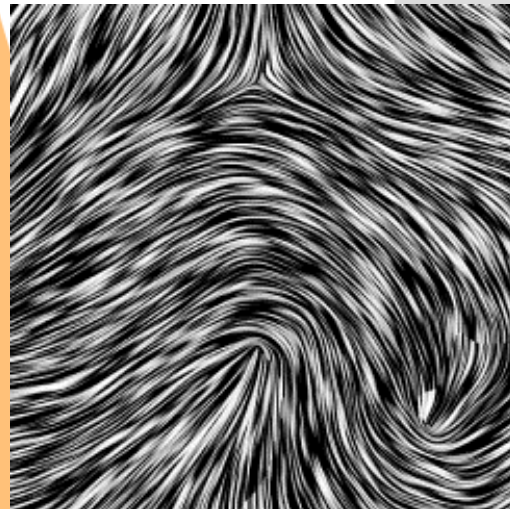




institut**Curie**

Active Cellular Nematics



Collective behaviors

Population scale



Fish

D. Hall



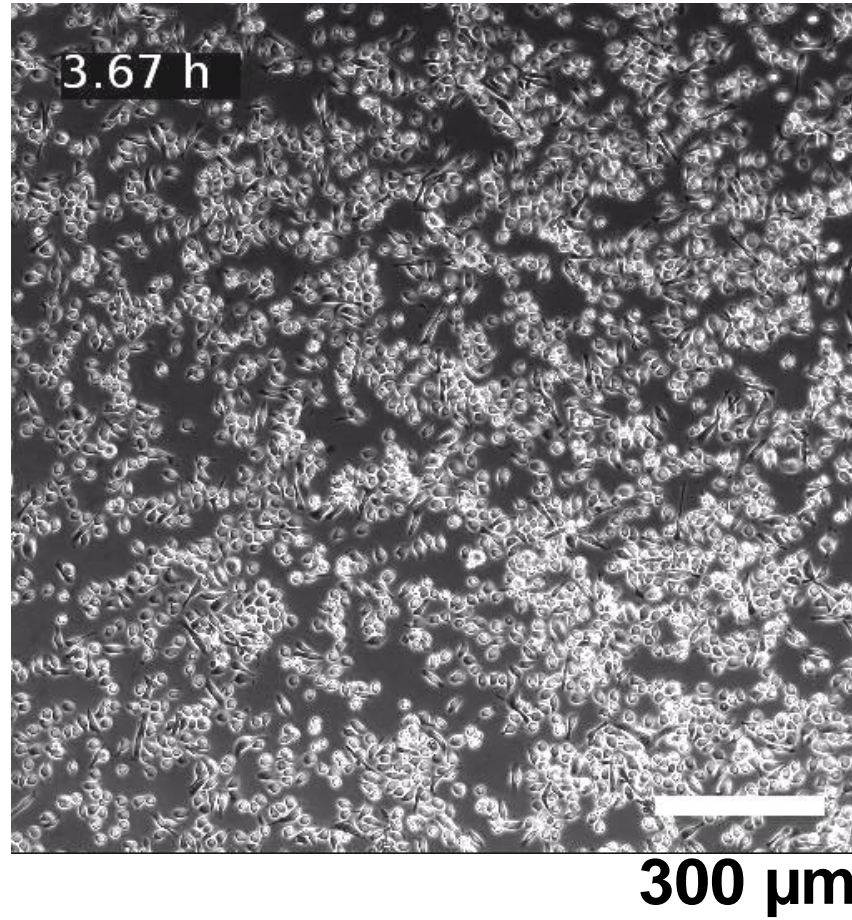
Starlings

M. Presti

Similar framework / concepts to multi-cellular systems

Review on collective cell migration: Hakim V., Silberzan P.: *Collective cell migration: a physics perspective*. *Rep. Prog. Phys.* **80**, (2017), 076601

HBE cells (Human Bronchial Epithelial)



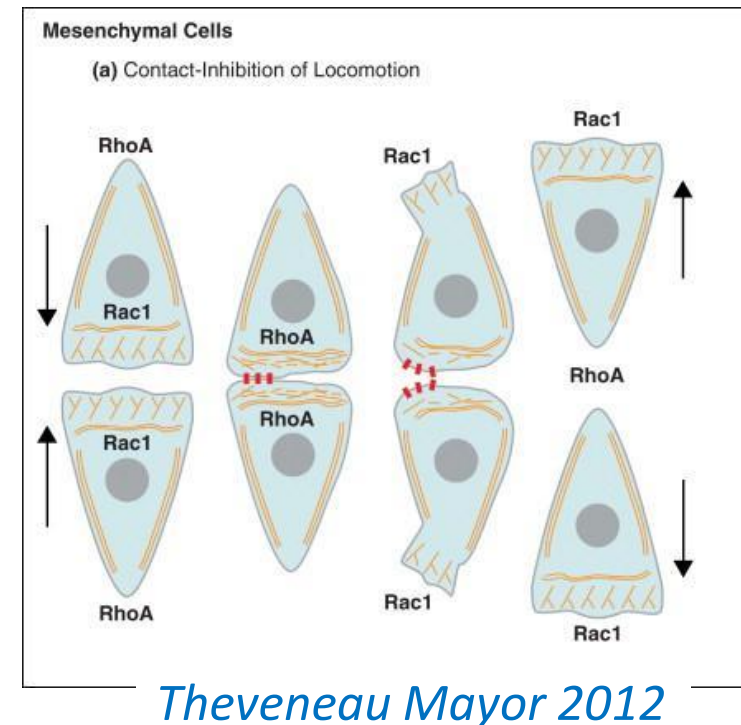
Slowing down as a consequence of
cell-cell interactions and cell
crowding:

Contact inhibition of locomotion

Contact inhibition of locomotion: a single cell concept

« *Contact inhibition consists of the **abolition or reduction of the power of the leading membrane to direct the general cell movement, and hence the assumption of dominance by another membrane, which then **redirects the cell.***** »

M. Abercrombie (1958)



A dark-field microscopy image showing a horizontal band of tumour cells. The cells appear as a dense, textured layer with some brighter spots. The background is black.

0 min

Tumour cells - S180

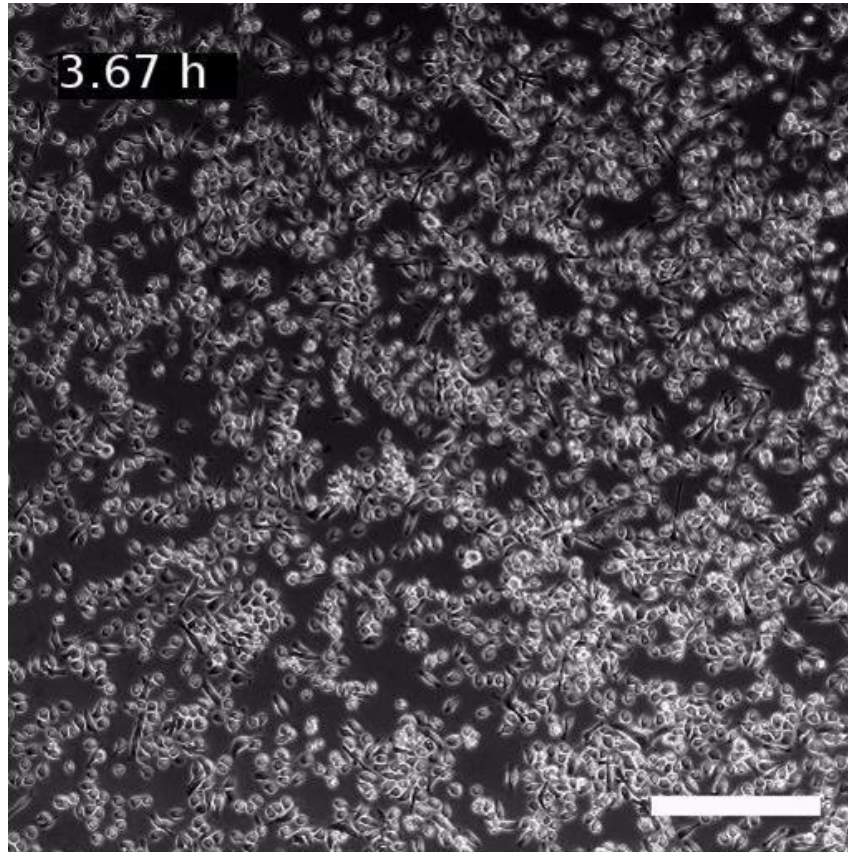
Michaël Abercrombie (UCL, 50's)

**wellcome
library**

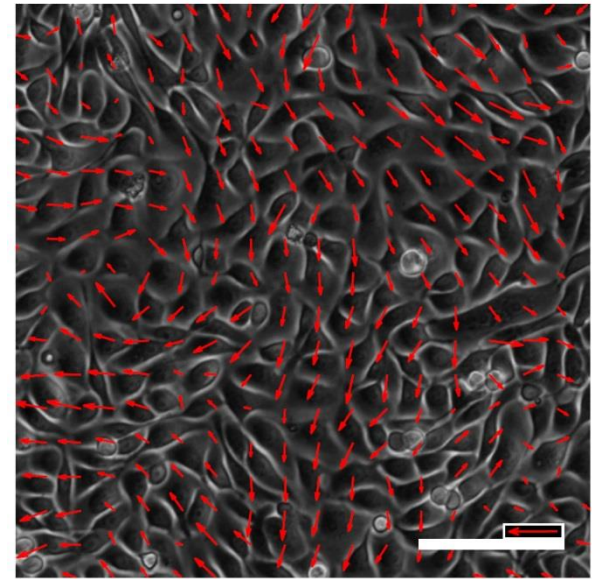
The library at Wellcome Collection

HBECs

(*Human bronchial epithelial cells*)



