagination: Gastrulation

The program of Invagination:  
Spatial and temporal control over cortical mechanics:  
active stress generation (MyosinII contractility)

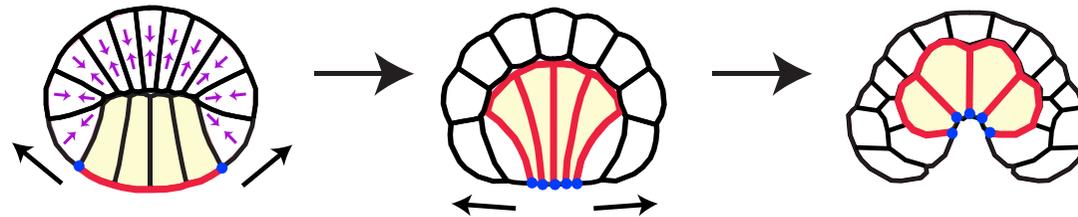
Information  
(genetics/  
biochemistry)



Mechanics

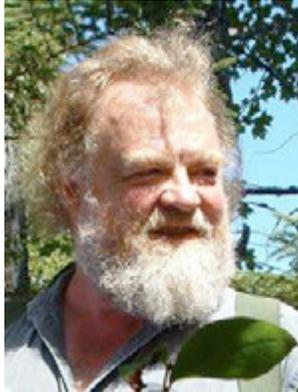


- Regionalisation
- Polarisation



# Self-organised tissue invagination

- **Self-organisation of invagination:**  
given initial trigger, cell deformation and spatial pattern emerges from cell mechanical properties



Garrett Odell  
1943-2018



George Oster  
1940-2018

## The Mechanical Basis of Morphogenesis

### 1. Epithelial Folding and Invagination

G. M. ODELL,<sup>1</sup> G. OSTER, P. ALBERCH, AND B. BURNSIDE

*University of California, Berkeley, California 94720*

In our model, coordination at the population level arises from the local behavior of each cell automatically; there is no need to introduce poorly understood devices such as morphogens and cellular clocks.

In this study we want to minimize both the number of complex instructions (e.g. morphogens and clocks) as well as the genetic programming required to generate morphogenetic patterns.

Second, we wanted to minimize the load of genetic preprogramming required to generate morphogenetic patterns. We felt that to equip each cell with an autonomous program, and a precise “clock” for activating that program, was unesthetic and probably unnecessary. At least part of the burden of pattern formation and regulation may be taken up by the equilibration of purely mechanical forces.

Our model, therefore, is built on Newton’s laws of motion and consists of a dynamical system of ordinary differential equations whose solution determines the global time history of the cell sheet geometry, given its initial configuration.

G.M. Odell, G. Oster, P. Alberch and B. Burnside. *J. Math. Biol.* (1980) 9:291-295

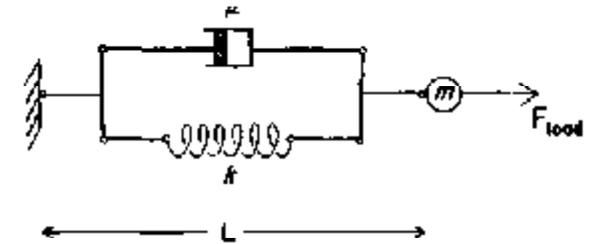
G.M. Odell, G. Oster, P. Alberch and B. Burnside. *Dev. Biol.* (1981) 85:446-462



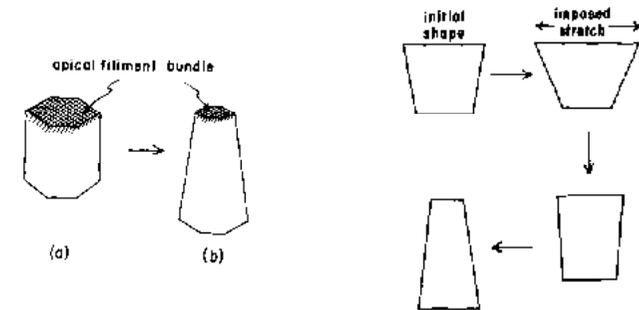
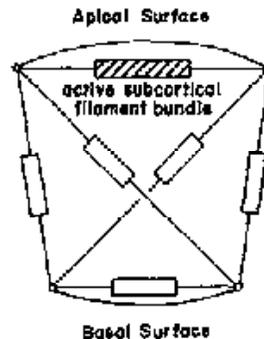
# Self-organised tissue invagination

- Task: coordinating at population level (tissue) individual cell behaviours in space and time

