

**Cours du 17-10-2011**

# The genetics of early telencephalon patterning: some assembly required

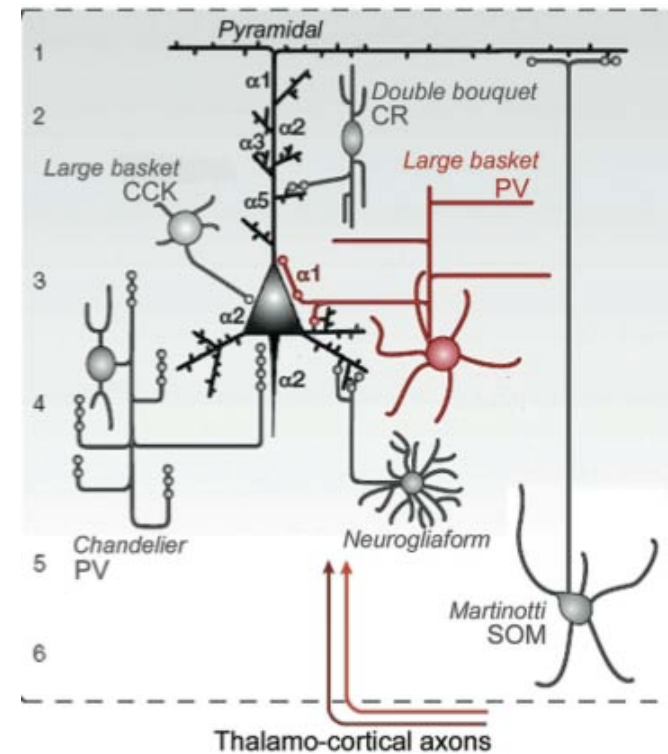
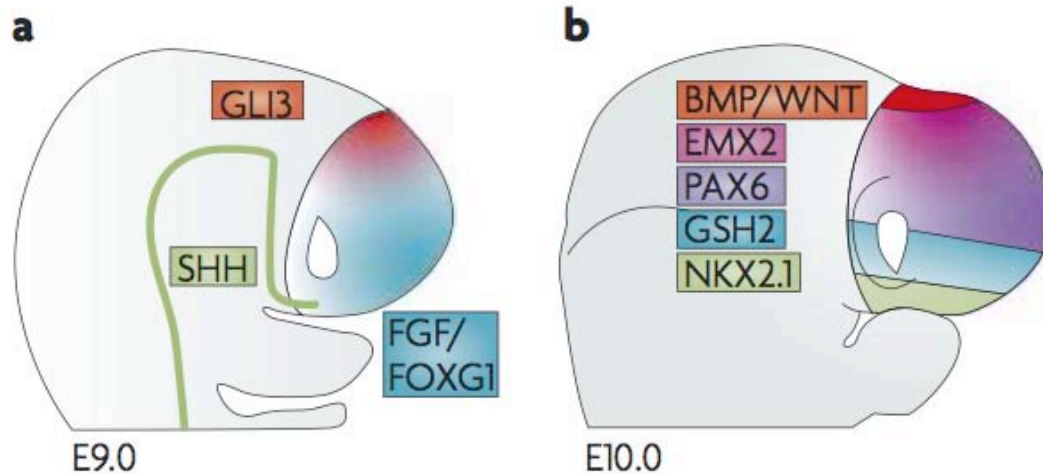
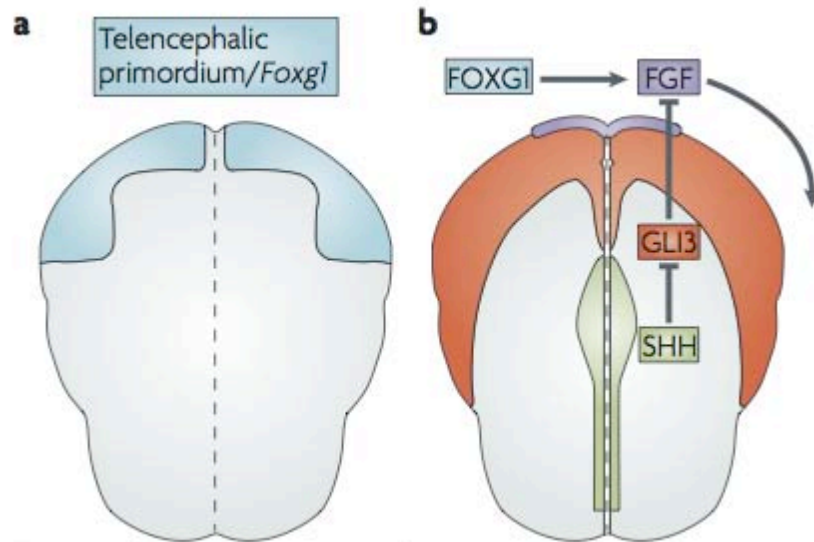
Hábert I, Fishell C

## From brain formation to plasticity: insights on Otx2 homeoprotein

Sugiyama S, Prochiantz A, Hensch T

Development, Growth & Differentiation

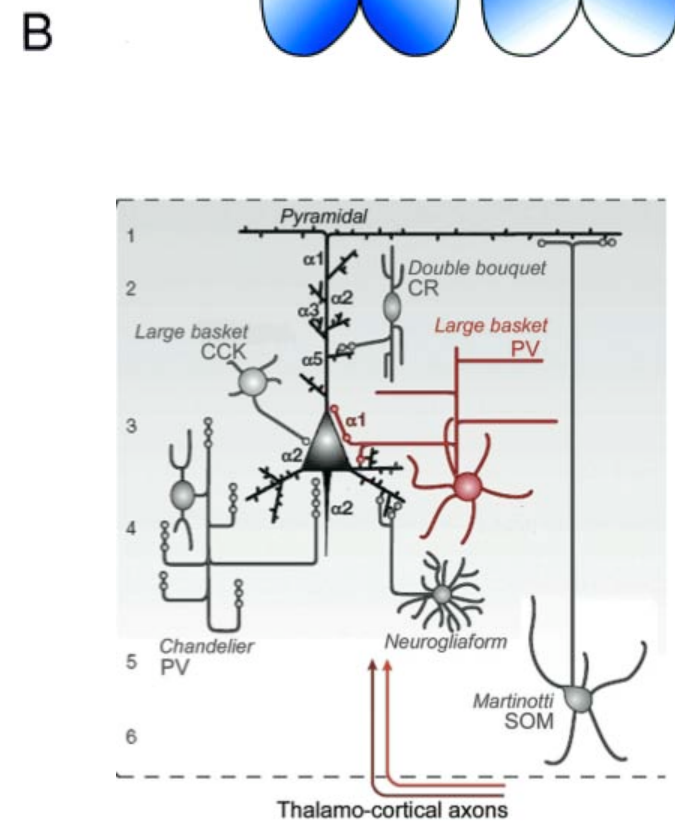
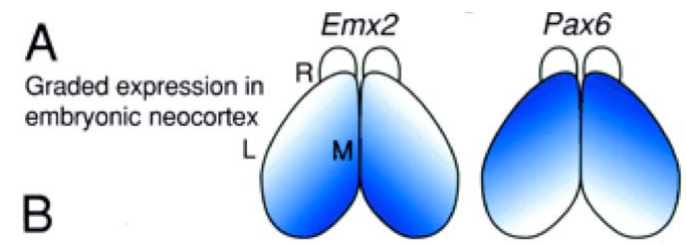
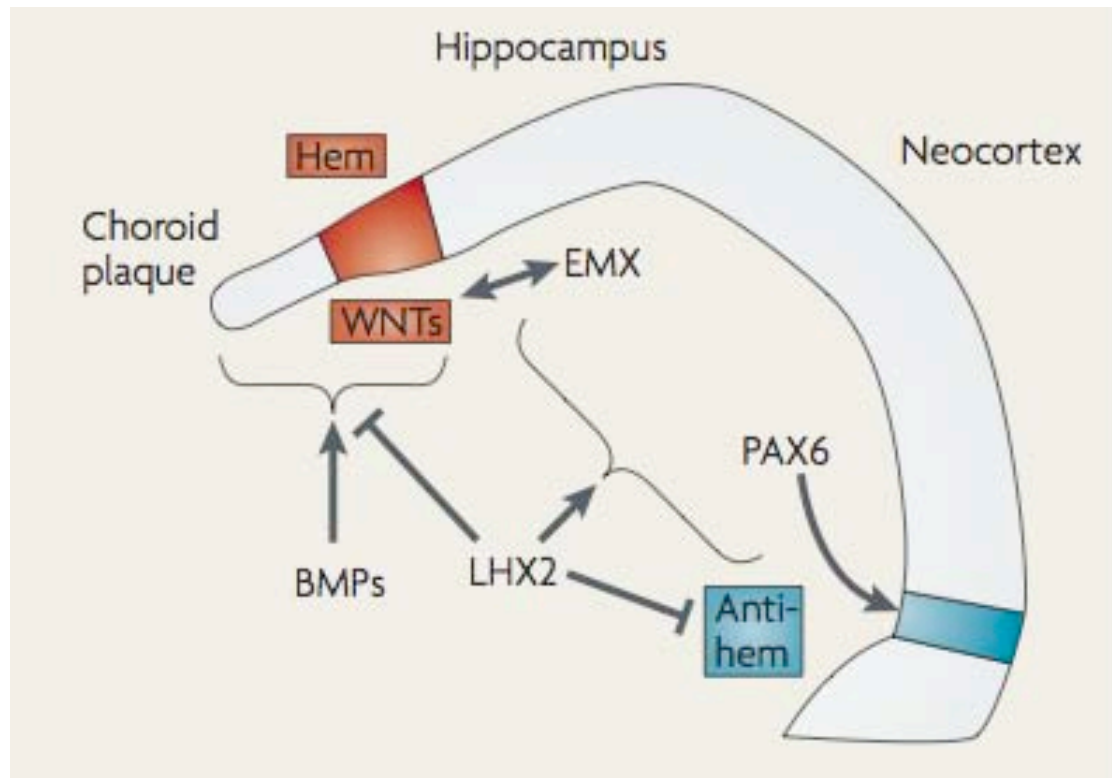
2009 vol. 51 (3) pp. 369-77



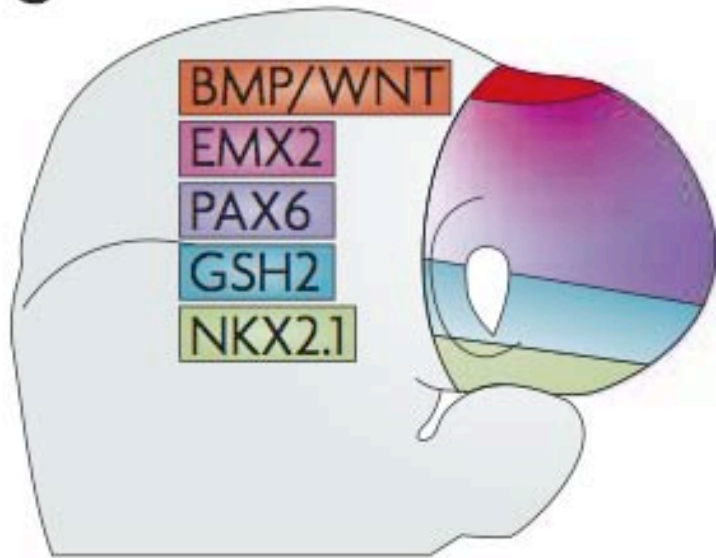
# The genetics of early telencephalon patterning: some assembly required