300 μm



100 $\mu\text{m}/\text{h}$



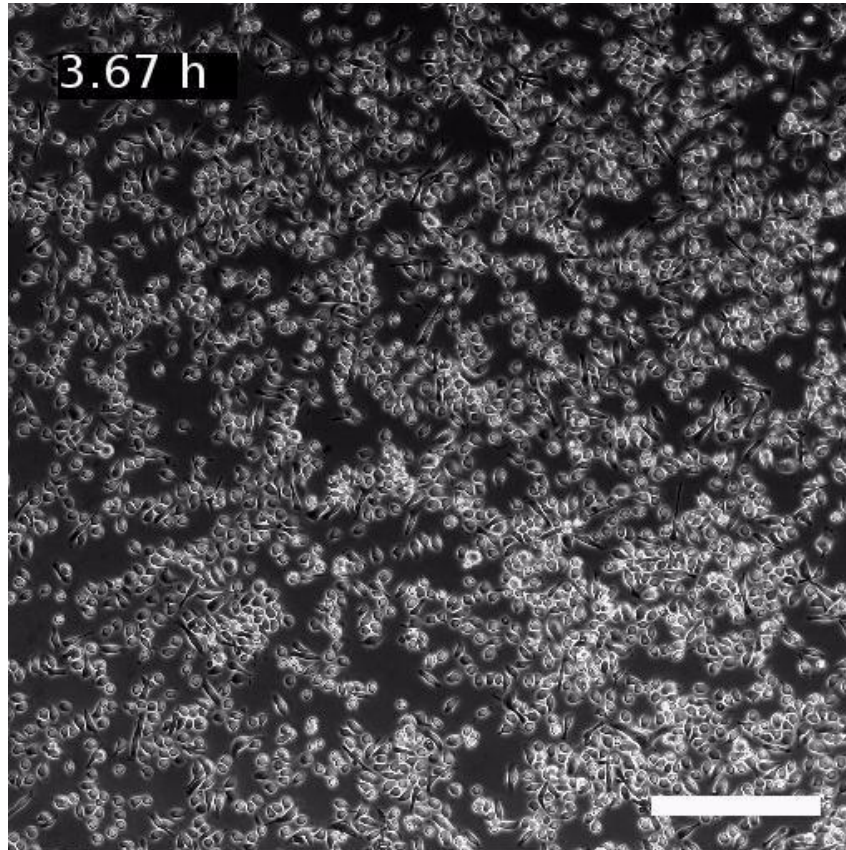
100 μm



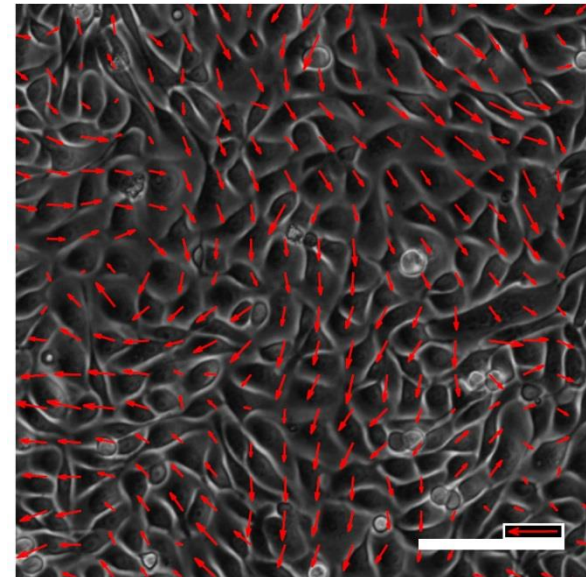
HBECs

(*Human bronchial epithelial cells*)

Cell jamming ?