- Cells are mechanically coupled
- Apical contractility
- Volume incompressibility
- **The apical cortex of cells is excitable (active spring):**
  - small cell stretching causes relaxation to rest configuration  $L_0$
  - above threshold, cell stretching causes cell « firing » of contractility and return to a new rest configuration



$$\frac{dL}{dt} = -\frac{k}{\mu}(L - L_0) + \frac{1}{\mu}F_{load}$$

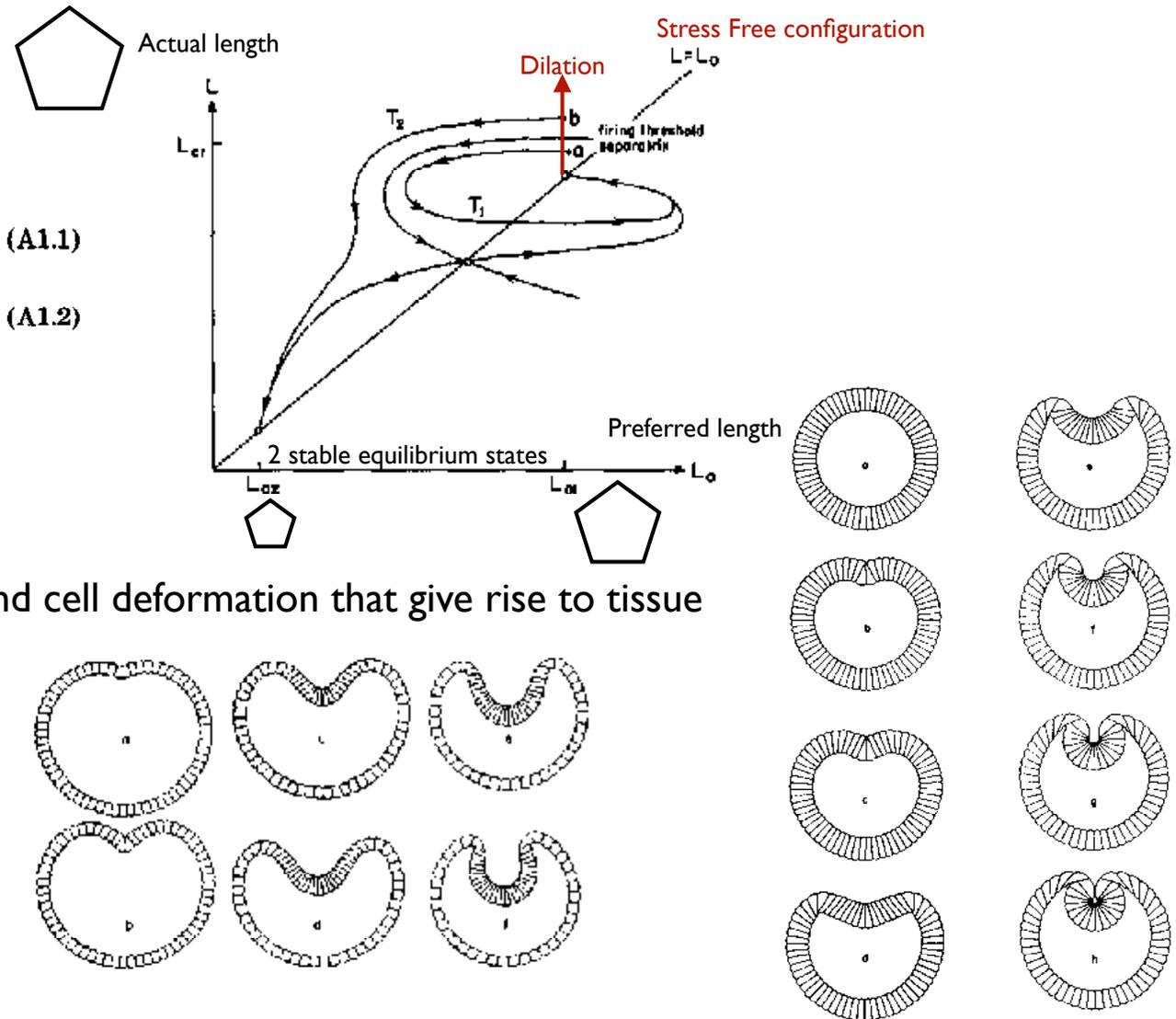