Hébert J, Fishell G

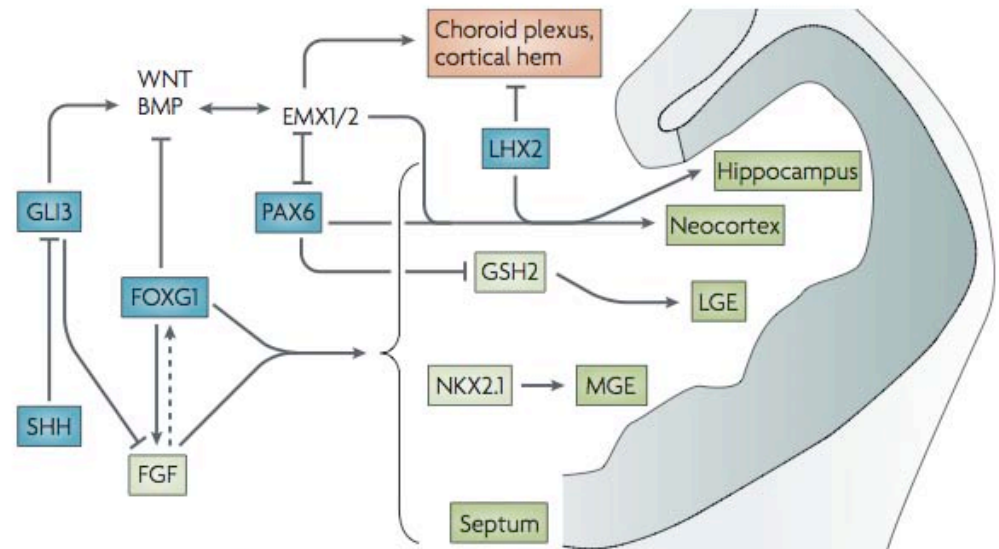
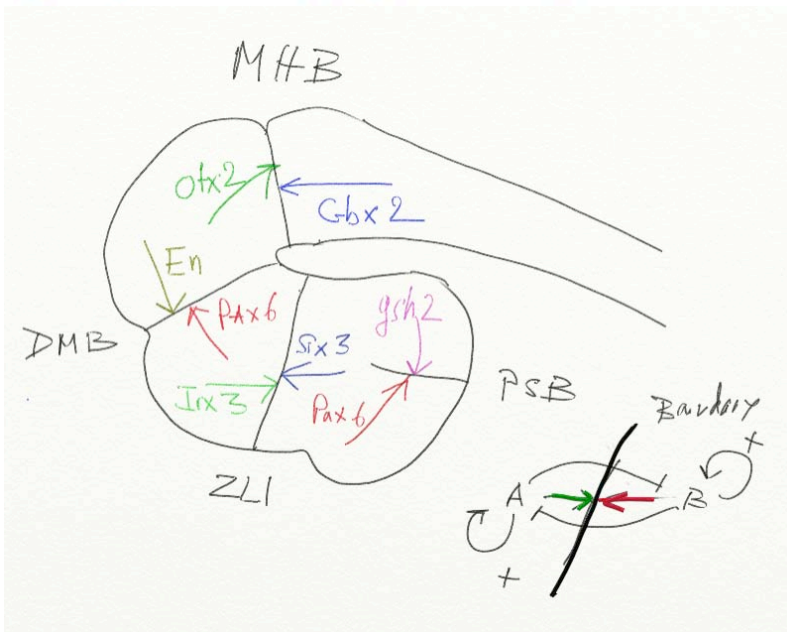
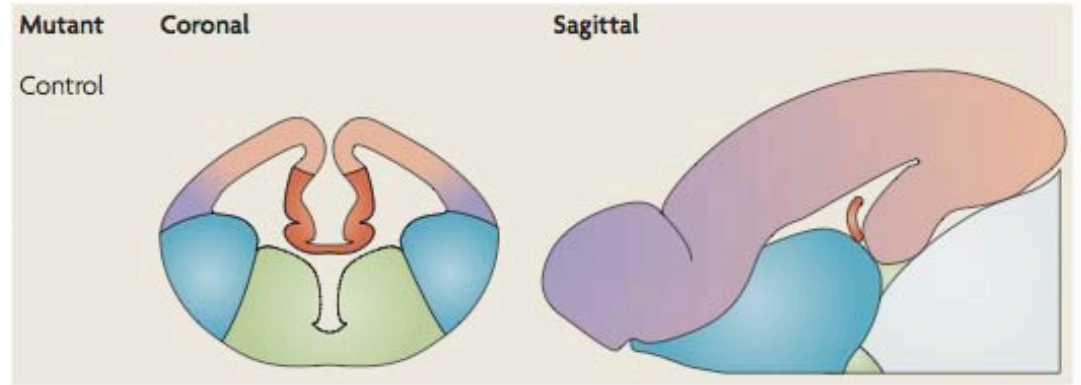
Nat Rev Neurosci  
2008 vol. 9 (9) pp. 678-85



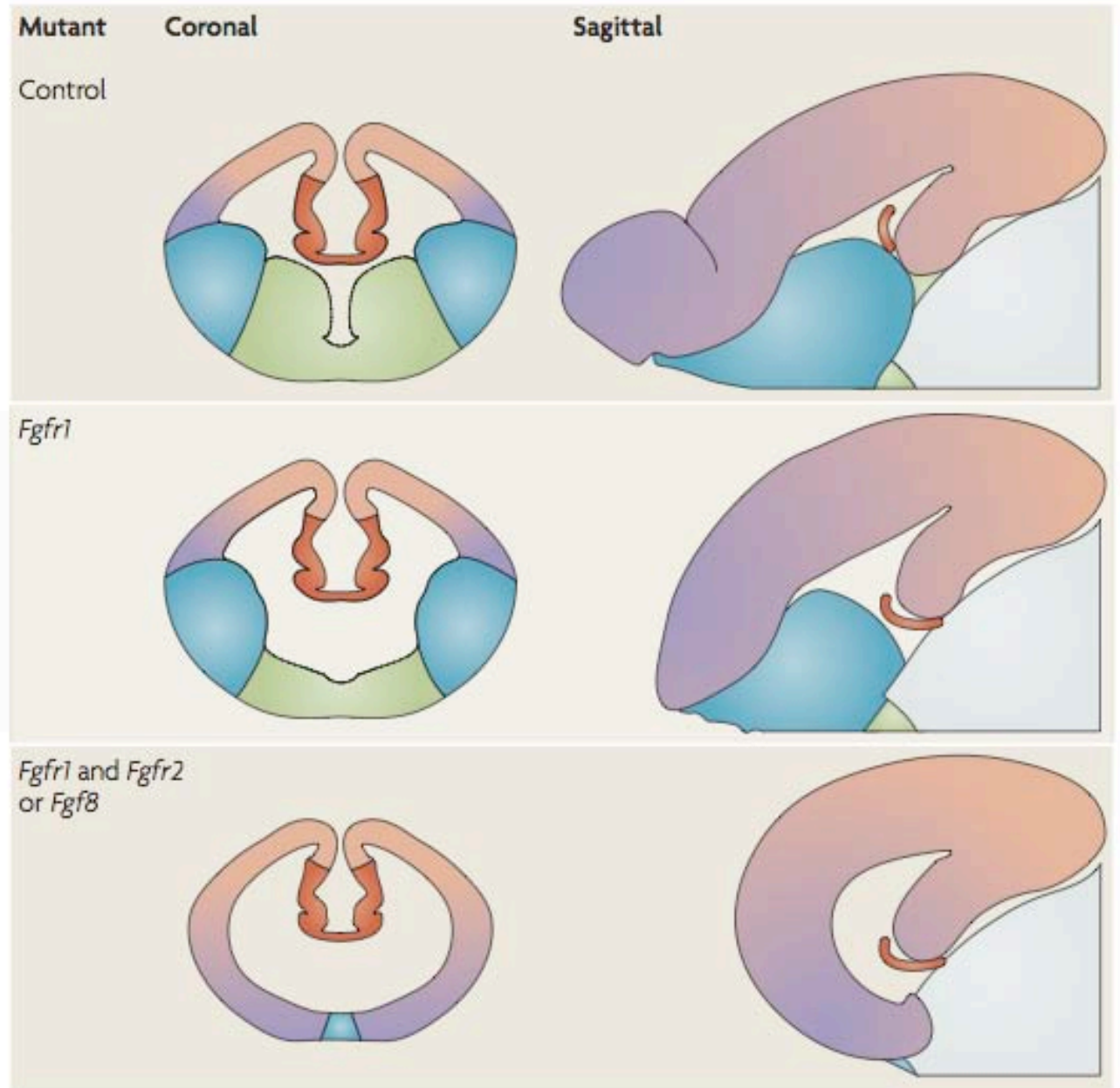
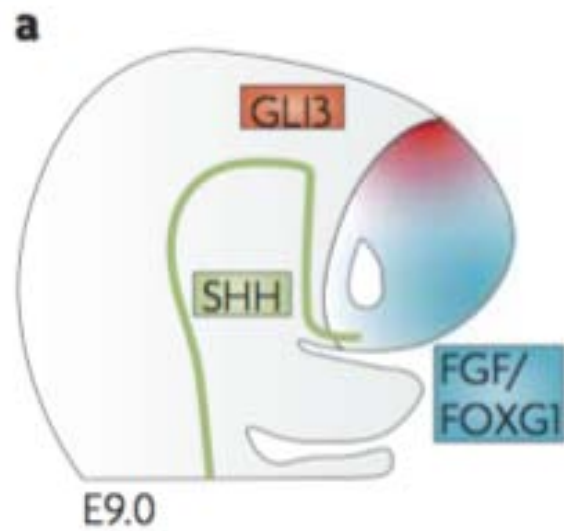
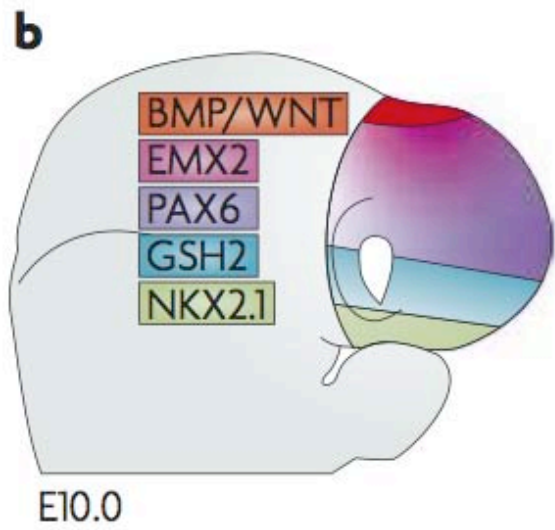
**b**