300 μm



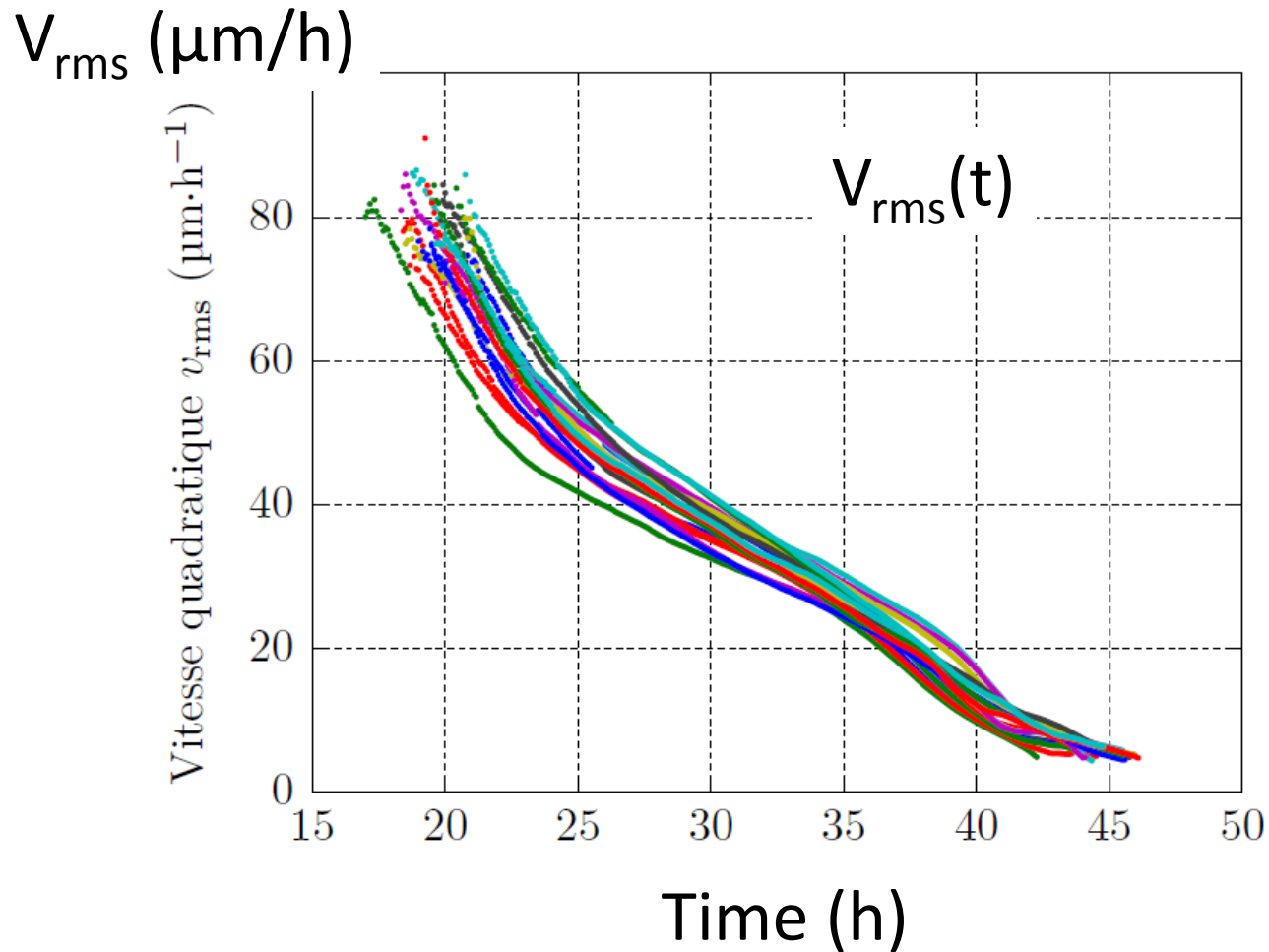
100 $\mu\text{m}/\text{h}$



100 μm

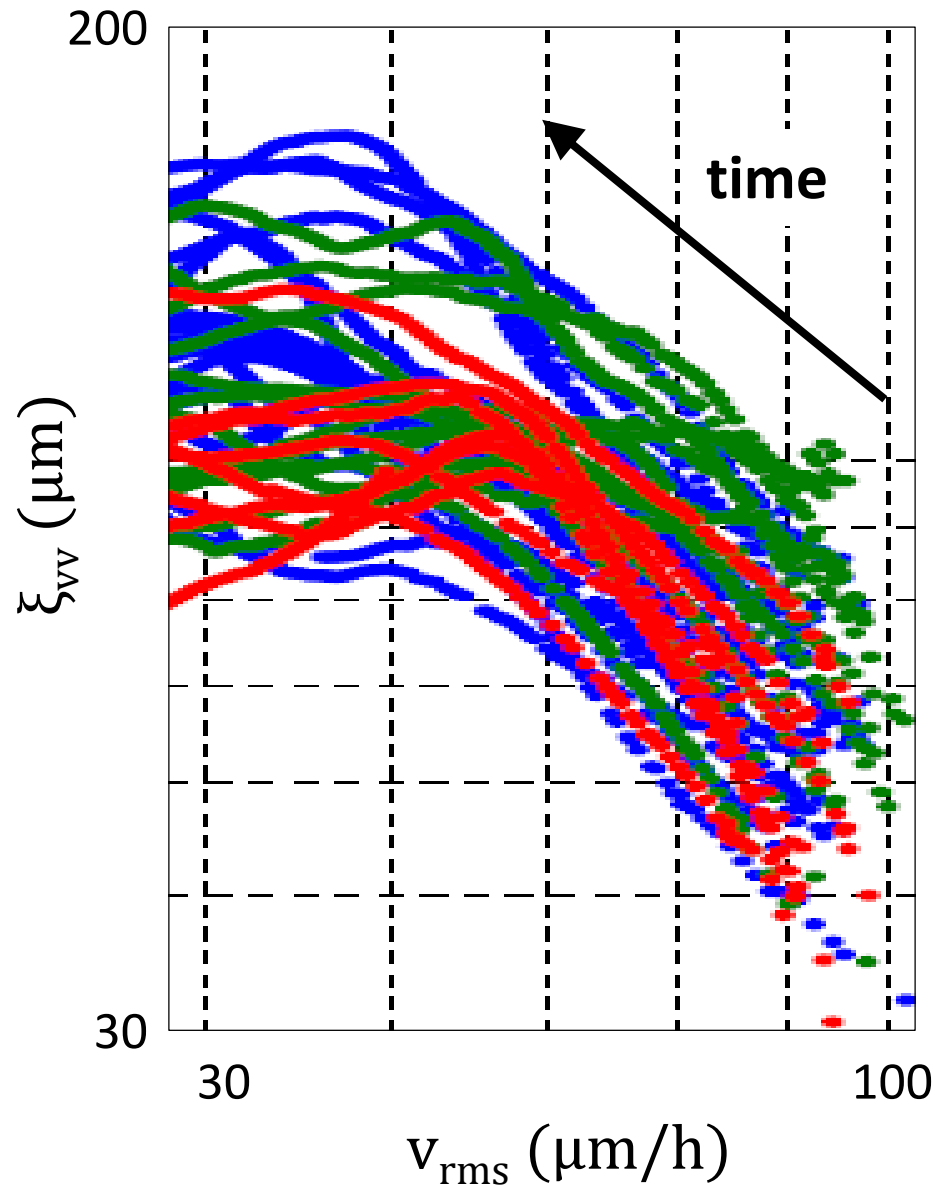


Slow down of displacements with time



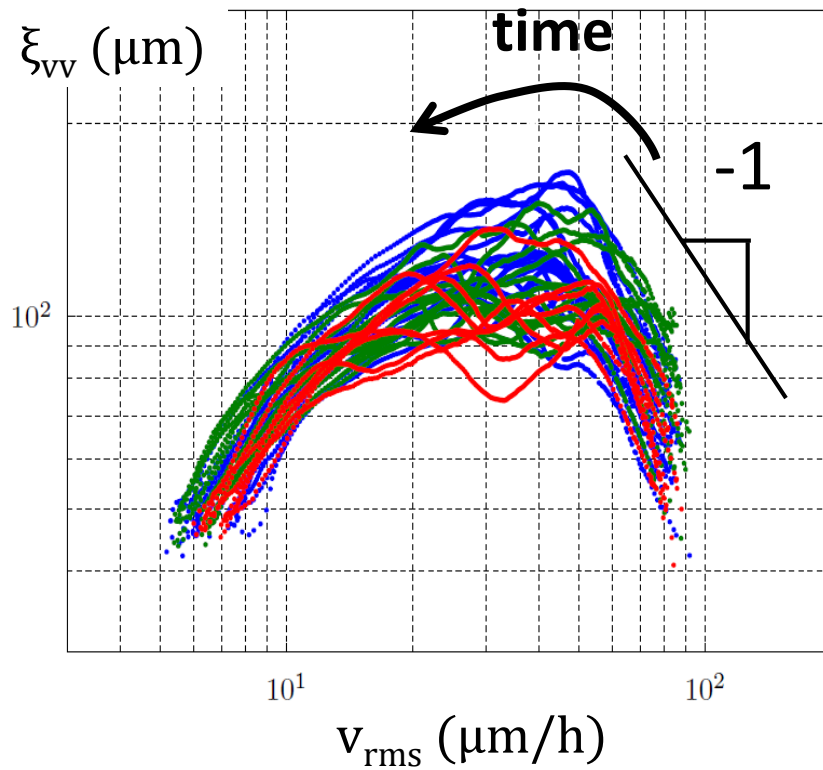
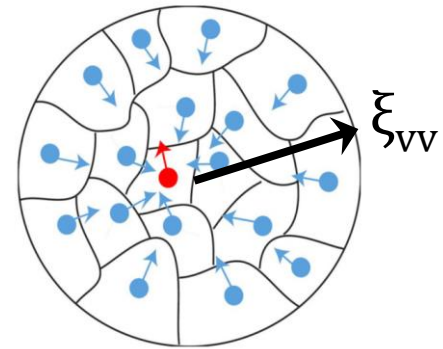
$$v_{\text{rms}}(t) = \sqrt{\langle \vec{v}(\vec{r}, t)^2 \rangle_{\vec{r}}}$$

Jamming of a HBEC monolayer

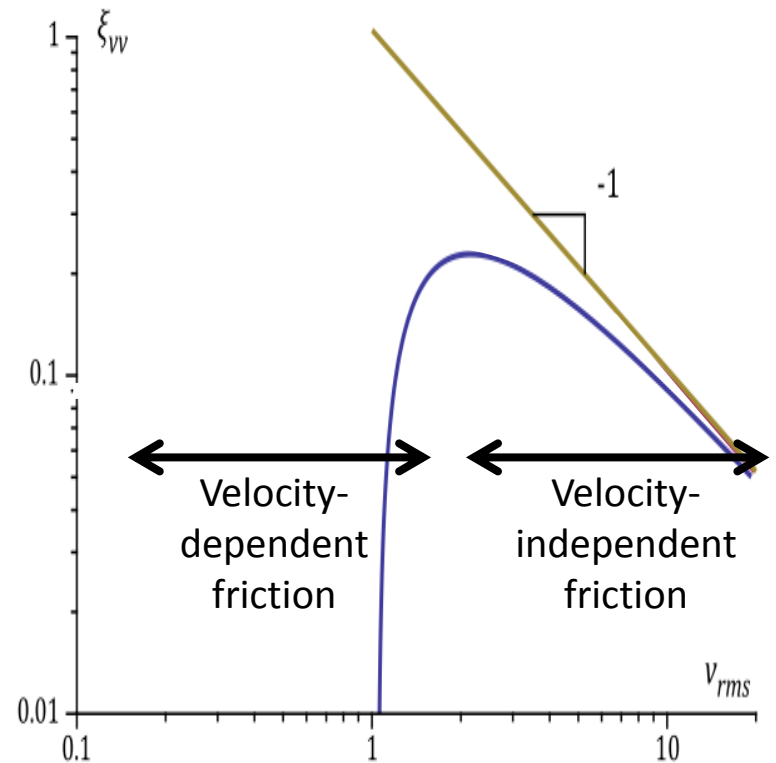


$$\xi_{vv} \propto \frac{1}{v_{\text{rms}}}$$

Jamming of a HBEC monolayer



Cluster model (Nir Gov)

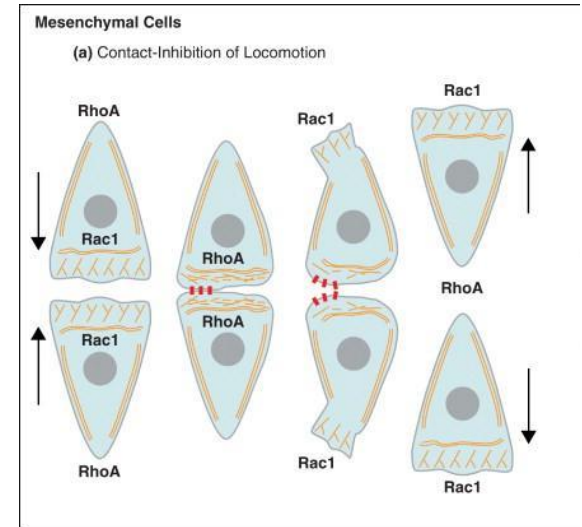


Garcia PNAS 2015

Cell Jamming

Contact Inhibition of Locomotion (M. Abercrombie (1958))

Single cell concept

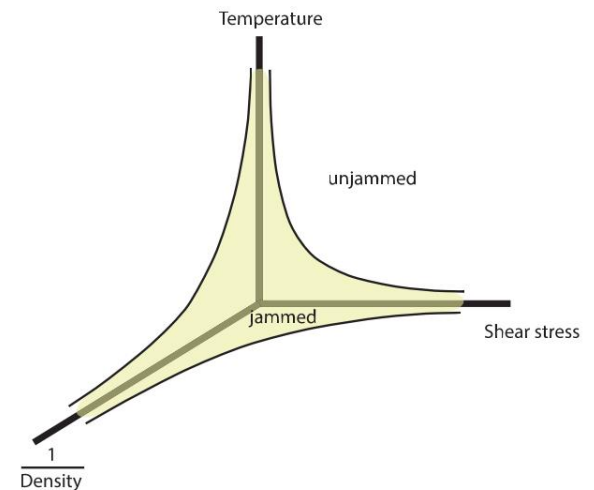


Theveneau Mayor 2012



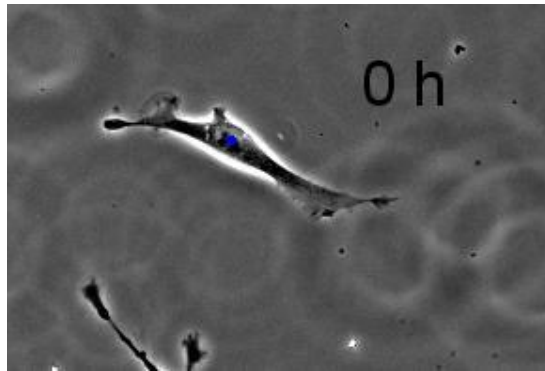
Cell Jamming (D. Weitz, J. Fredberg (2010's))