# Self-organised tissue invagination

- Rest configuration is a function of cell deformation

$$\frac{dL}{dt} = -\frac{k(\cdot)}{\mu} [L - L_0] \quad (\text{A1.1})$$

$$\frac{dL_0}{dt} = G(L, L_0) \quad (\text{A1.2})$$



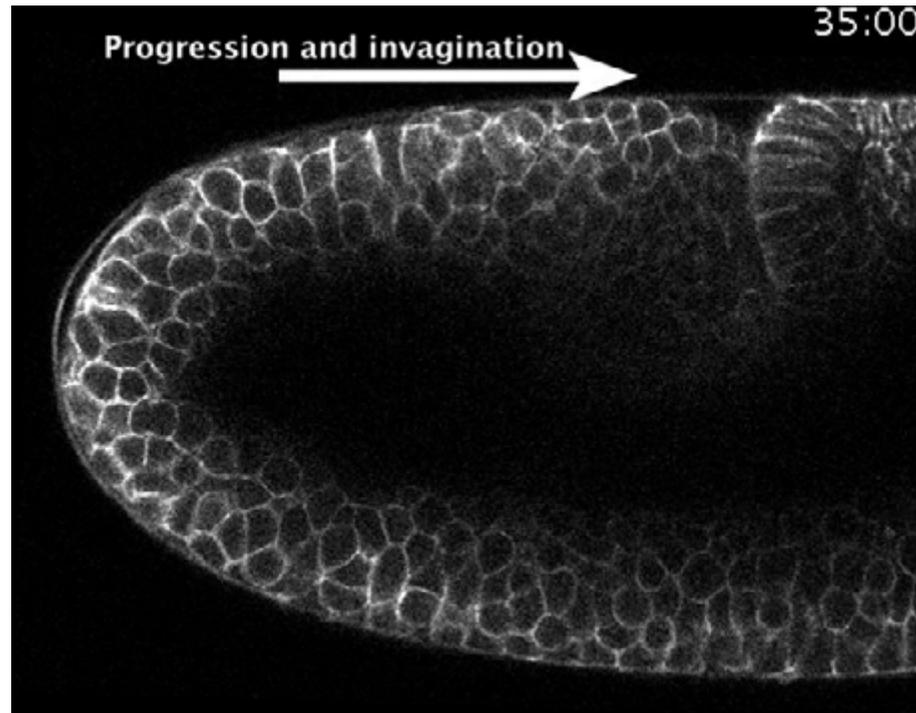
- Predicts wave of contractility and cell deformation that give rise to tissue folding and invagination



# Programmed and Self-Organised Invagination

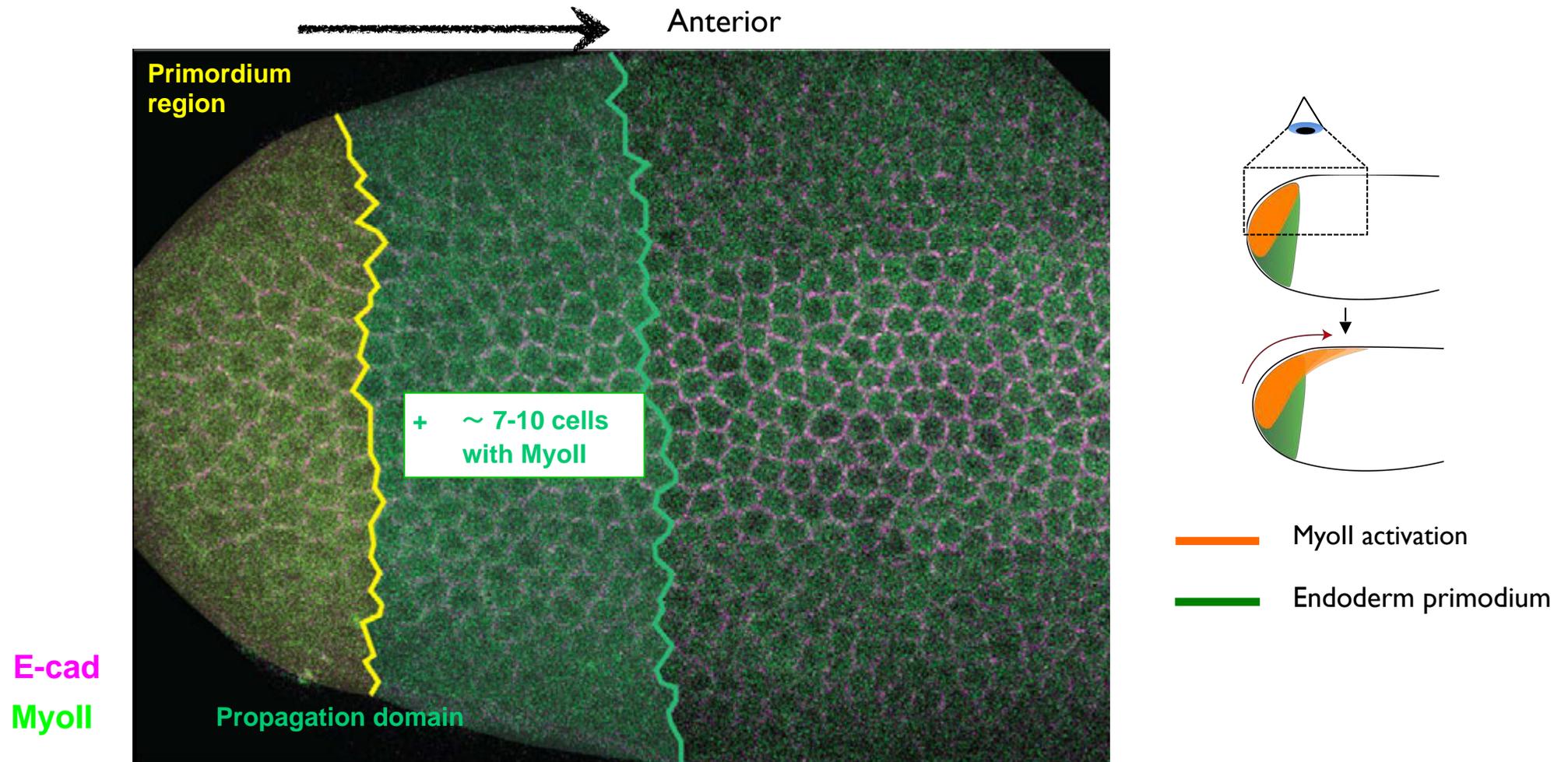
- Interplay between control and self-organisation of tissue morphogenesis