E10.0



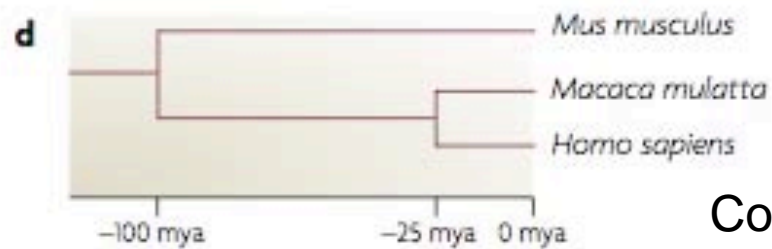
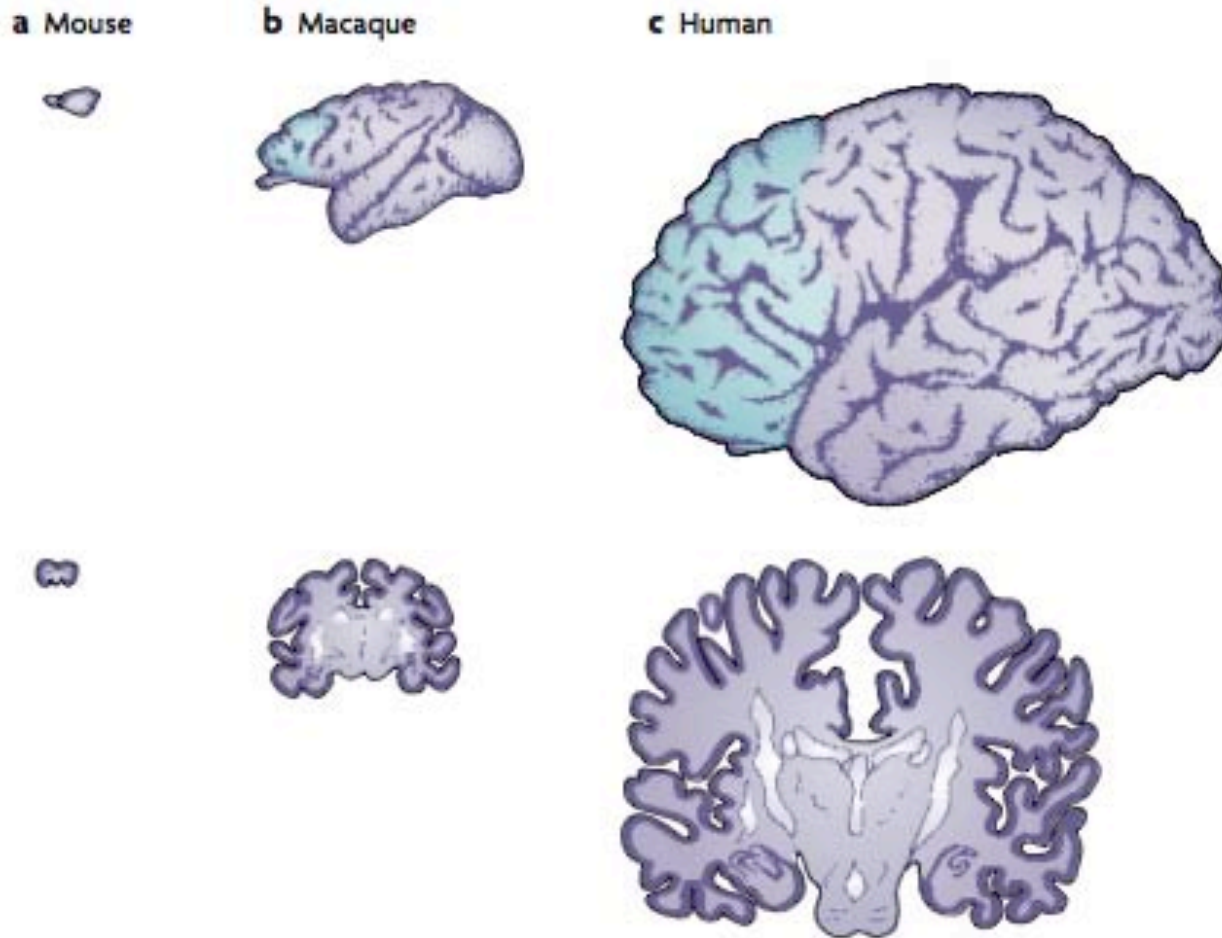




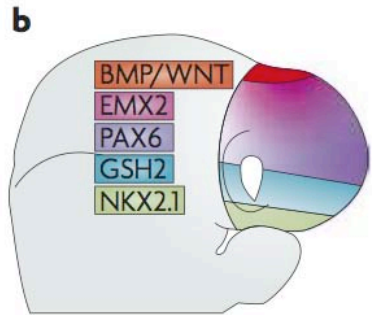
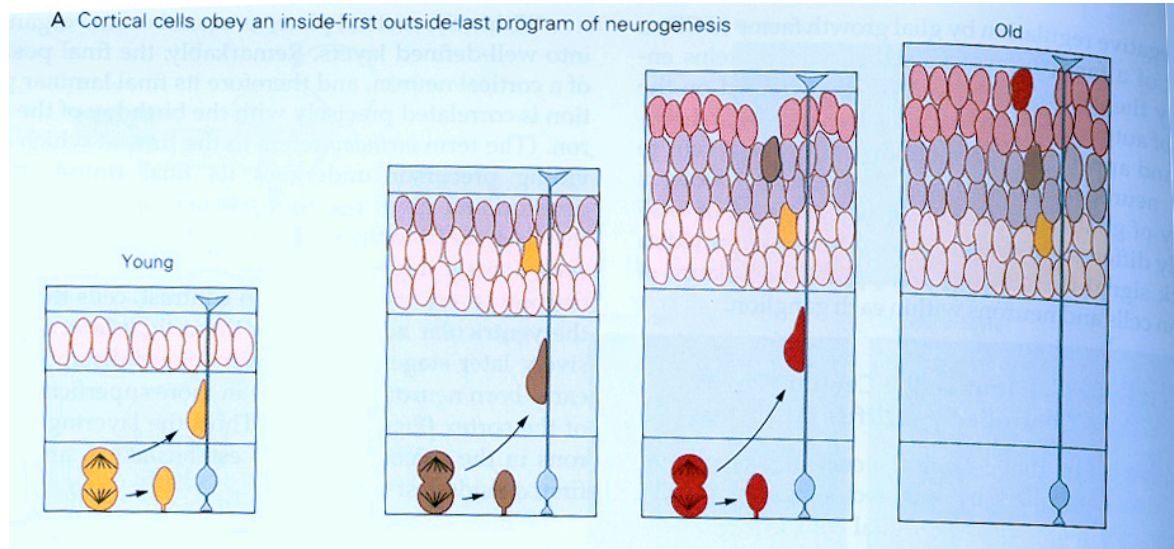
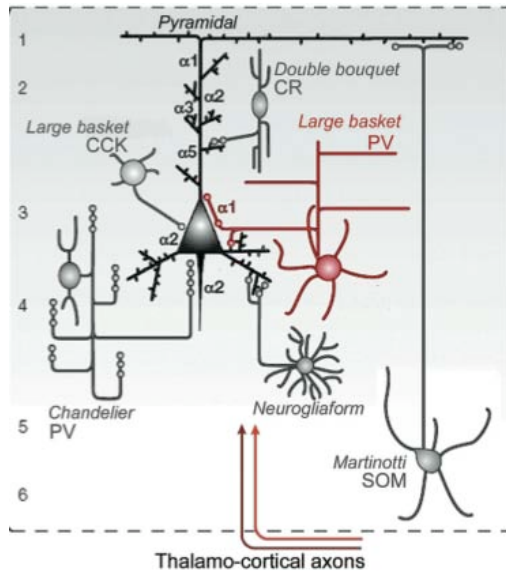
# Evolution of the neocortex: a perspective from developmental biology

Rakic P

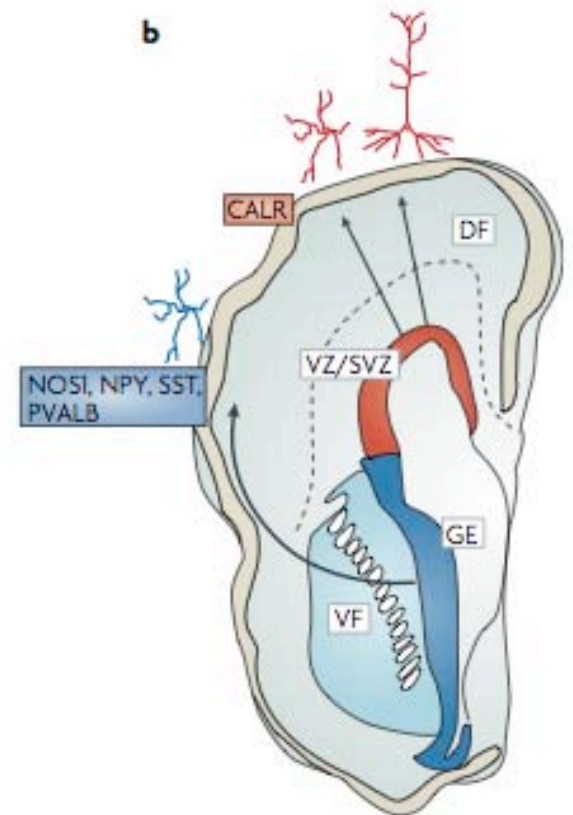
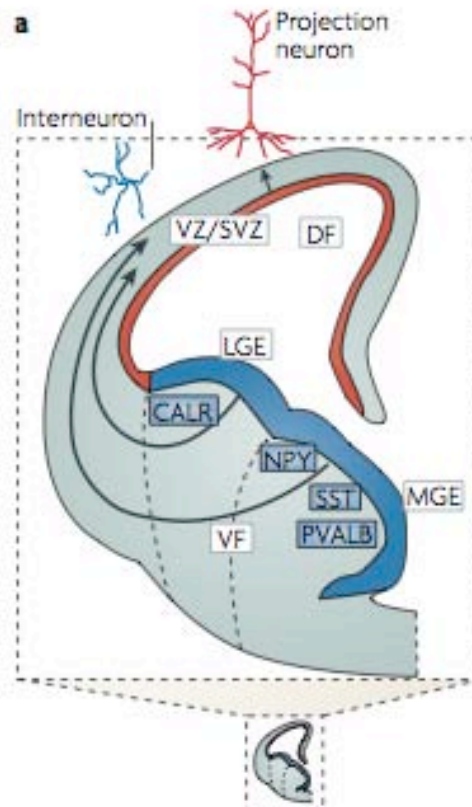
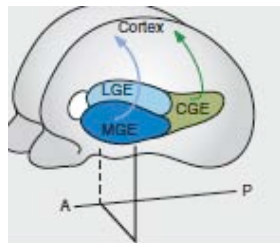
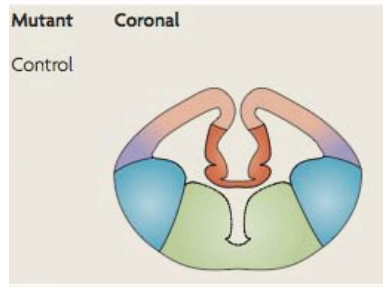
Nat Rev Neurosci  
2009 vol. 10 (10) pp. 724-35