Collective



***Dynamics and architecture of cells
in their route to jamming***

3T3 fibroblasts

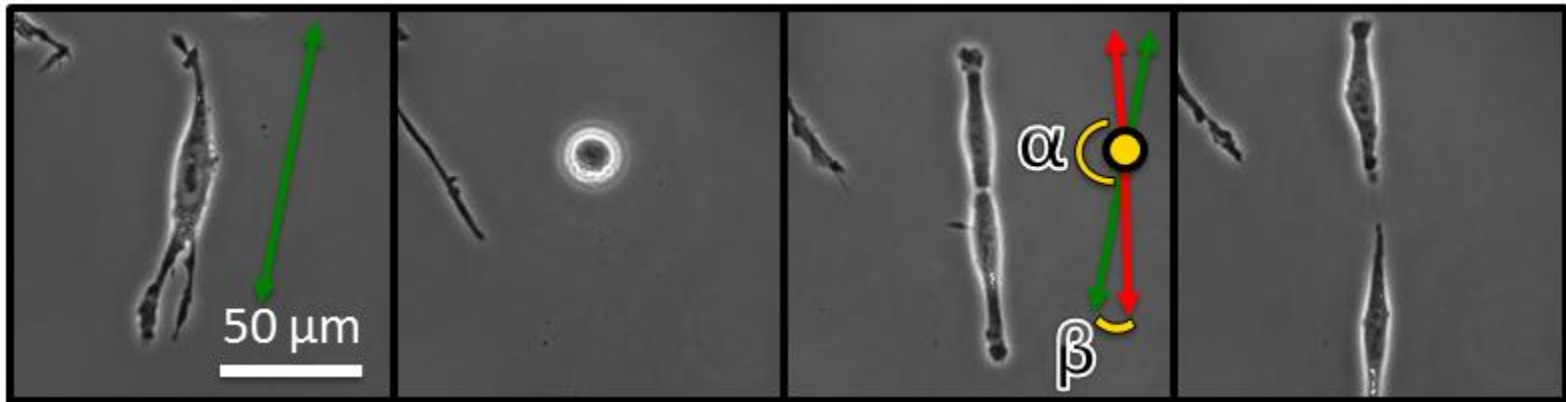


50 μm



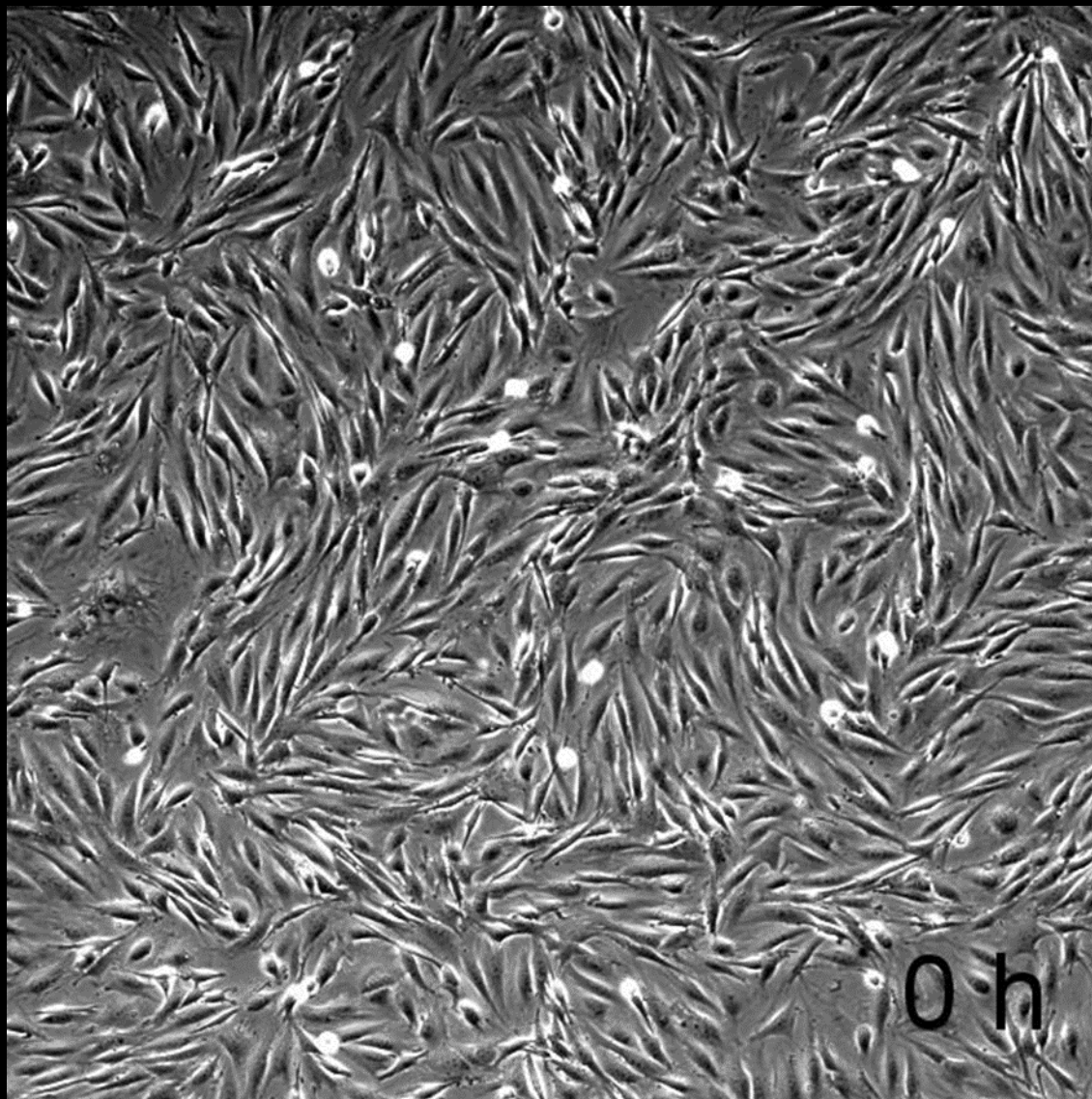
Spindle-shaped cells
Apolar

Cell division :