Endoderm morphogenesis in *Drosophila*



# Programmed and Self-Organised Invagination

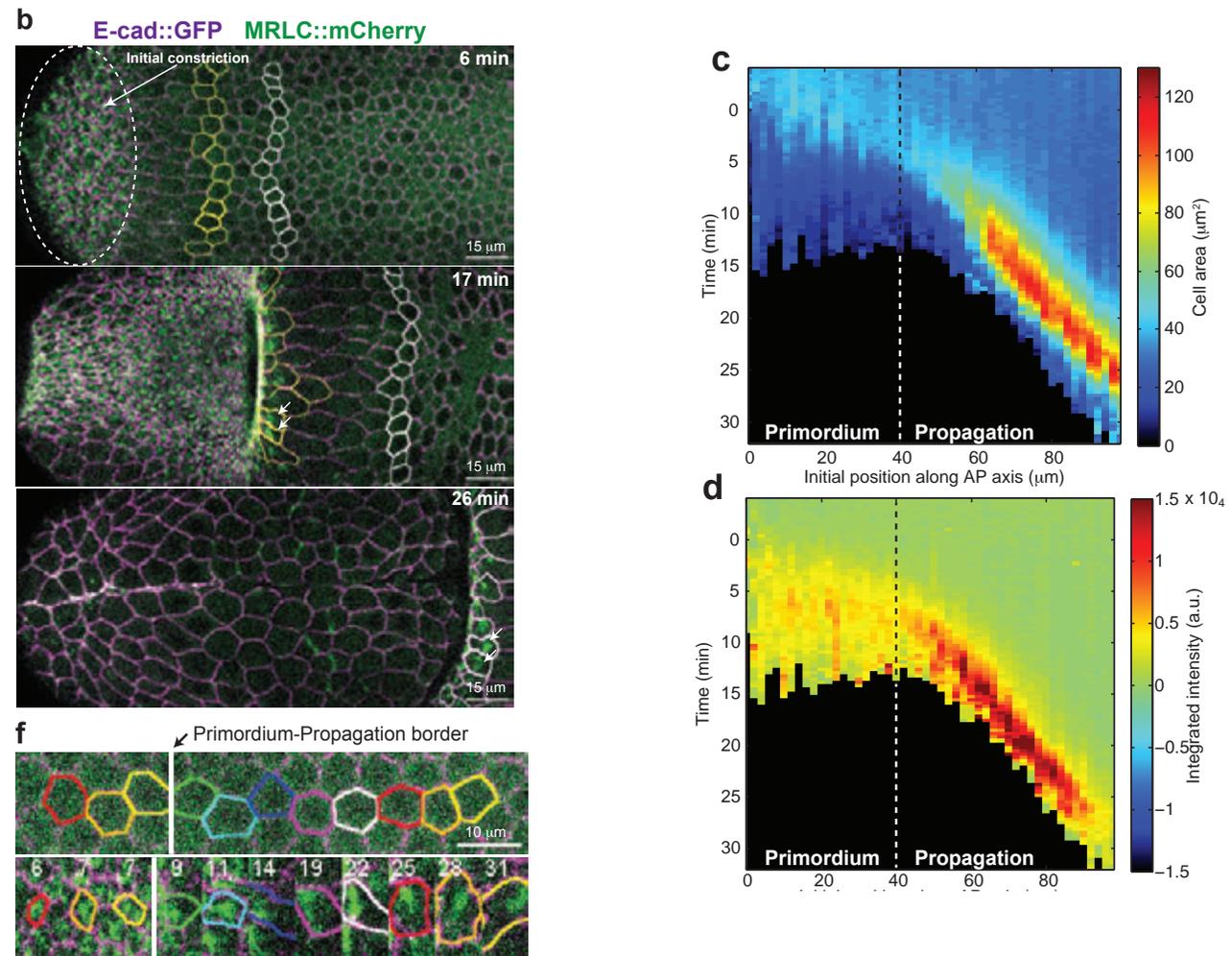
- A tissue scale wave of actomyosin activation is associated with wave of invagination