Cortex préfrontal en bleu, absent chez la souris



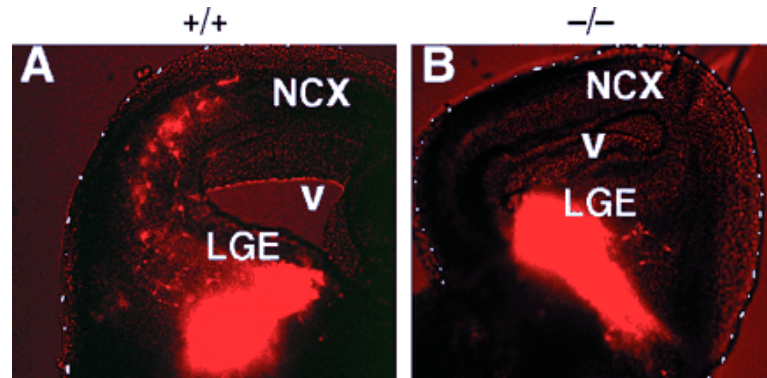
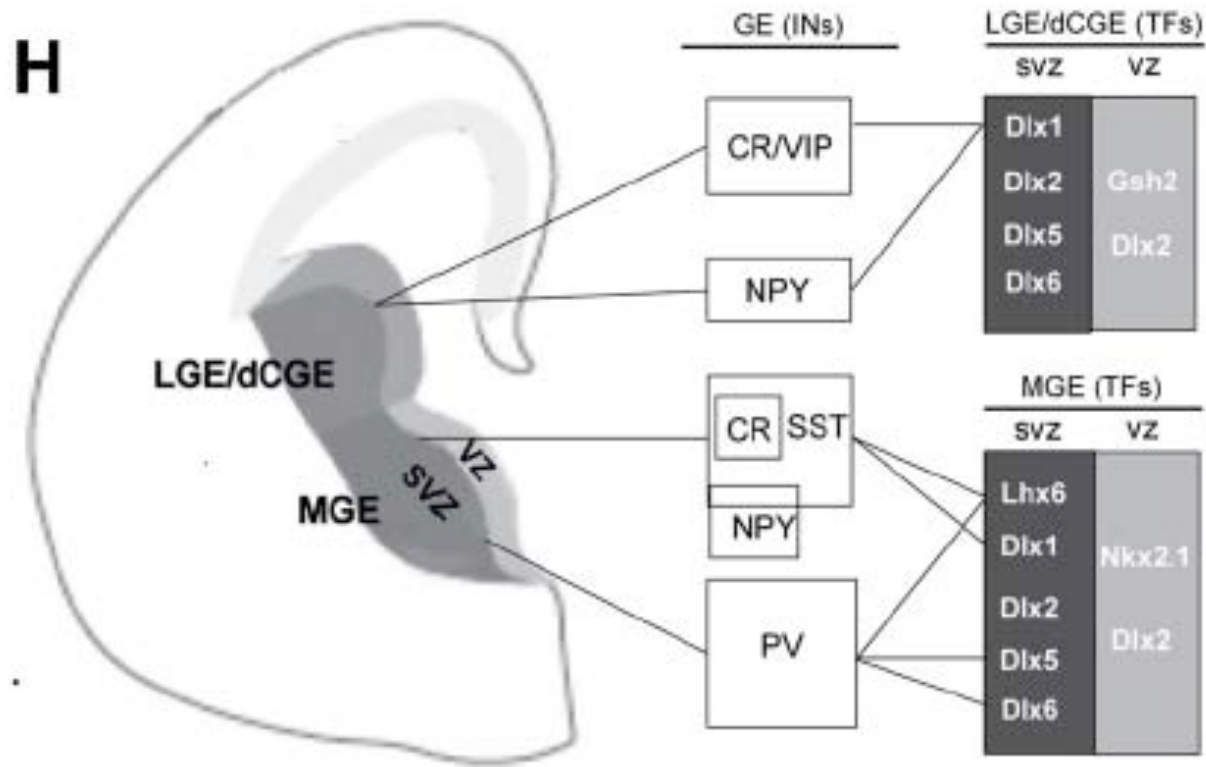
E10.0





# *Dlx5* and *Dlx6* Regulate the Development of Parvalbumin-Expressing Cortical Interneurons

Yanling Wang,<sup>1\*</sup> Catherine A. Dye,<sup>1,2\*</sup> Vikaas Sohal,<sup>7</sup> Jason E. Long,<sup>1,3</sup> Rosanne C. Estrada,<sup>4</sup> Tomas Roztocil,<sup>5</sup> Thomas Lufkin,<sup>6</sup> Karl Deisseroth,<sup>7</sup> Scott C. Baraban,<sup>4</sup> and John L. R. Rubenstein<sup>1</sup>

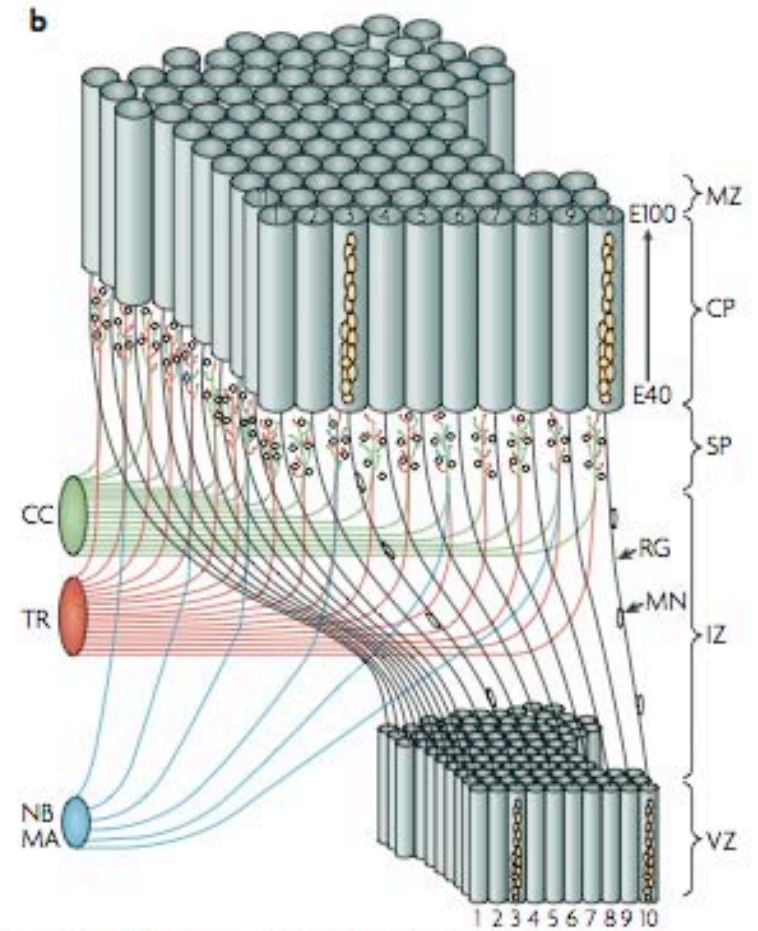
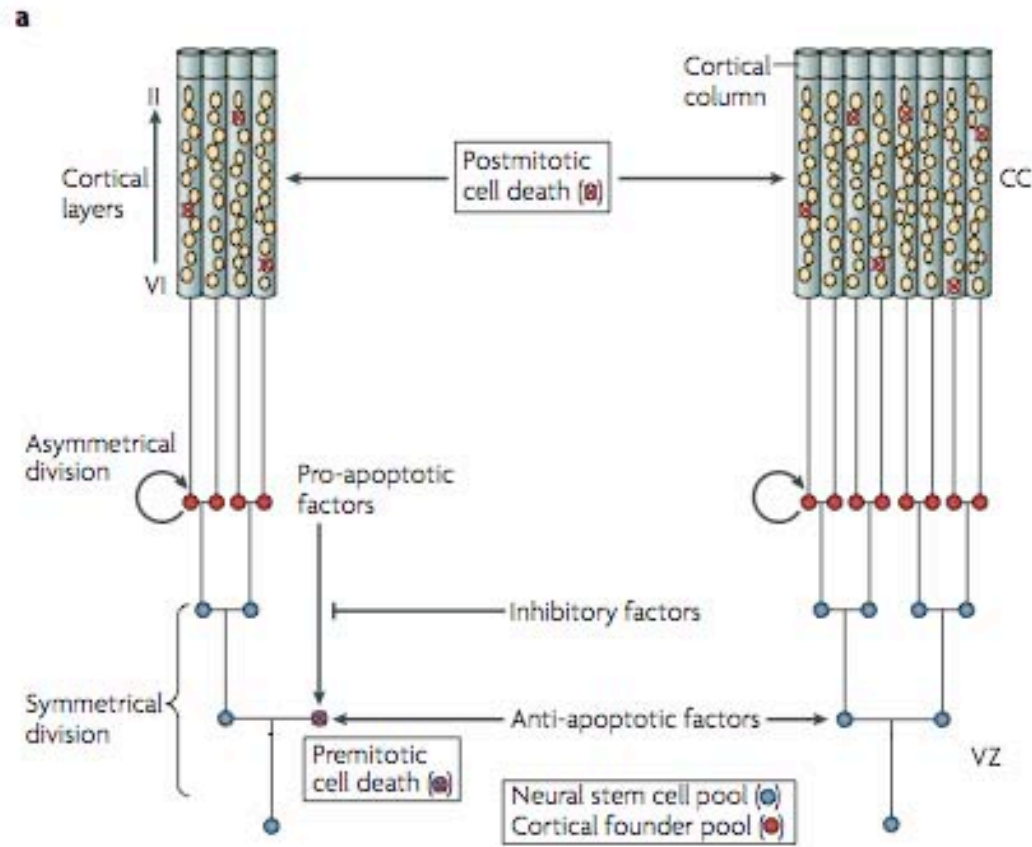


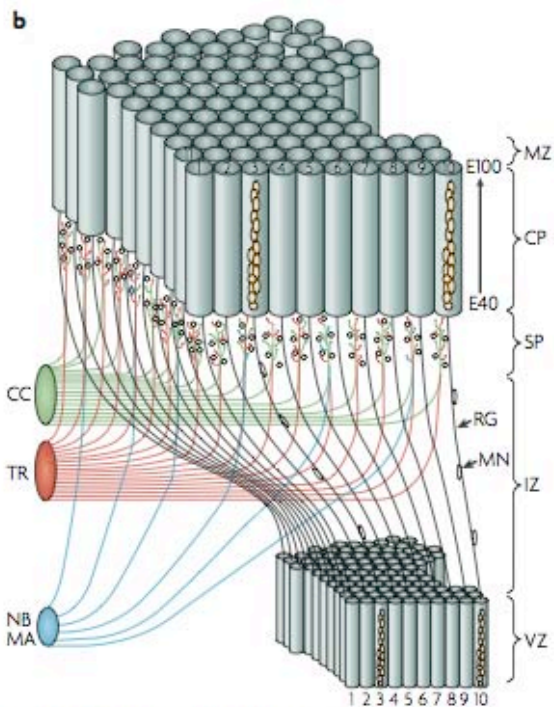
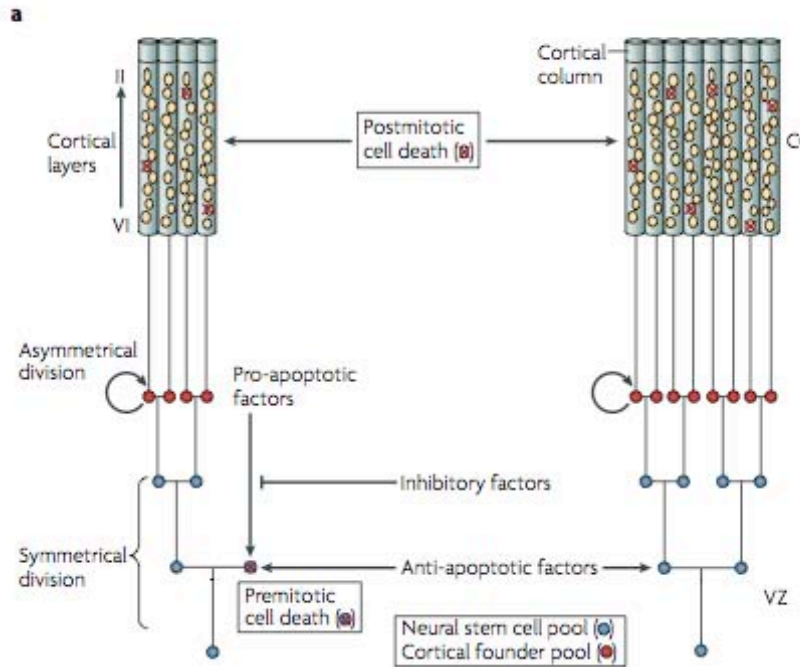