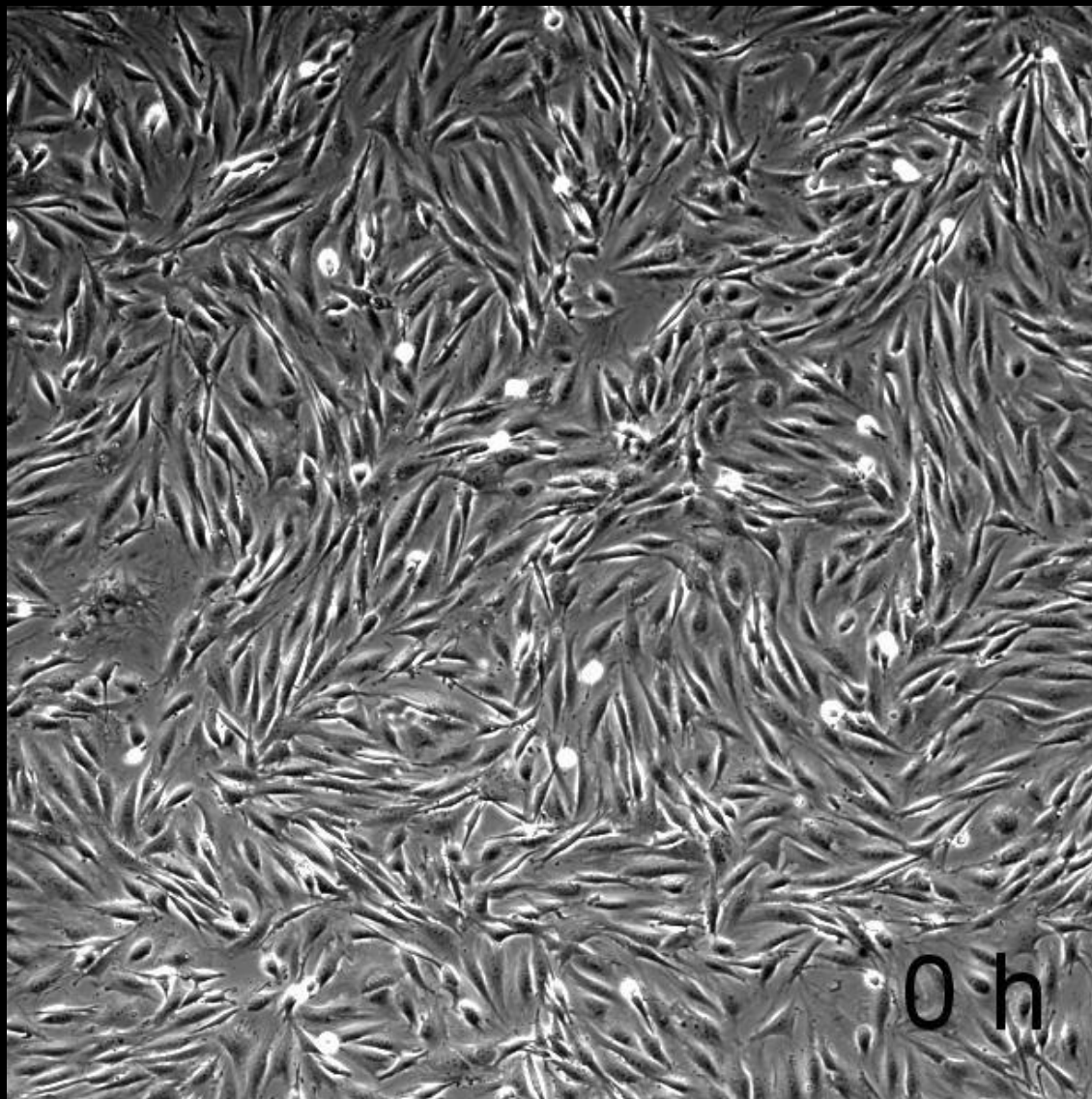
No cadherin-mediated cell-cell adhesion

Fibroblasts
NIH-3T3



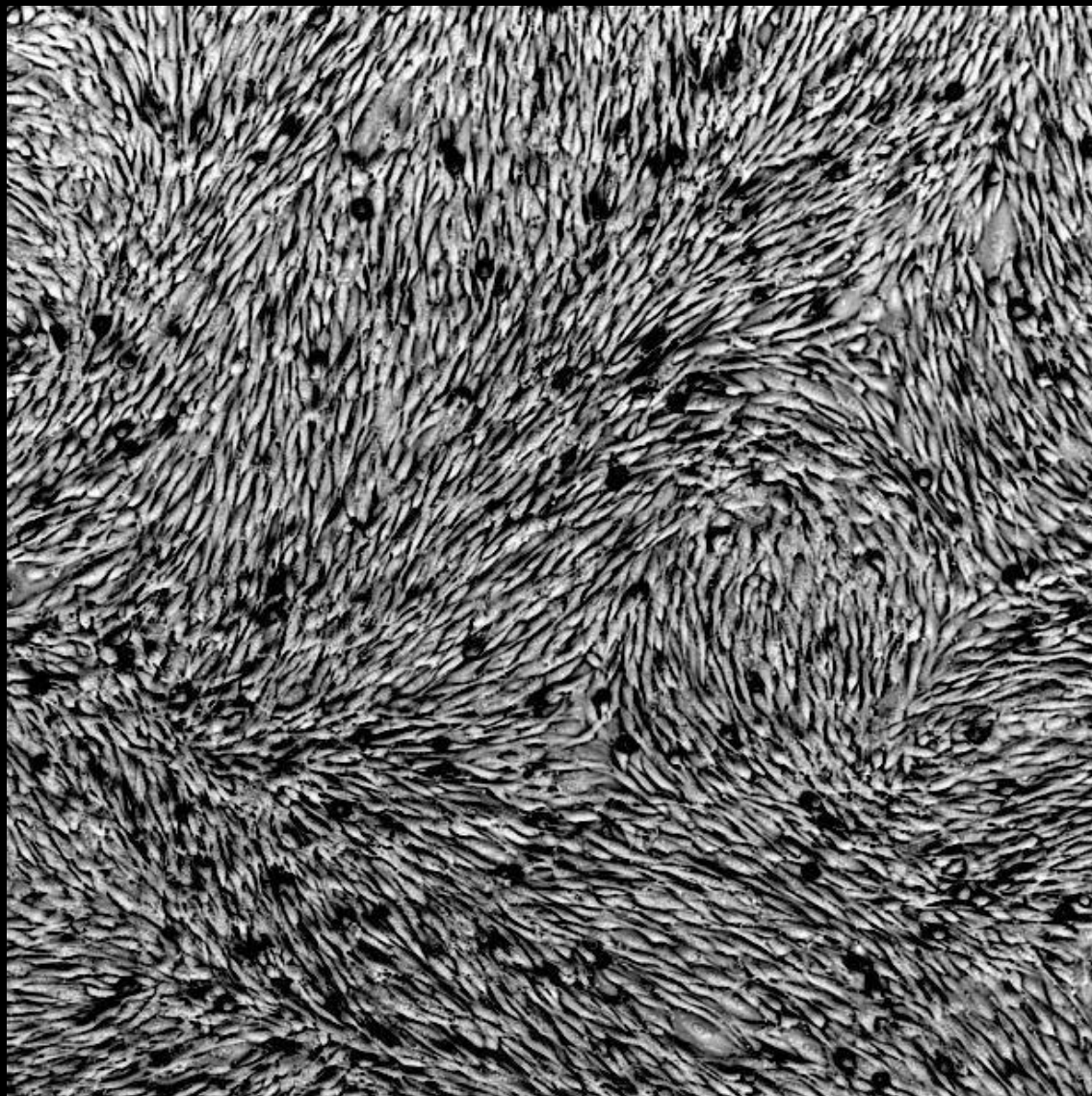
400 μm

Fibroblasts
NIH-3T3



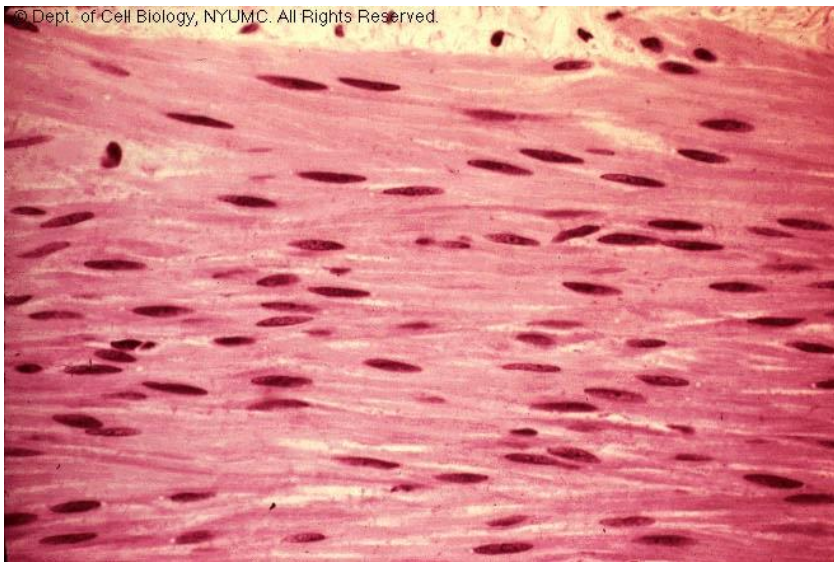
400 μm

Fibroblasts
NIH-3T3



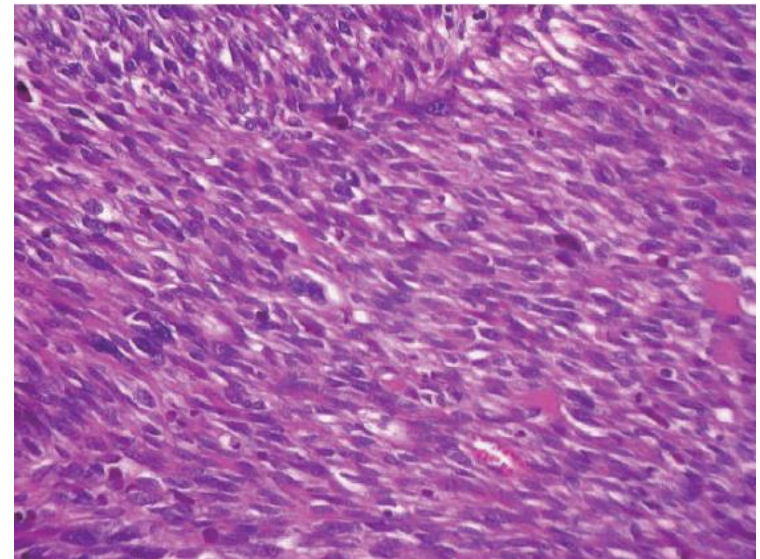
400 μm

A nematic-like ordering also observed in vivo



Smooth muscle

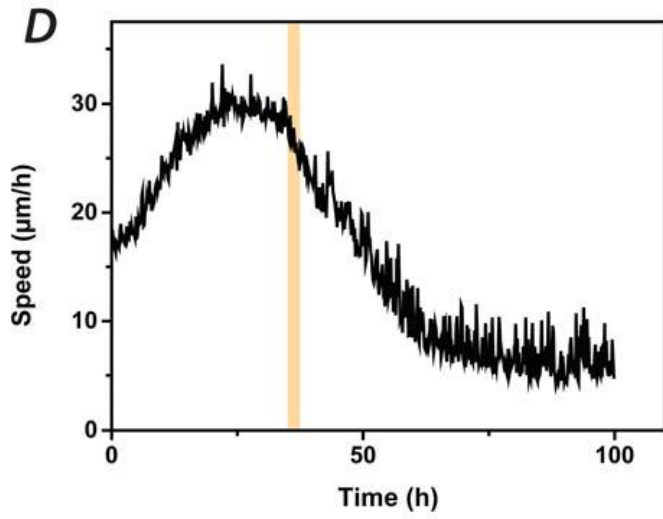
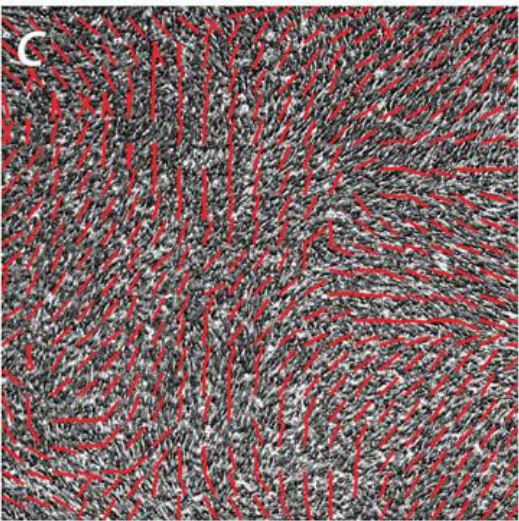
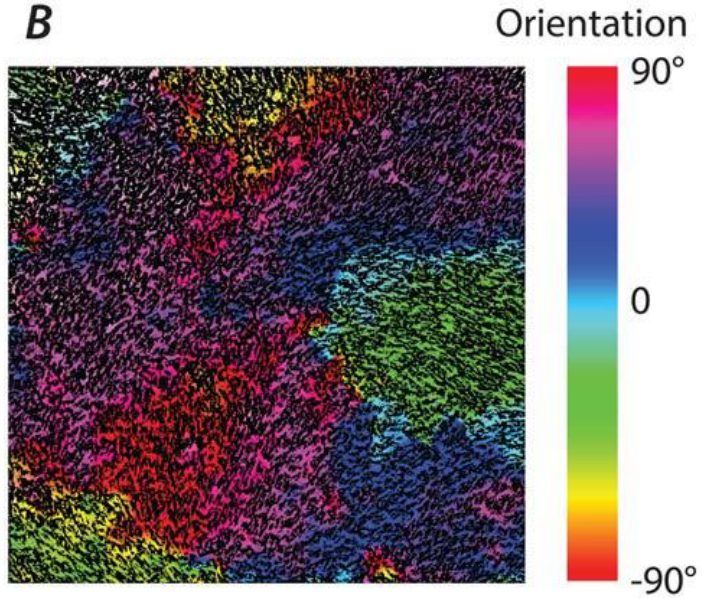
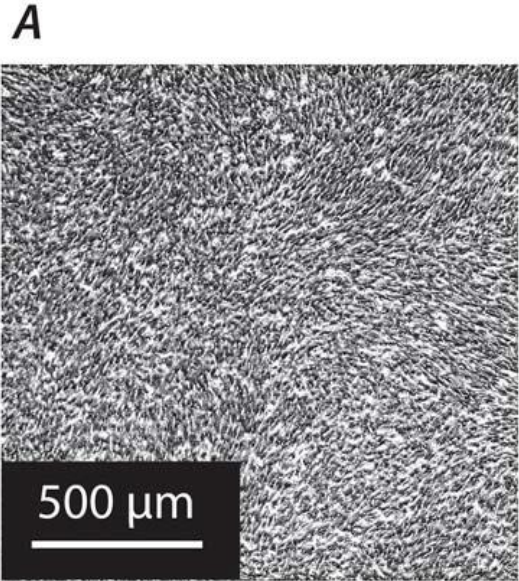
Dpt of Cell Biology NYUMC



Sarcoma

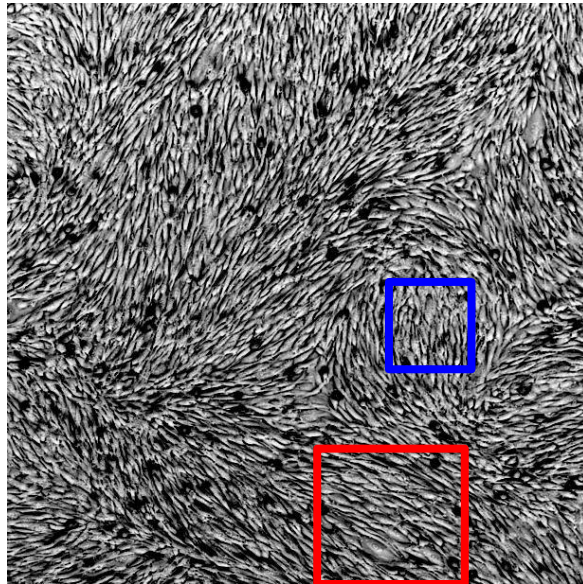
Braham Ann Trans Med 2013

Order in a fibroblasts monolayer

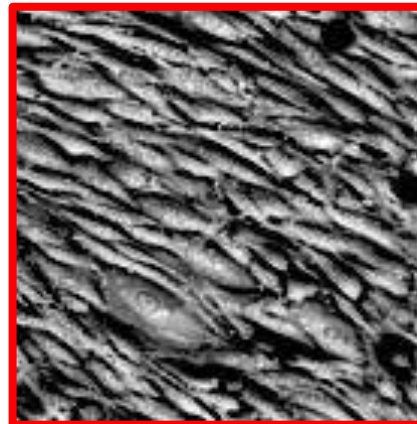
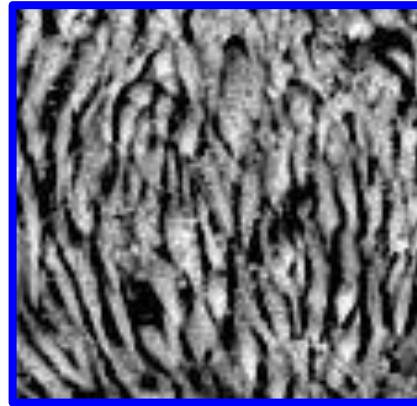


Nematic ordering of spindle-shaped cells

Fibroblasts
NIH-3T3

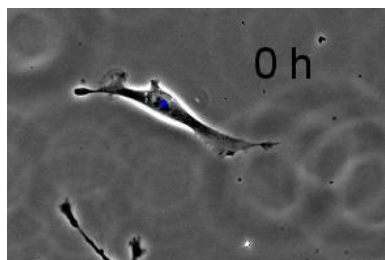


400 μm

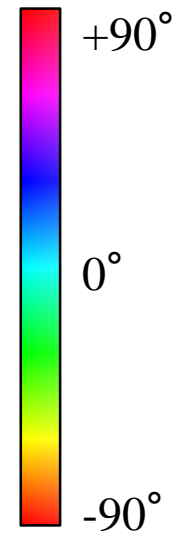
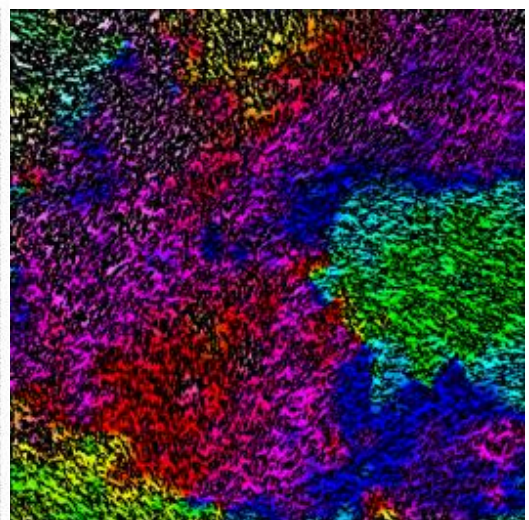
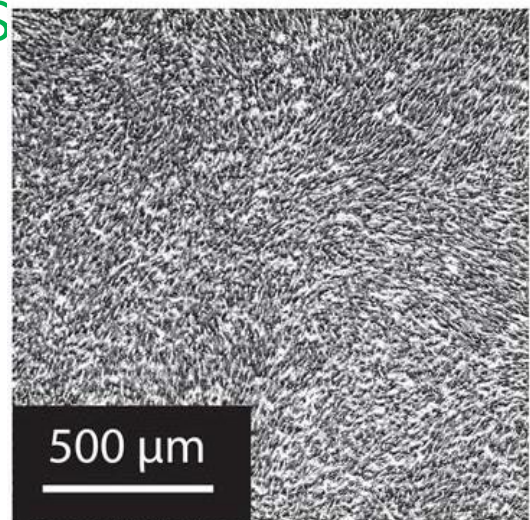