A. Bailles & C. Collinet et al, E. Munro and T. Lecuit. *bioRxiv* 430512; doi: <https://doi.org/10.1101/430512>

# Programmed and Self-Organised Invagination

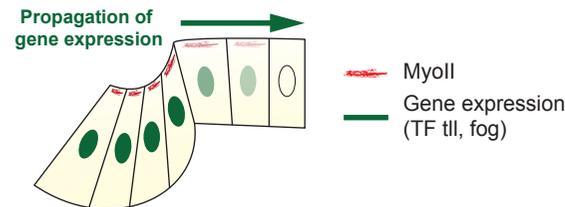
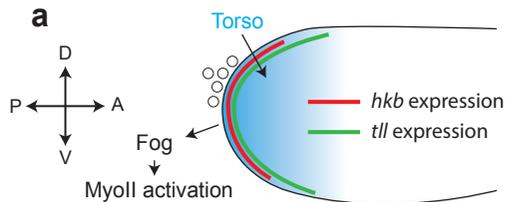
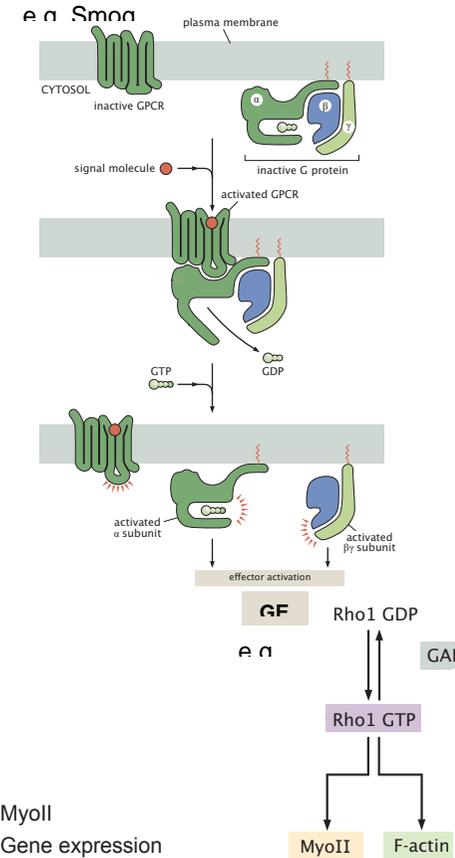
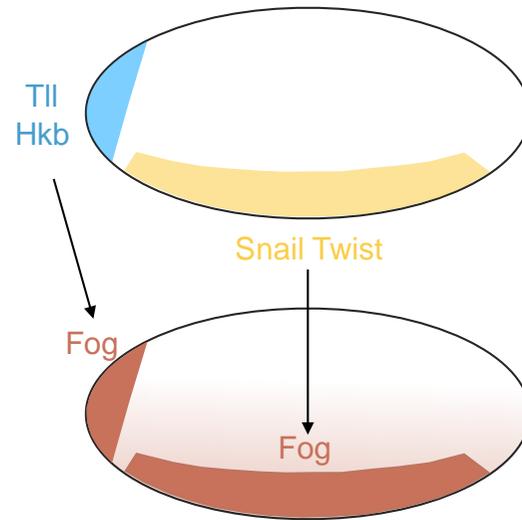
- A tissue scale wave of actomyosin activation is associated with wave of invagination