# Evolution of the neocortex: a perspective from developmental biology

Rakic P

Nat Rev Neurosci  
2009 vol. 10 (10) pp. 724-35



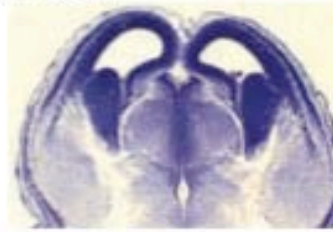


## Evolution of the neocortex: a perspective from developmental biology

Rakic P

Nat Rev Neurosci  
2009 vol. 10 (10) pp. 724-35

**a Wild type**



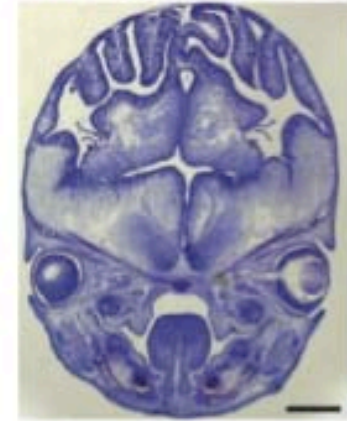
**Caspase 9 knockout**



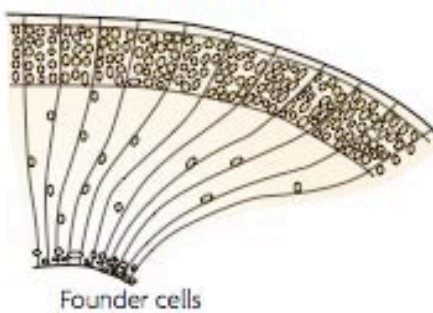
**b Wild type**



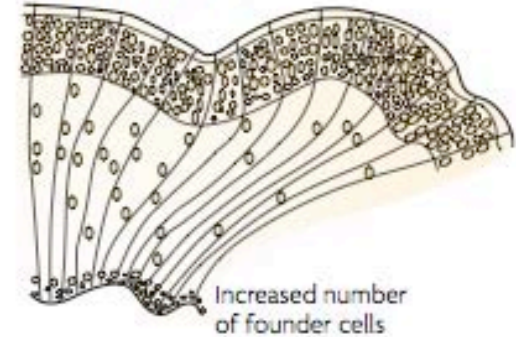
**$\Delta 90\beta$ -catenin-GFP**



**c Lissencephalic cerebrum**



**Convolutated cerebrum**





# Postnatal Hippocampal Neurogenesis in Rats <sup>1</sup>

J. COMP. NEUR., 124: 319-336.

JOSEPH ALTMAN AND GOPAL D. DAS

Psychophysiological Laboratory, Massachusetts Institute of Technology,  
Cambridge, Massachusetts

HIPPOCAMPAL NEUROGENESIS

321

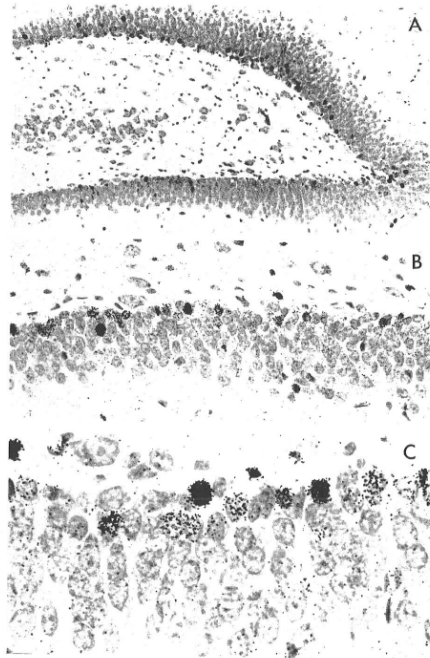


Fig. 1. Low and high power microphotographs of autoradiograms from the area of the dentate gyrus of the hippocampus in a rat injected with thymidine- $H^3$  at the age of ten days and killed two months after the injection. Note labeling of granule cells, predominantly in the internal border (basal surface) of the granular layer. A, 100 $\times$ ; B, 250 $\times$ ; C, 640 $\times$ .

Our interpretation of these results, namely that the granule cells of the dentate gyrus multiply at a high rate in young animals, a process which declines rapidly but does not cease altogether in the mature animals, is difficult to reconcile with the absence of signs of mitotic activity in differentiated granule cells. We have, accordingly (un-

dertaken to investigate the possibility that undifferentiated precursor cells take up thymidine, multiply, and subsequently differentiate into granule cells. This possibility was suggested to us by the presence of small cells with darkly staining nuclei in the inner border or base of the granular layer, a region where most of the labeled cells were encountered. There is little information in the literature about the properties of these small cells with dark nuclei, presumably because they are considered to be glia cells (they are sometimes described as astrocytes). However, in mature animals neuroglia cells are seldom encountered in the dentate gyrus, whereas in young animals, as we shall describe in detail later, these cells with small dark nuclei are quite abundant. We have, therefore, postulated that these are undifferentiated cells which can take up thymidine and multiply, and may subsequently differentiate into granule cells. If this hypothesis were correct, then we should find a larger proportion of labeled small cells with dark nuclei in animals killed soon after injection with thymidine- $H^3$ , whereas in animals surviving for longer periods after injection the labeled small cells should decrease and the labeled differentiated cells should increase at the expense of the former. We could investigate this hypothesis in the

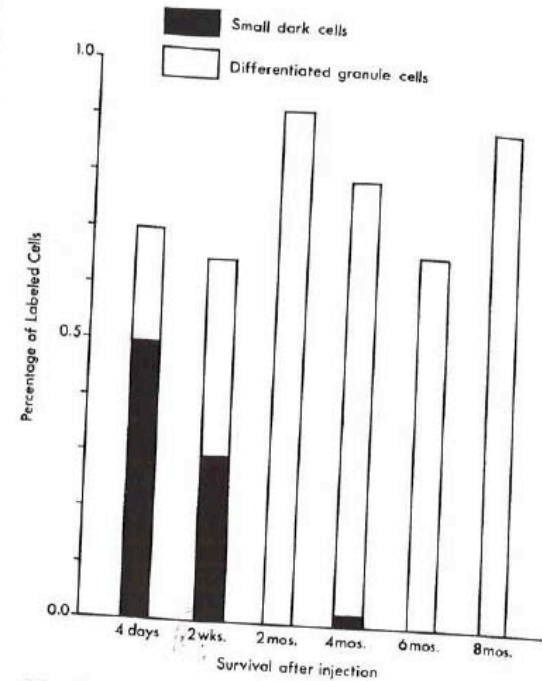
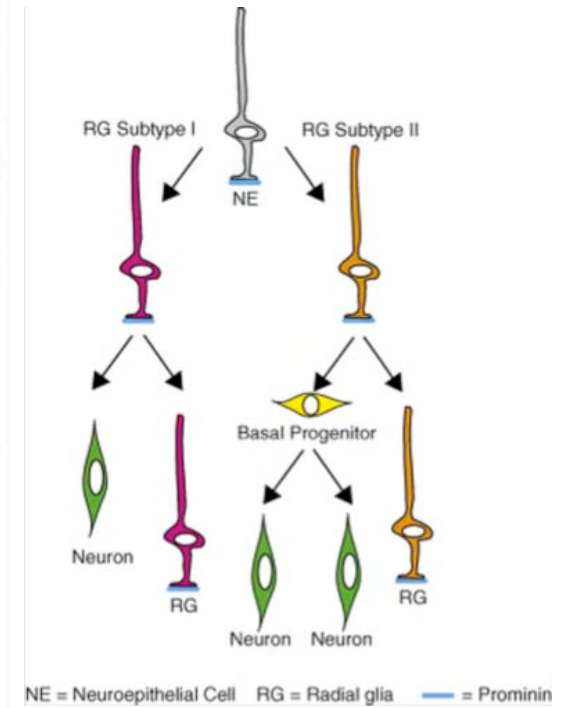
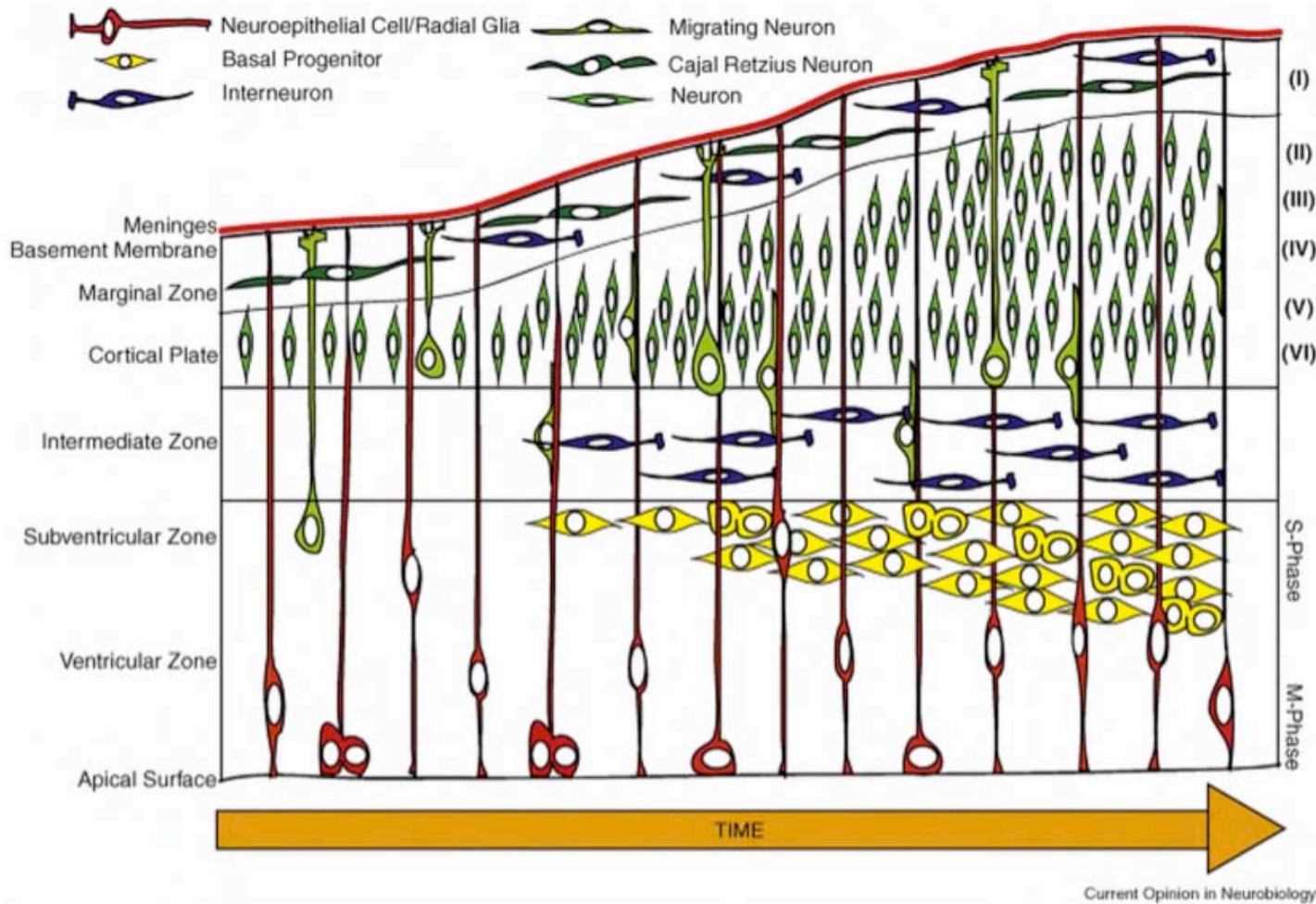


Fig. 5. Types of cells labeled in the granular layer in rats injected as adults (4 months), following which they survived for the periods indicated.