Duclos Soft Matter 2014
Duclos Nat Phys 2017
Duclos Nat Phys 2018

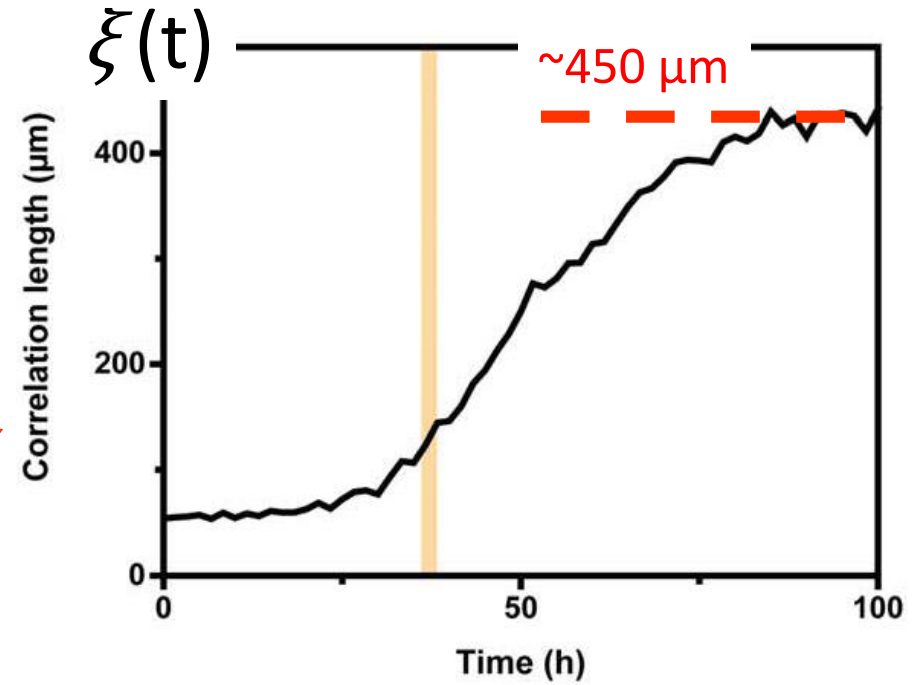
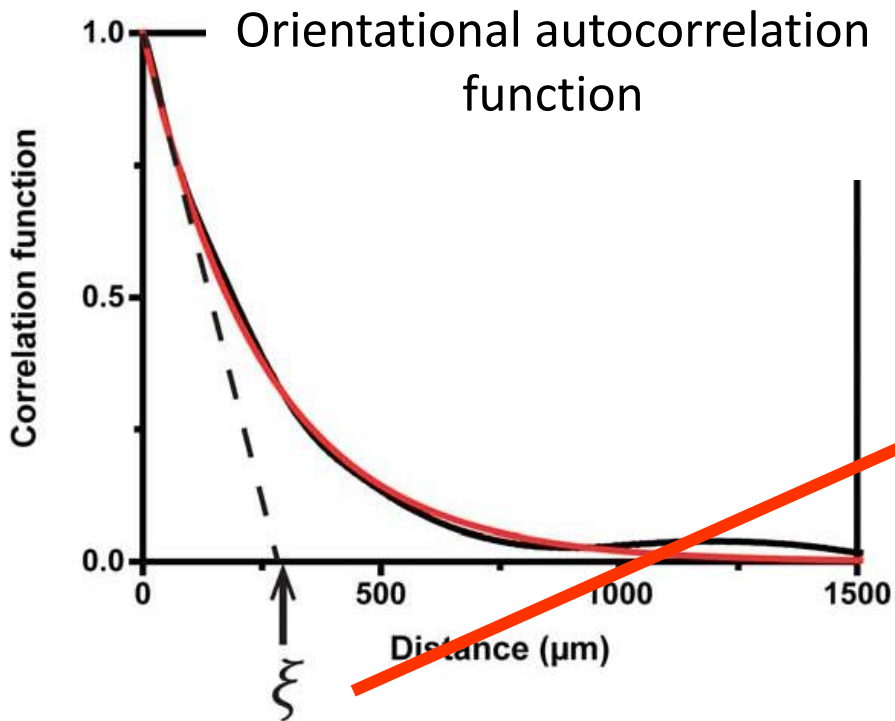
Orientation clusters



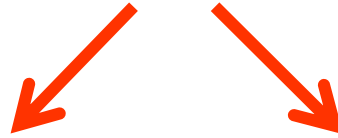
50 μm
3T3 cells



Activity

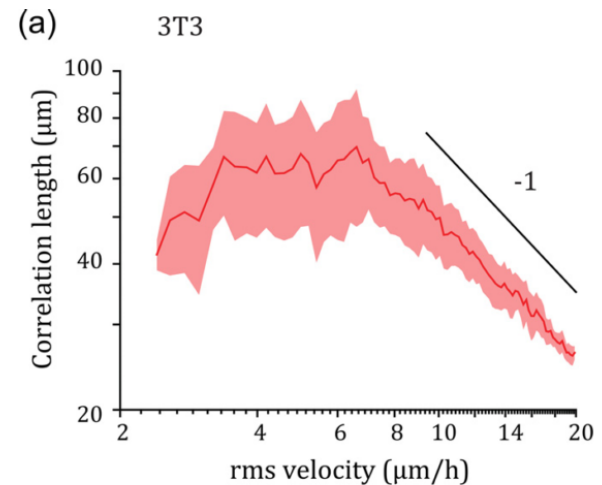
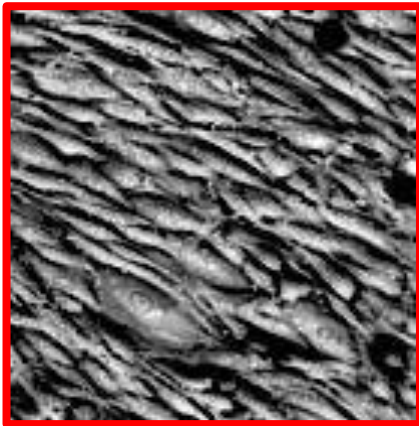


As **time** goes on



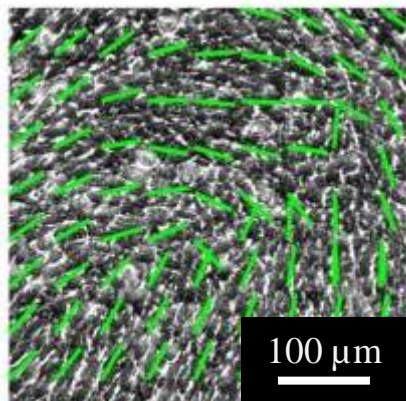
Better orientation
Steric interactions

Less movement
Jamming

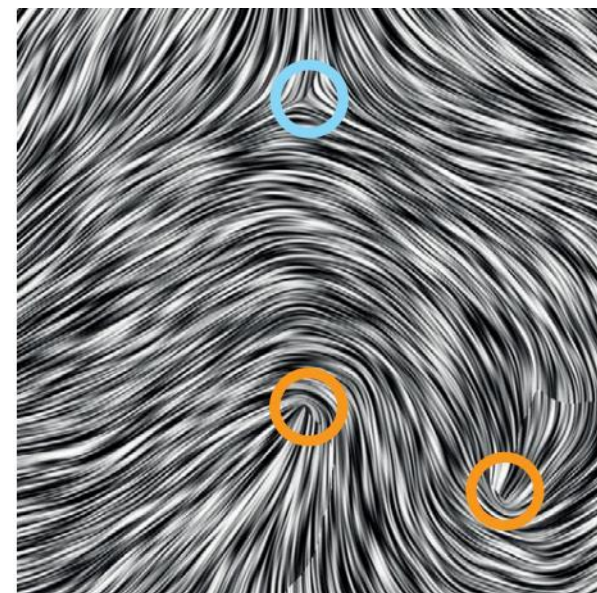
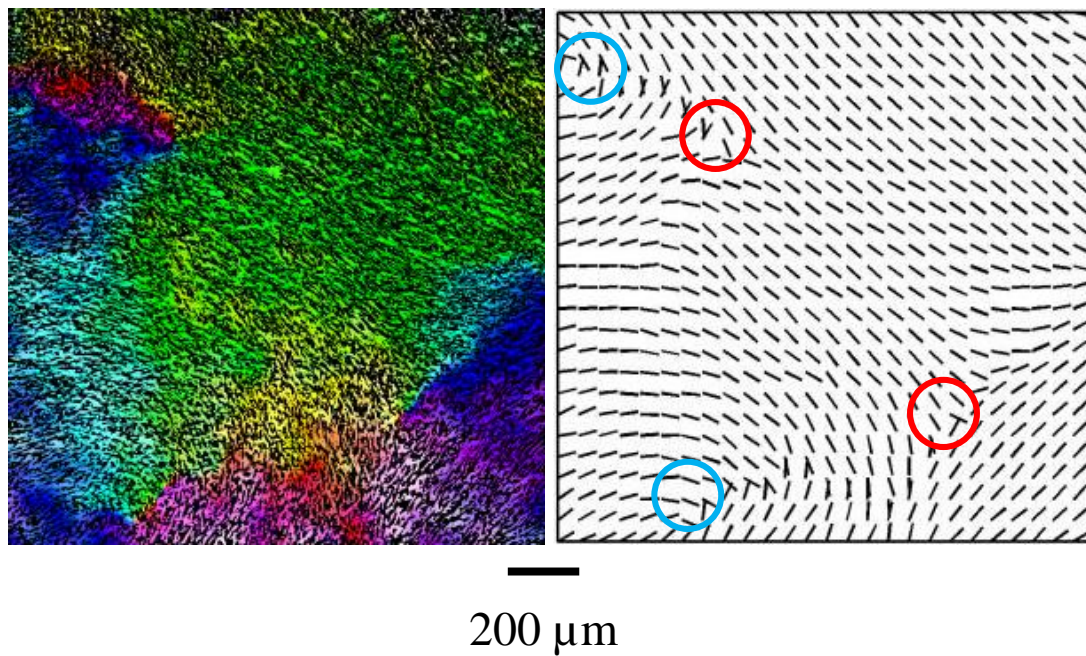
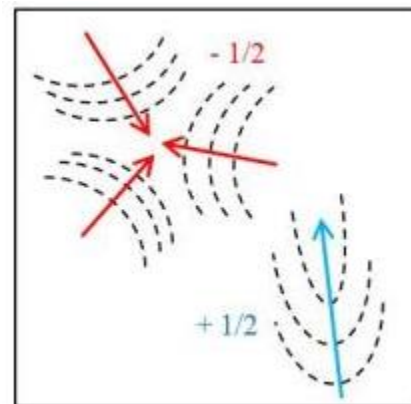
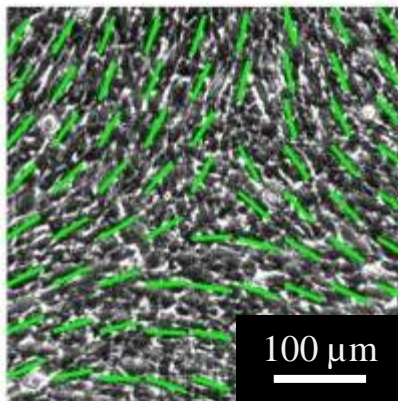