A. Bailles & C. Collinet et al, E. Munro and T. Lecuit. *bioRxiv* 430512; doi: <https://doi.org/10.1101/430512>

# Programmed and Self-Organised Invagination

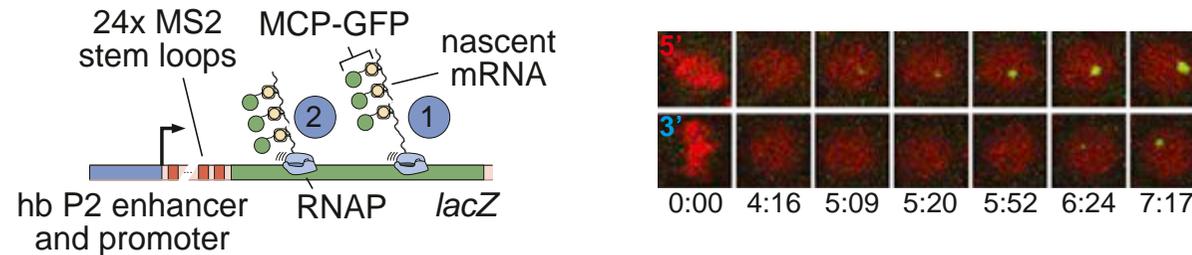
- Initiation of invagination: the transcriptional program



A. Bailles & C. Collinet et al, E. Munro and T. Lecuit. *bioRxiv* 430512; doi: <https://doi.org/10.1101/430512>

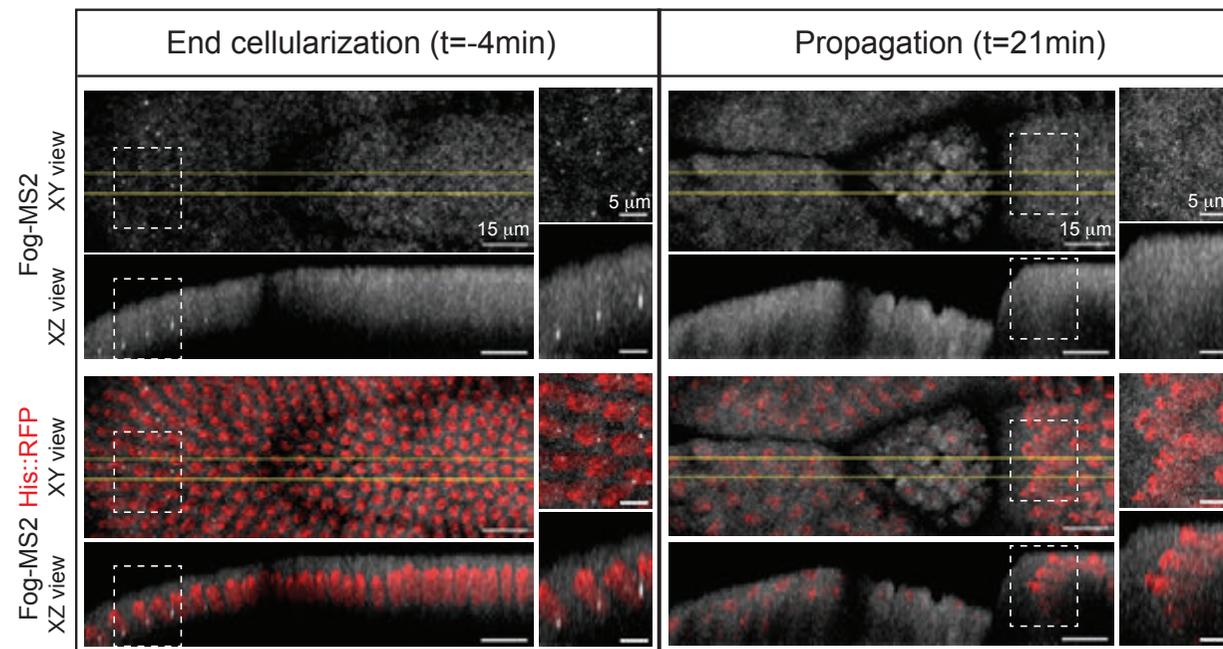
# Programmed and Self-Organised Invagination