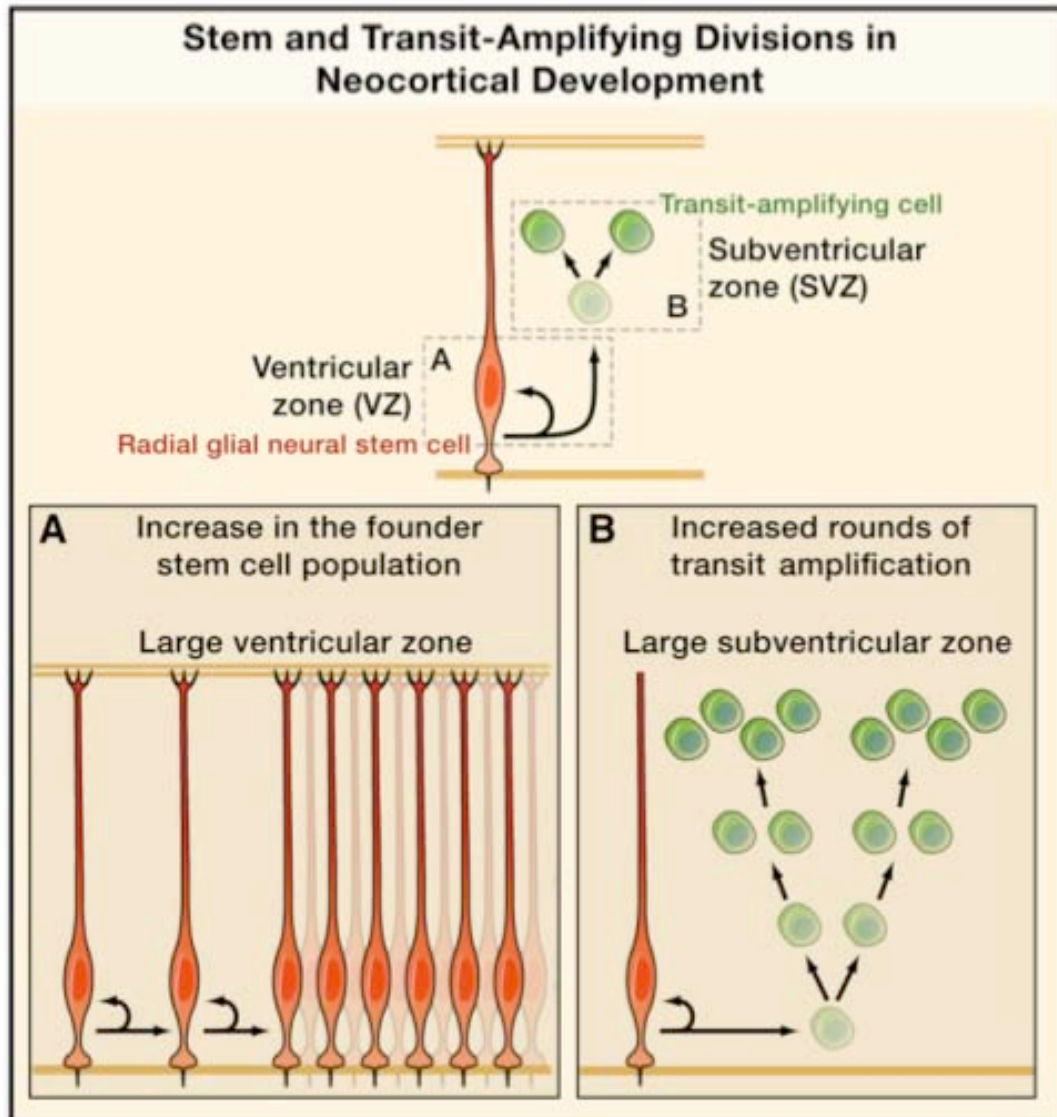
# Stem cells niches during development--lessons from the cerebral cortex

Johansson PA, Cappello S, Götz M

Current Opinion in Neurobiology  
2010 vol. 20 (4) pp. 400-7

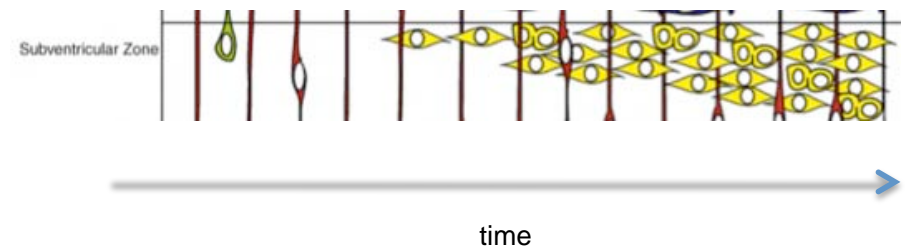


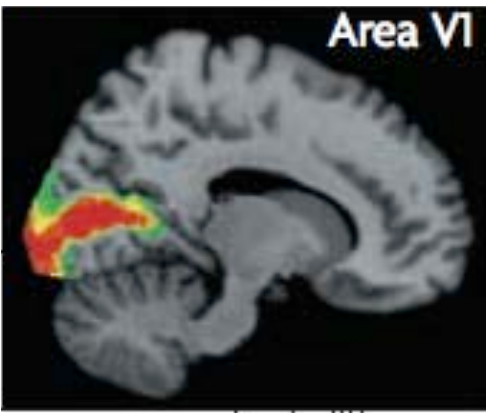
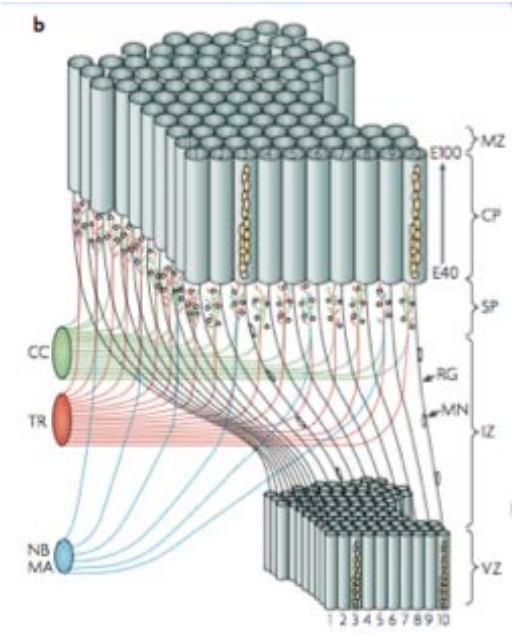
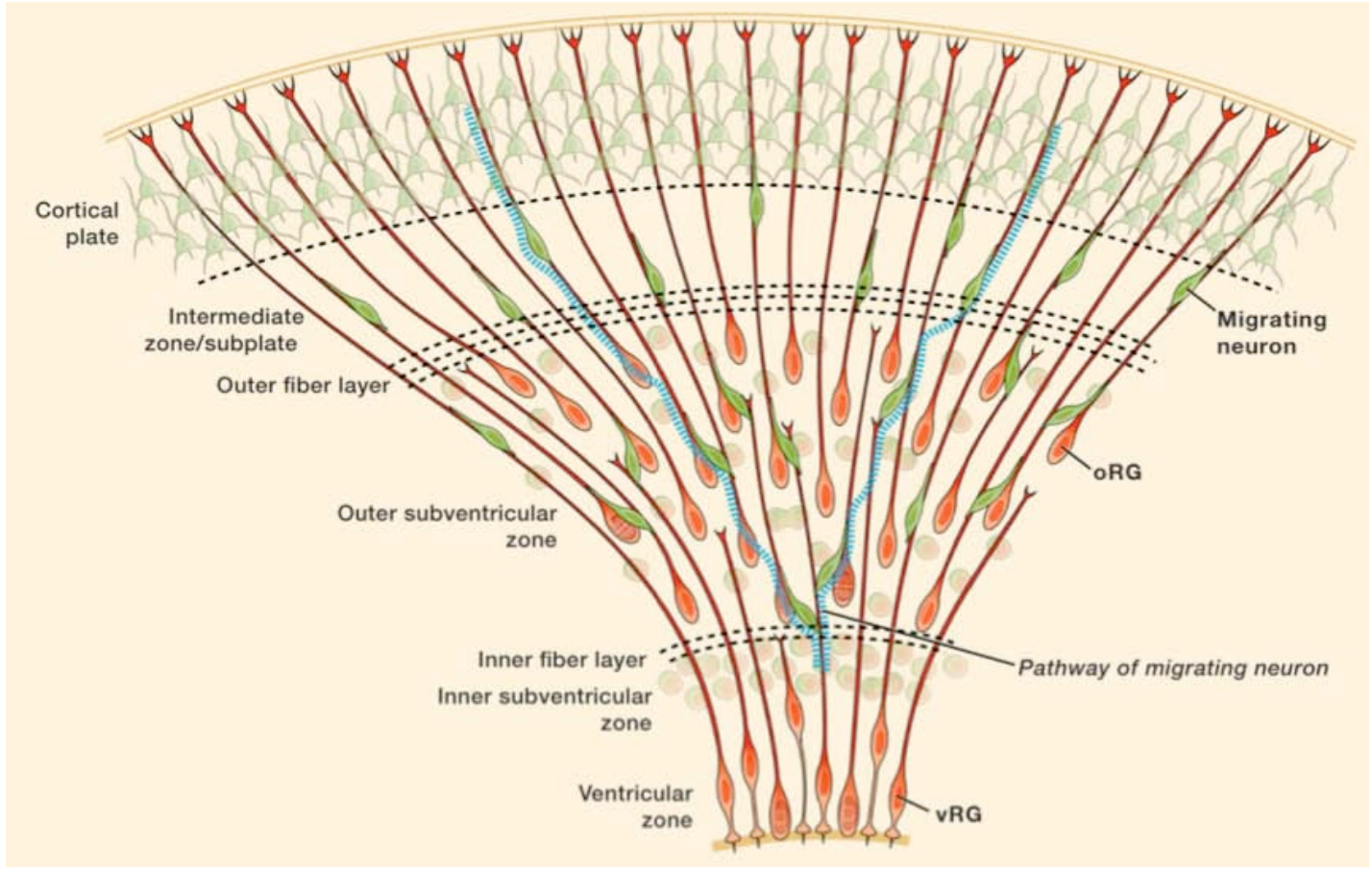




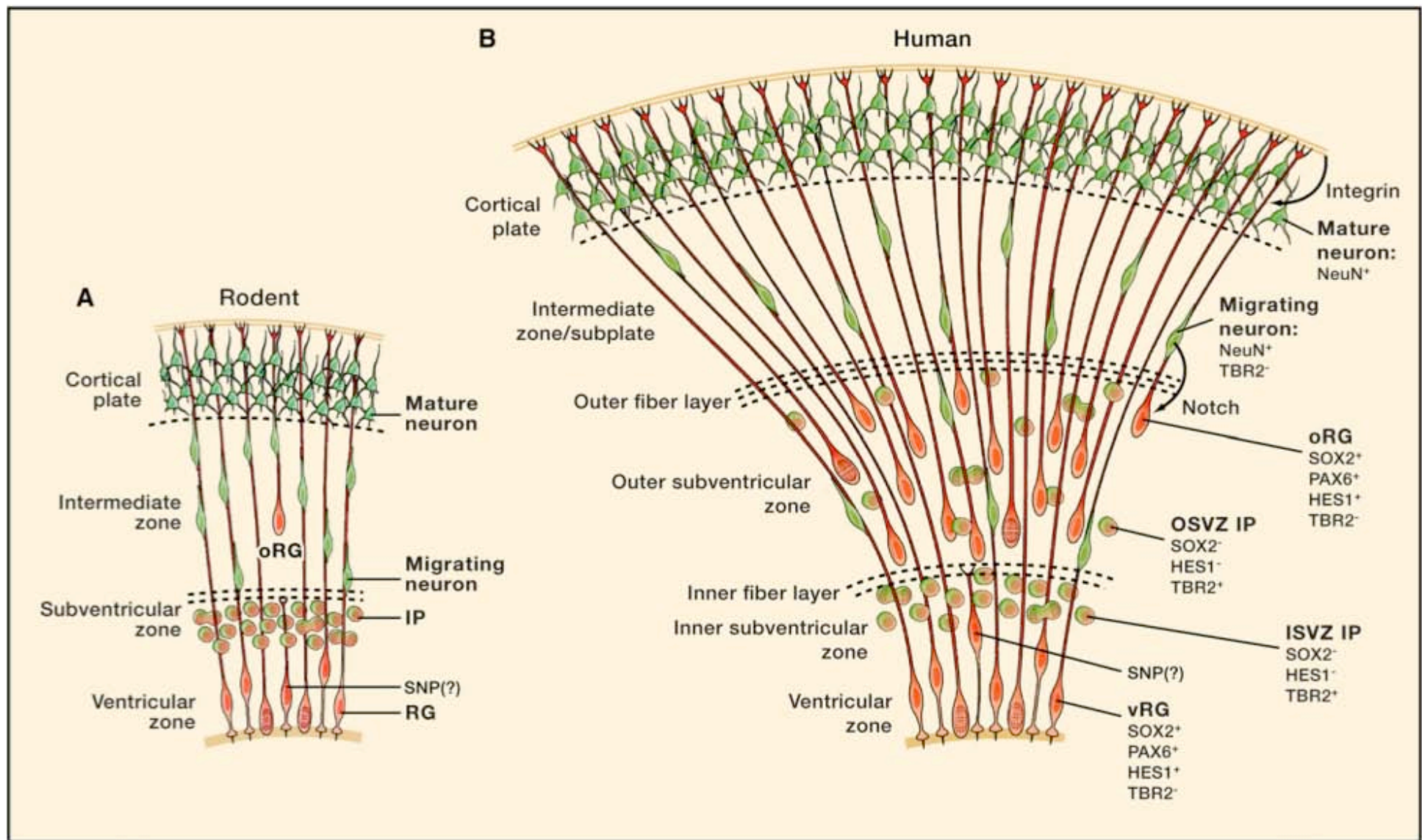
**Figure 1. Progenitor Cell Expansion Can Underlie Neocortical Enlargement**