A true nematic ? The proof is in the defects

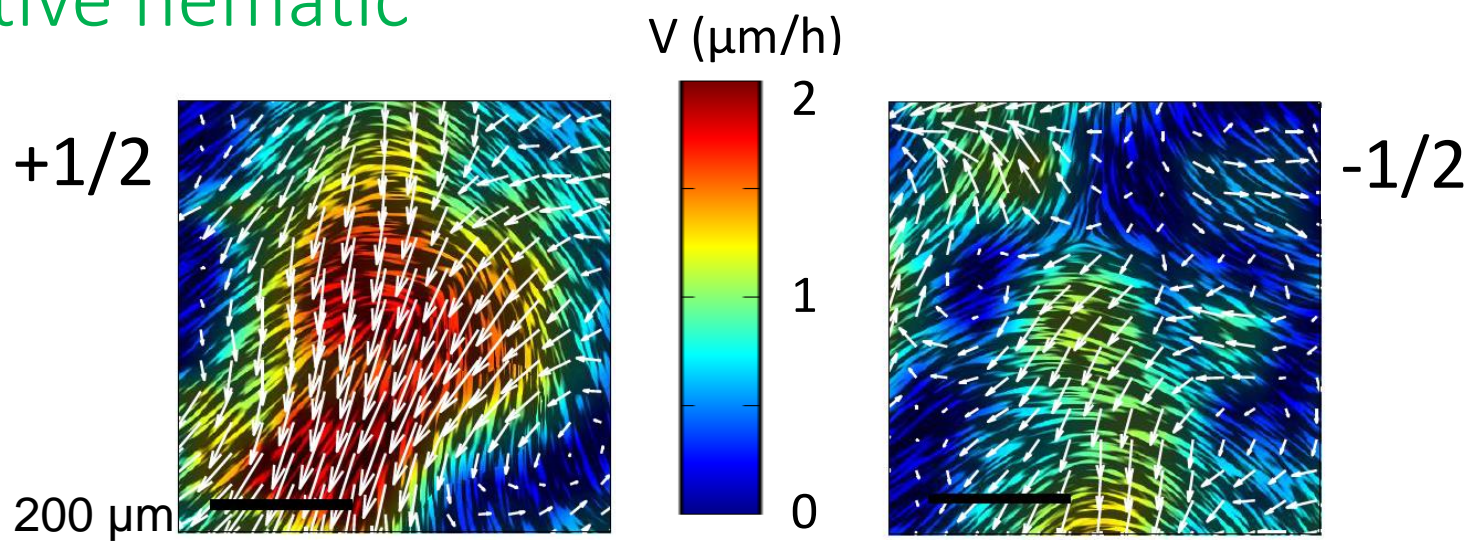
+1/2



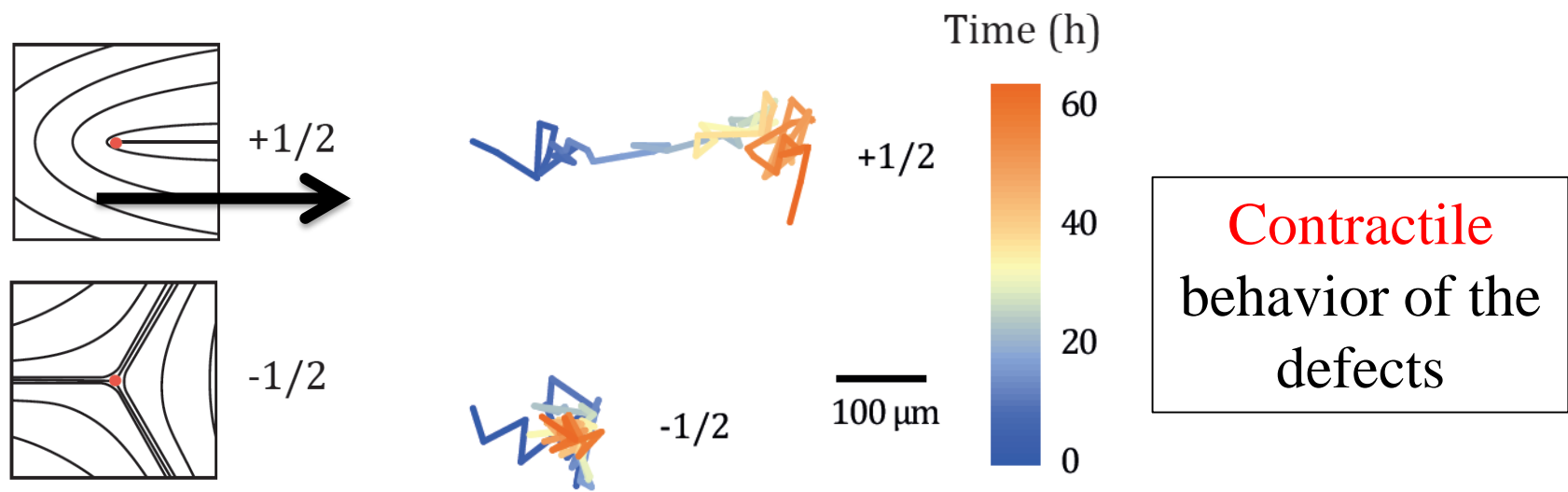
-1/2



An active nematic



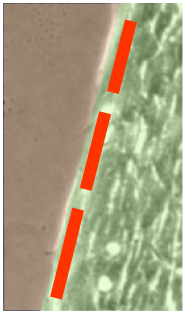
- Self-propulsion of $+1/2$ defects
- Active nematic backflow via the monolayer itself



How to reach perfect alignment ?

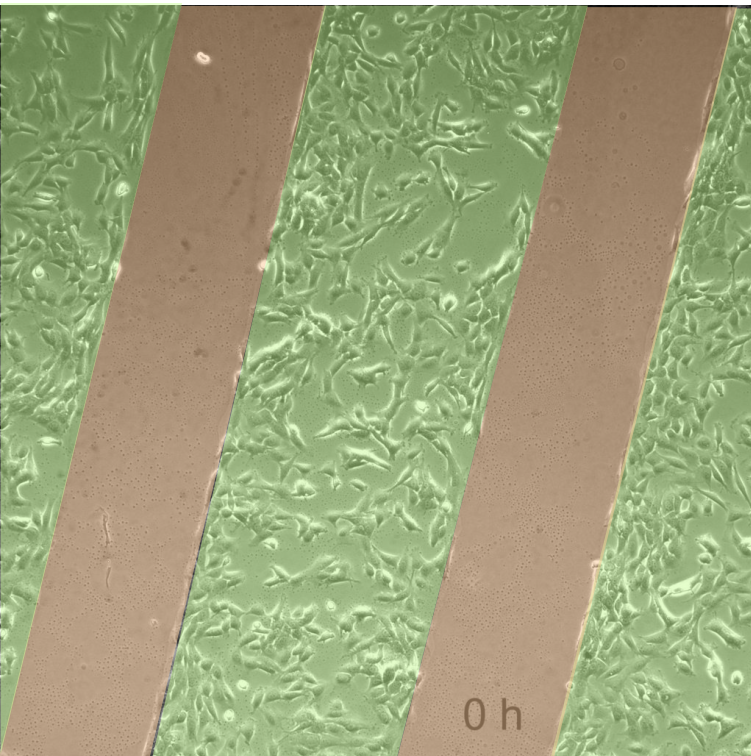
Liquid Crystal strategy :
Use **boundaries**