- The tissue wave of invagination is not controlled transcriptionally



H. Garcia et al., 2013

Monitoring the dynamics of transcription in vivo with the MS2/MCP::GFP system

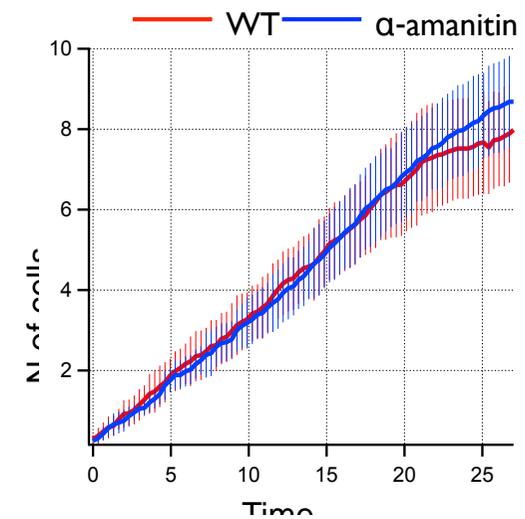
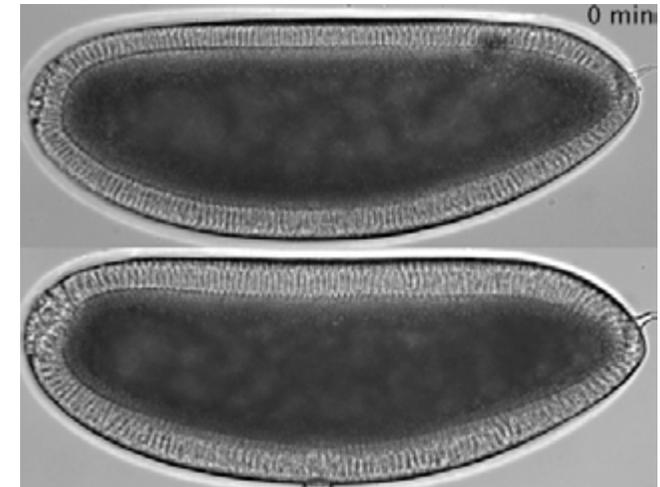


A. Bailles & C. Collinet et al, E. Munro and T. Lecuit. *bioRxiv* 430512; doi: <https://doi.org/10.1101/430512>

# Programmed and Self-Organised Invagination

- The tissue wave of invagination is not controlled transcriptionally

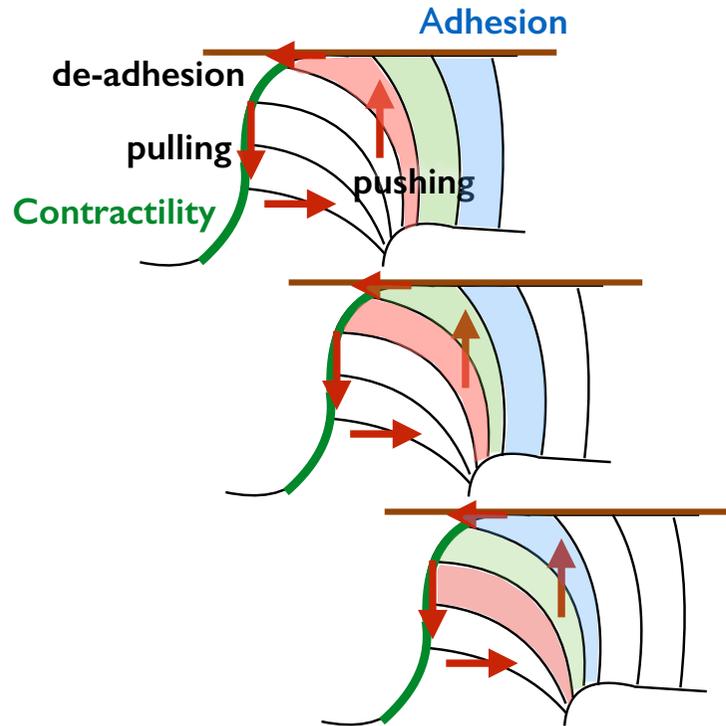
▶  $\alpha$ -amanitin is potent inhibitor of RNA Pol-II