Neuronal number is a key determinant of neocortex size and shape. Neurons are produced from a lineage of radial glia (RG) stem cells (red) and transit-amplifying intermediate progenitor (IP) cells (green). Expansion in one or both cell populations has been proposed as potential mechanisms that underlie neocortical expansion. Expansion of the founder RG cell population prior to the onset of neurogenesis (A) predicts a large ventricular zone (VZ). Expansion in the number of transit-amplifying divisions (B) predicts a large subventricular zone (SVZ).









**Figure 4. Contrasting Rodent and Human Neocortical Development**

(A) Current views of rodent corticogenesis are illustrated. Radial glial (RG) cells most often generate intermediate progenitor (IP) cells that divide to produce pairs of neurons. These neurons use RG fibers to migrate to the cortical plate. The historical view of neocortical development was that RG and neuronal progenitor cells were lineally distinct and that RG did not have a role in neurogenesis. Our current appreciation of the lineage relationship between RG cells, IP cells, and neurons has revised this view. The recent observation that small numbers of outer subventricular zone radial glia-like (oRG) cells exist in the mouse is also illustrated.

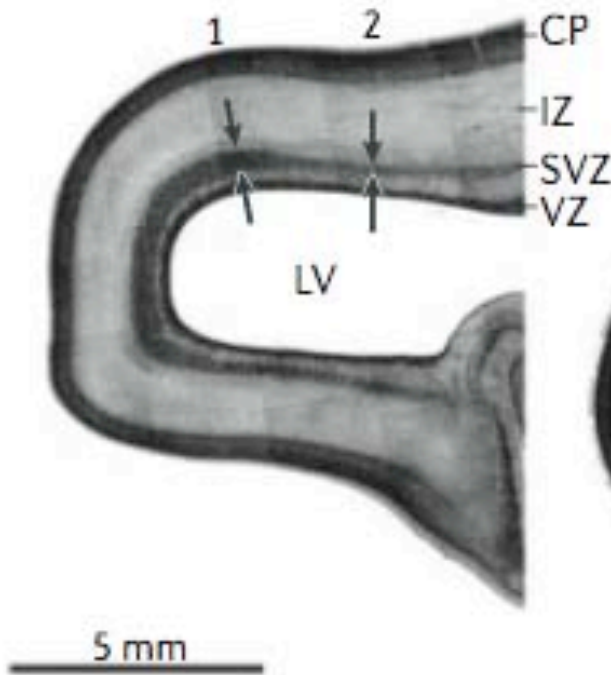
(B) We highlight the lineage of oRG cells, IP cells, and migrating neurons (red to green) present in the human outer subventricular zone (OSVZ) and the increased number of radial fibers that neurons can use to migrate to the cortical plate. The number of ontogenetic "units" is significantly increased with the addition of oRG cells over ventricular RG (vRG) cells. Maintenance of oRG cells by Notch and integrin signaling is shown. Short neural precursors (SNP), a transitional cell form between RG and IP cells, are also depicted in (A) and (B). For simplicity, we do not illustrate all of the cell types described in Figure 2E.

# Patterns of neural stem and progenitor cell division may underlie evolutionary cortical expansion

Kriegstein A, Noctor S, Martínez-Cerdeño V

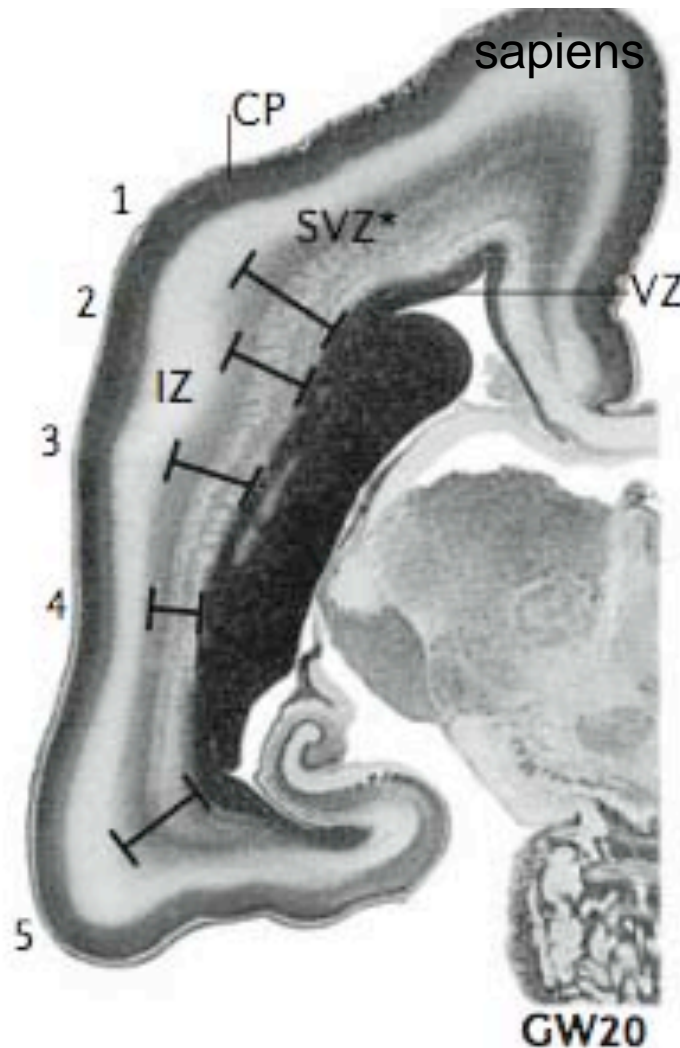
Nat Rev Neurosci  
2006 vol. 7 (11) pp. 883-90

macaque

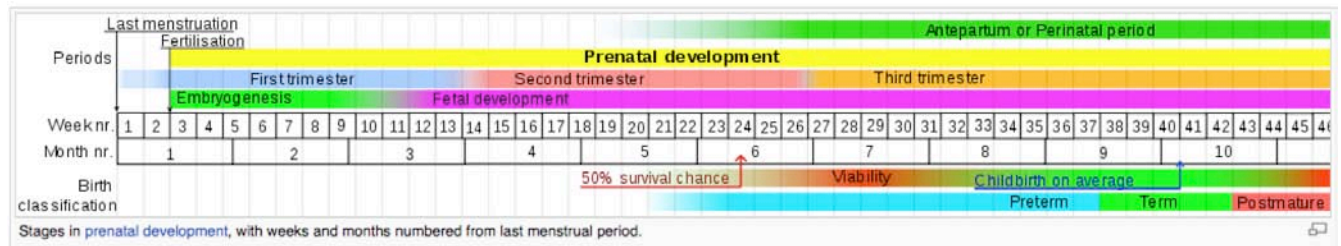
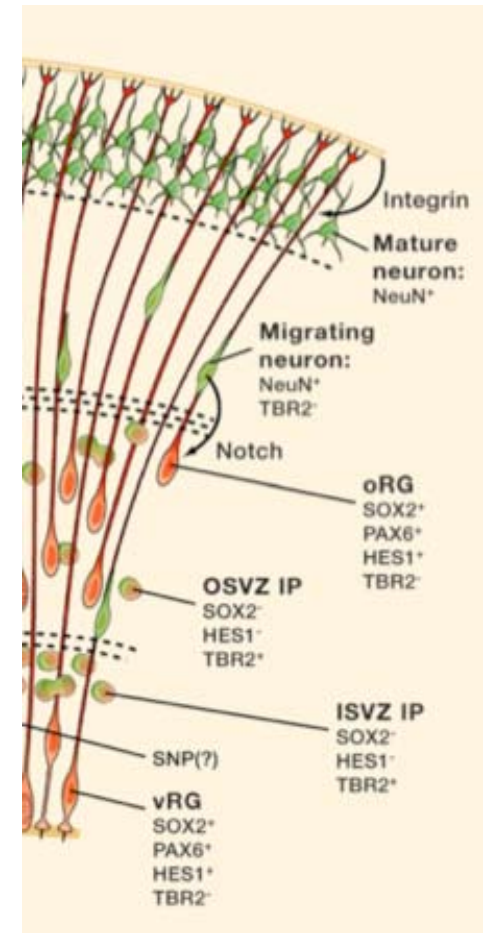


E78

sapiens



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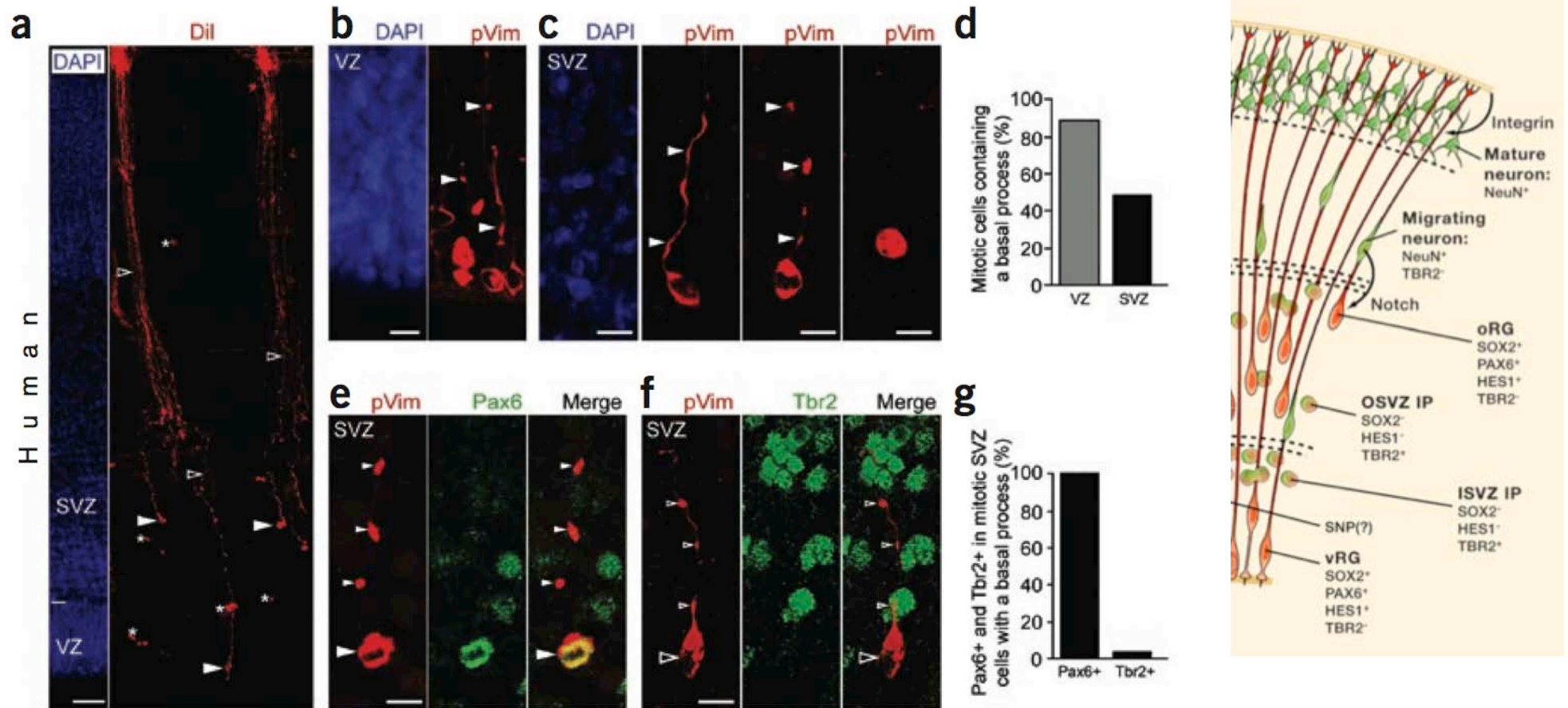




# OSVZ progenitors of human and ferret neocortex are epithelial-like and expand by integrin signaling

Fietz SA, Kelava I, Vogt J, Wilsch-Bräuninger M, Stenzel D, Fish JL, Corbeil D, Riehn A, Distler W, Nitsch R, Huttner WB

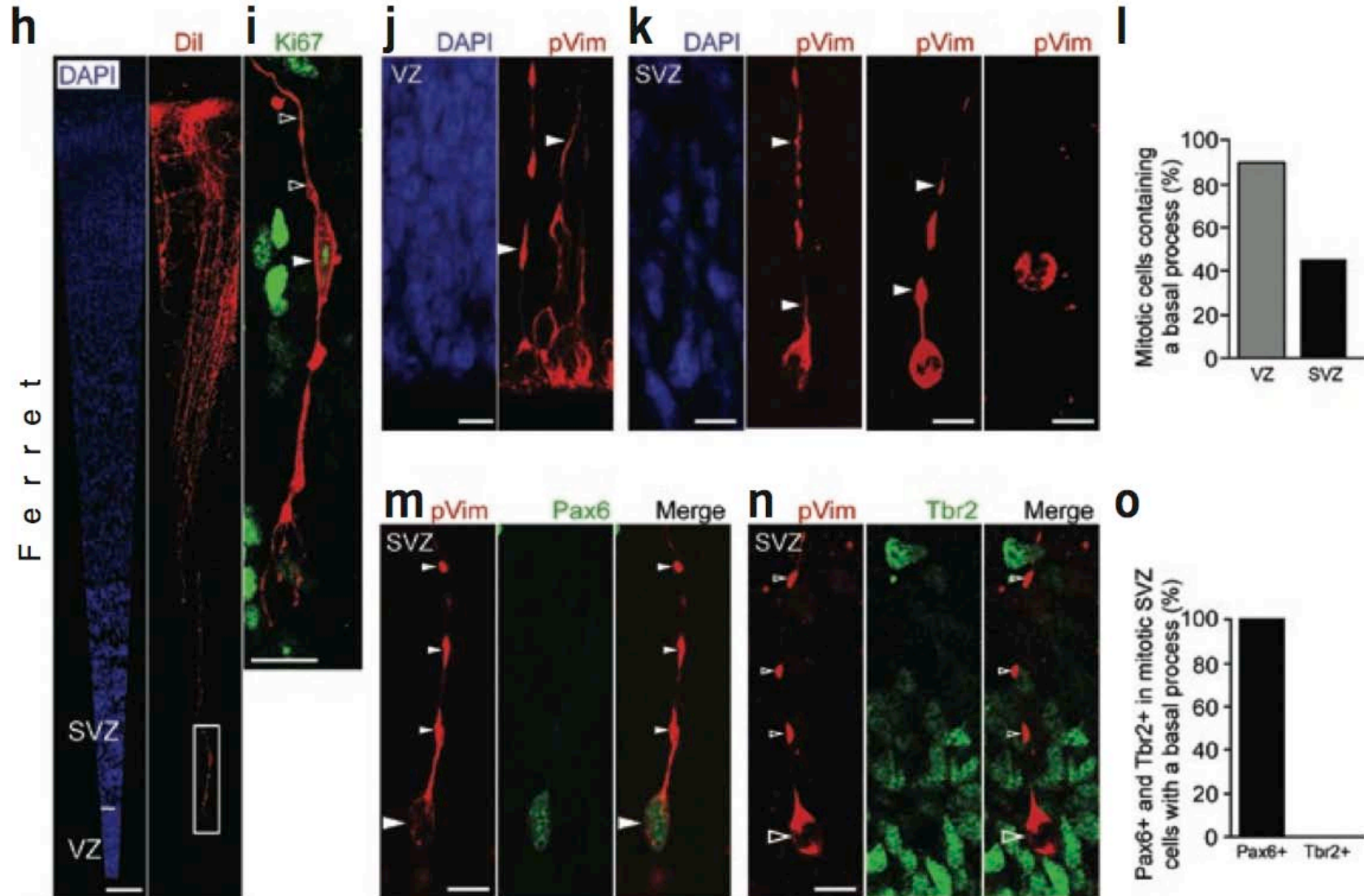
Nature Neuroscience  
2010 vol. 13 (6) pp. 690-9



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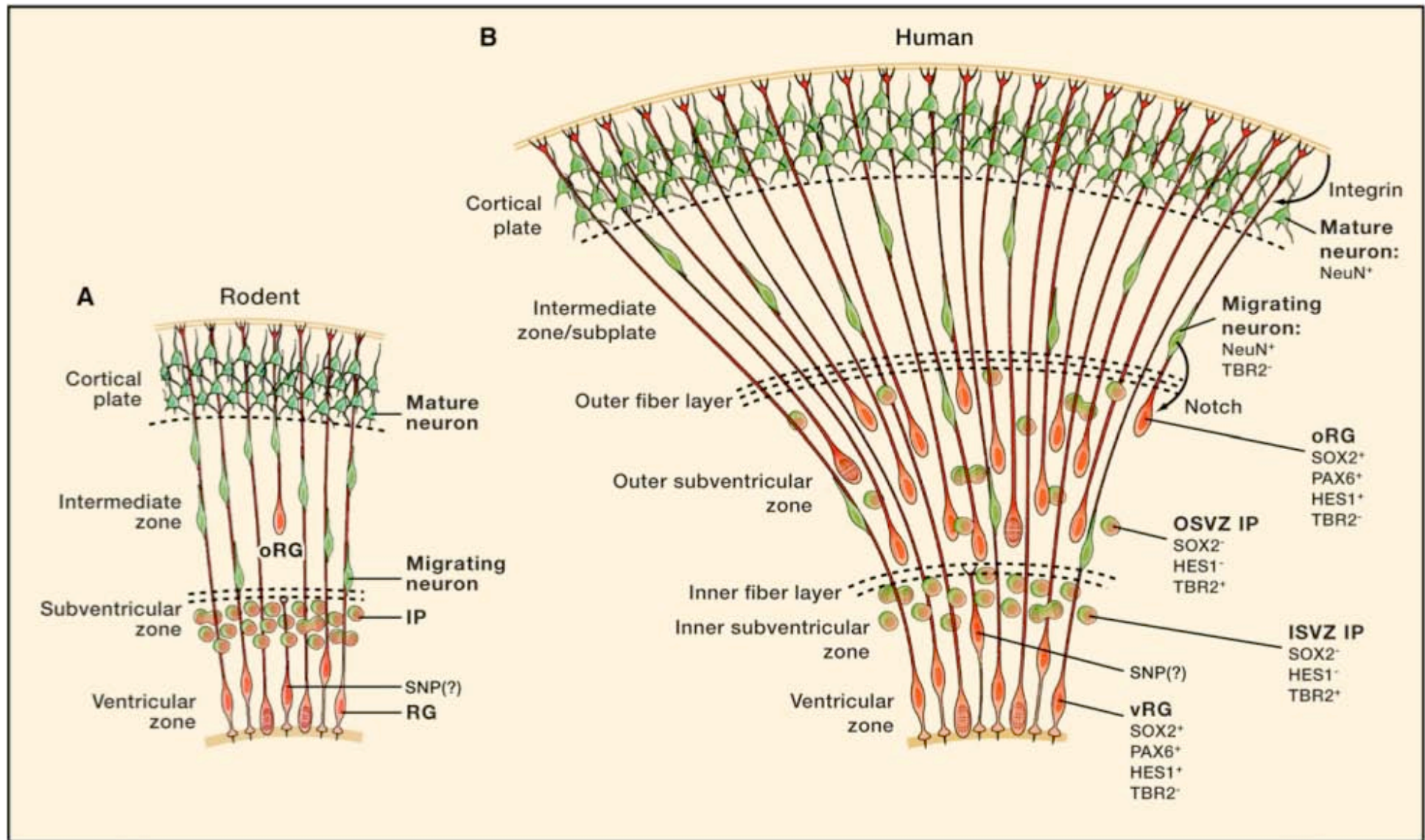
Nature Neuroscience  
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# Development and evolution of the human neocortex

Cell  
2011 vol. 146 (1) pp. 18-36

Lui JH, Hansen DV, Kriegstein AR



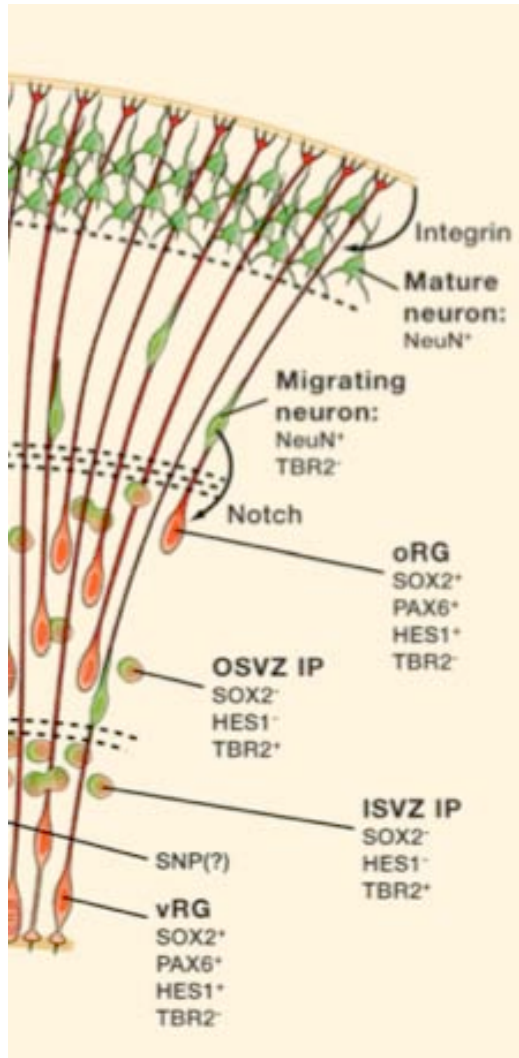


**Development and evolution of the human neocortex**

Lui JH, Hansen DV, Kriegstein AR

**Cell**

2011 vol. 146 (1) pp. 18–36



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