A. Bailles & C. Collinet et al, E. Munro and T. Lecuit. *bioRxiv* 430512; doi: <https://doi.org/10.1101/430512>

# Tissue scale Model:

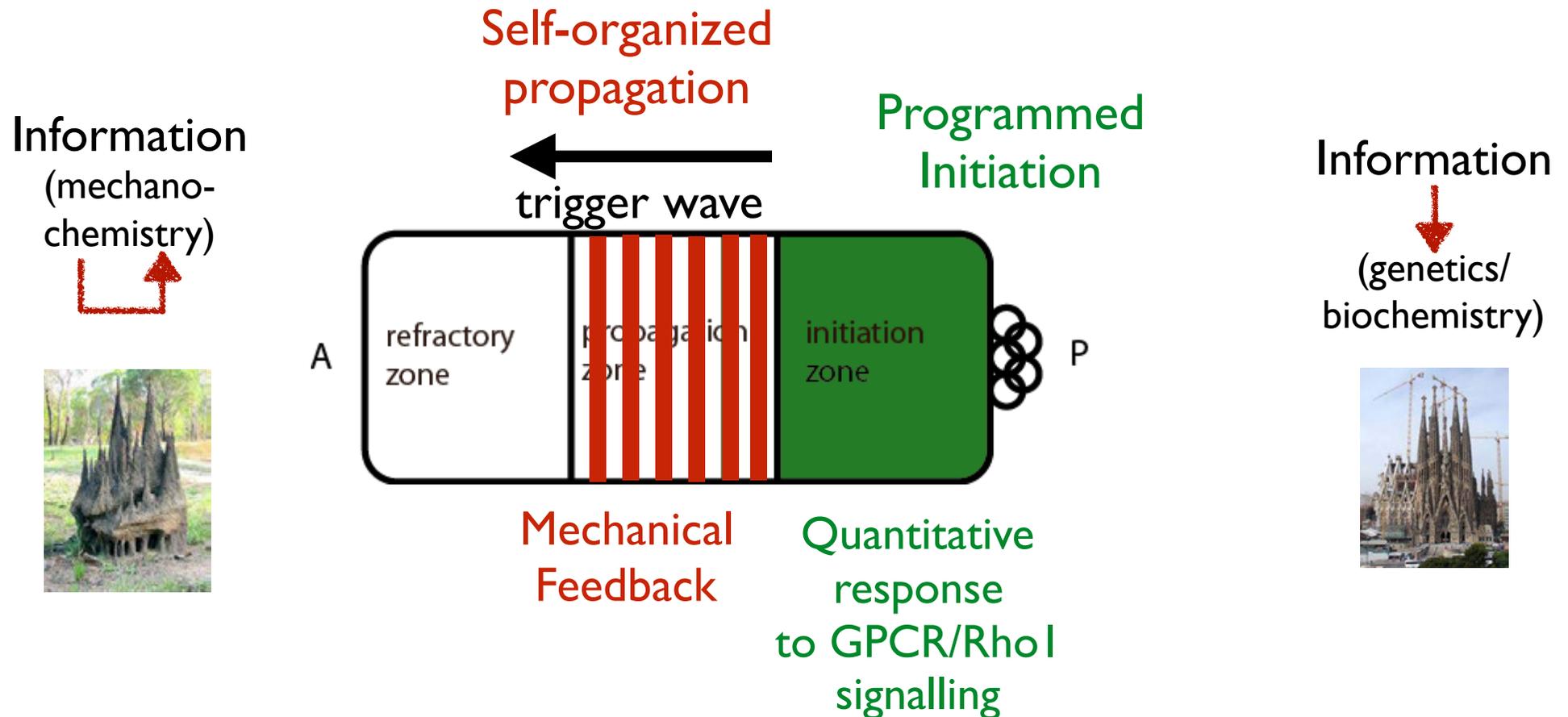
## Self-propagating mechanical cycle



- Invagination compresses cells anteriorly and against vitelline membrane
- Cells adhere to vitelline membrane and subsequently detach as they join invagination
- New cycle begins

A. Bailles & C. Collinet et al, E. Munro and T. Lecuit. *bioRxiv* 430512; doi: <https://doi.org/10.1101/430512>

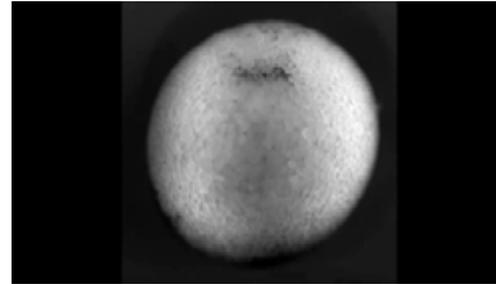
# Self-organisation with mechano-chemical information



A. Bailles & C. Collinet et al, E. Munro and T. Lecuit. *bioRxiv* 430512; doi: <https://doi.org/10.1101/430512>

# Next Course 18 December

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## Tissue extension and growth: control and self-organisation