non adhesive

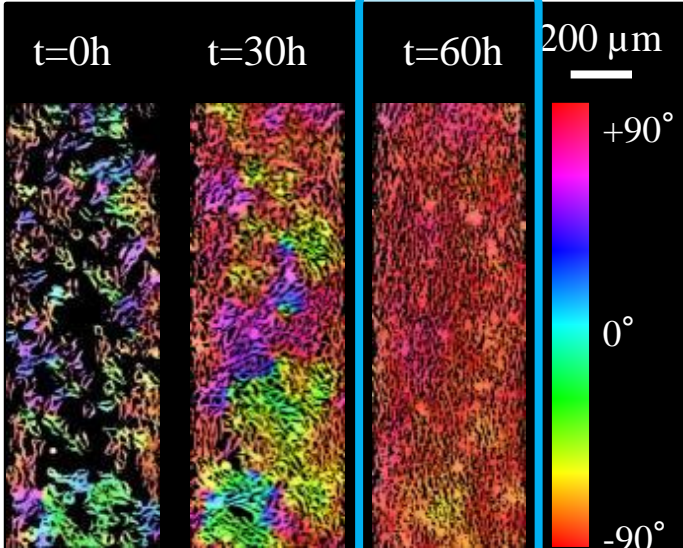


adhesive

Cells align **along** boundaries

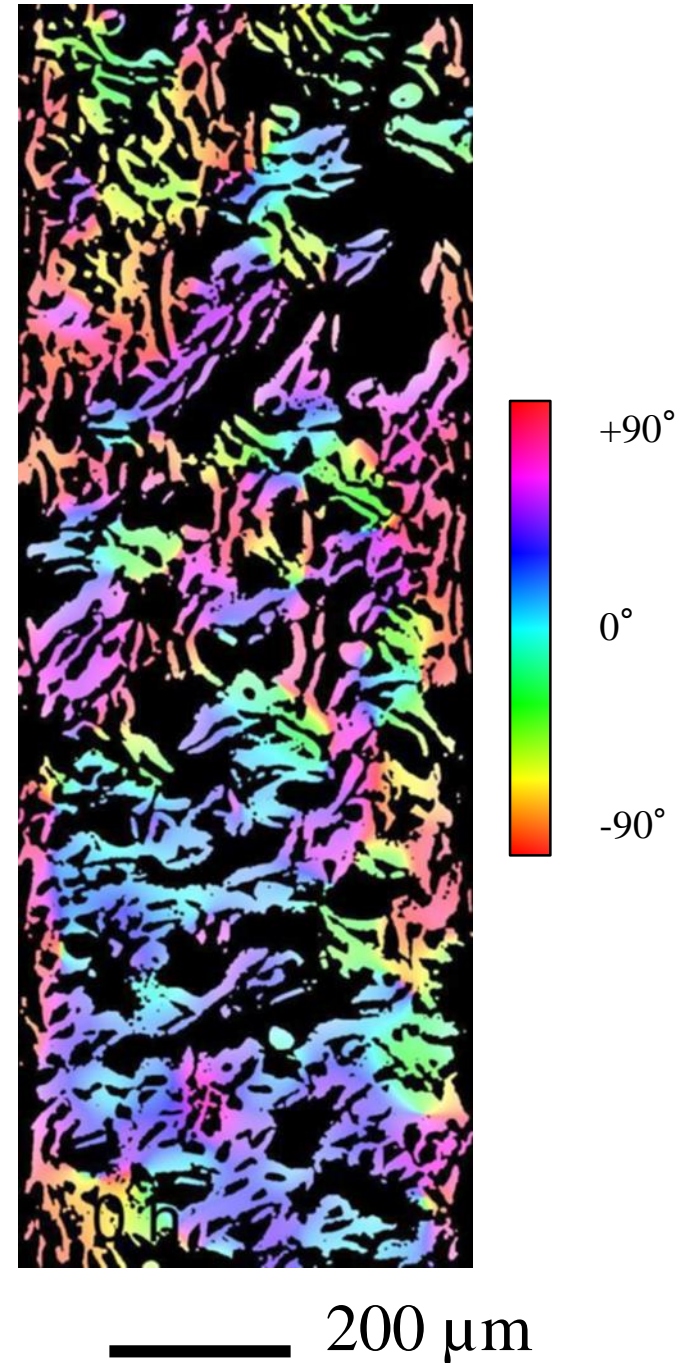
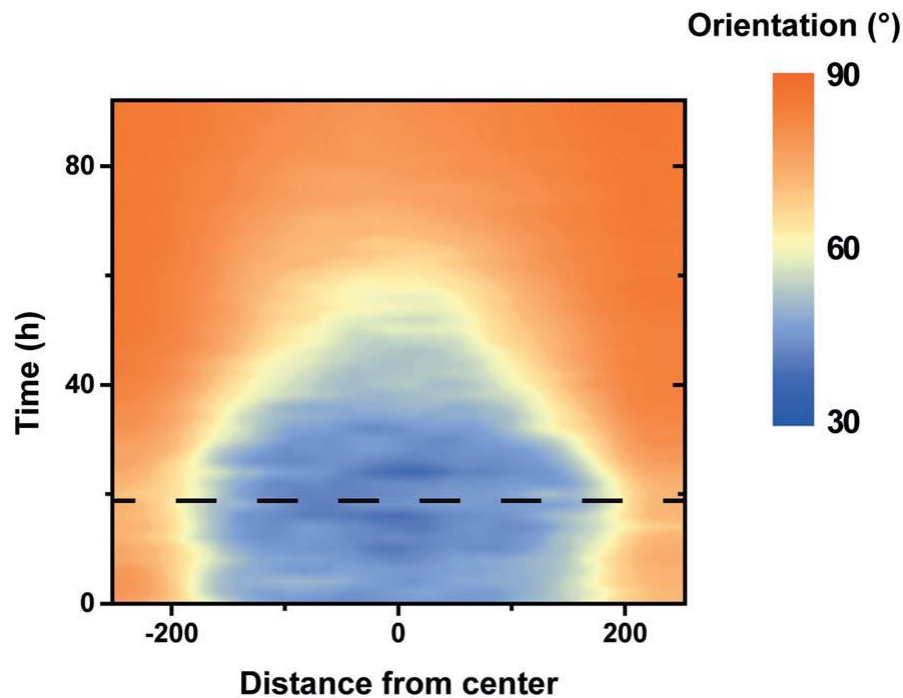


200 μm

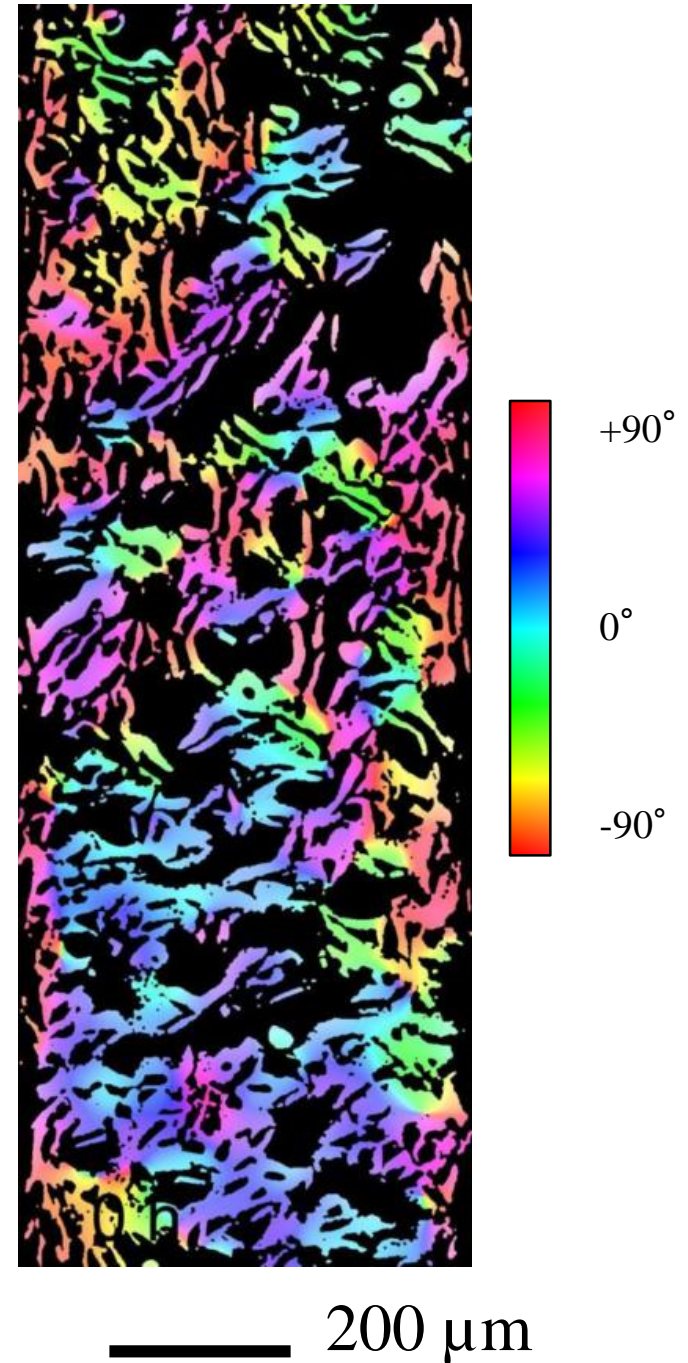
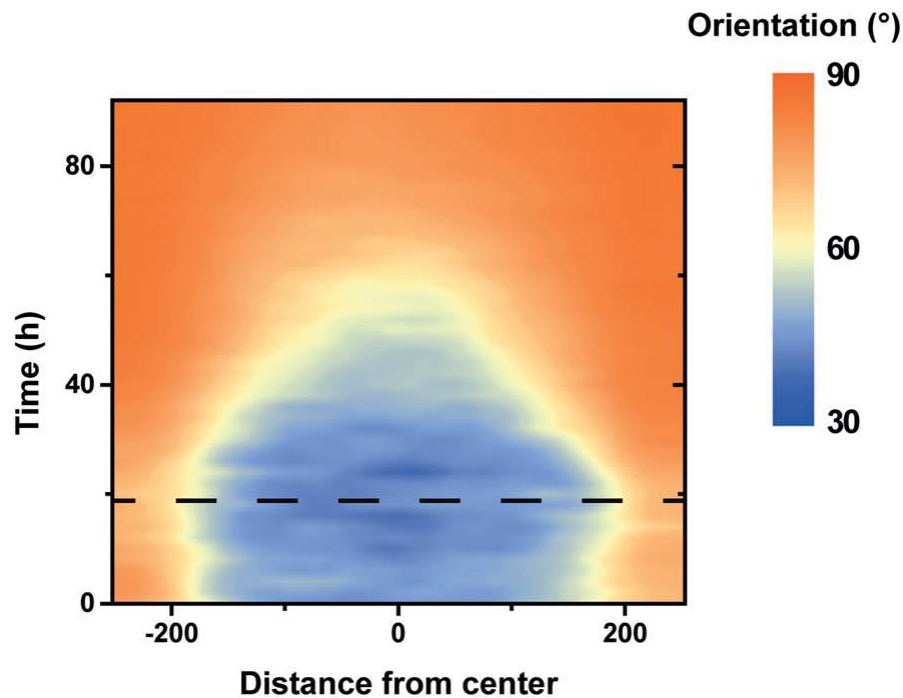


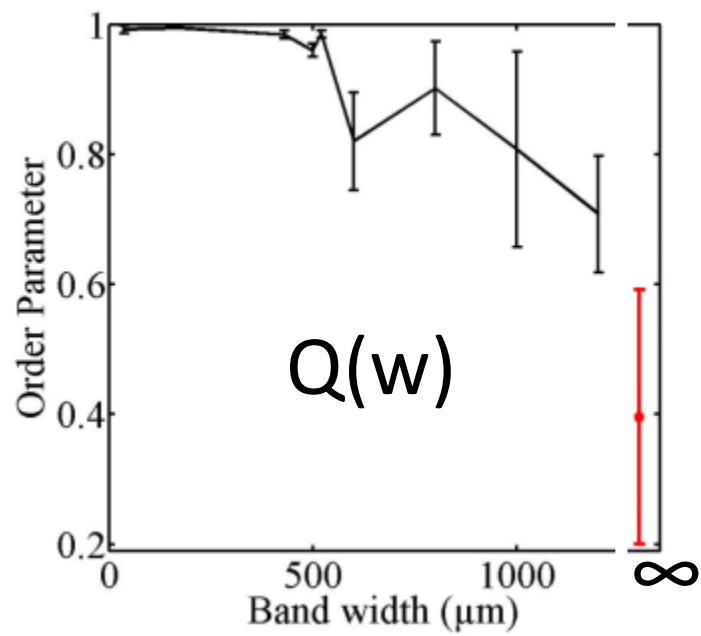
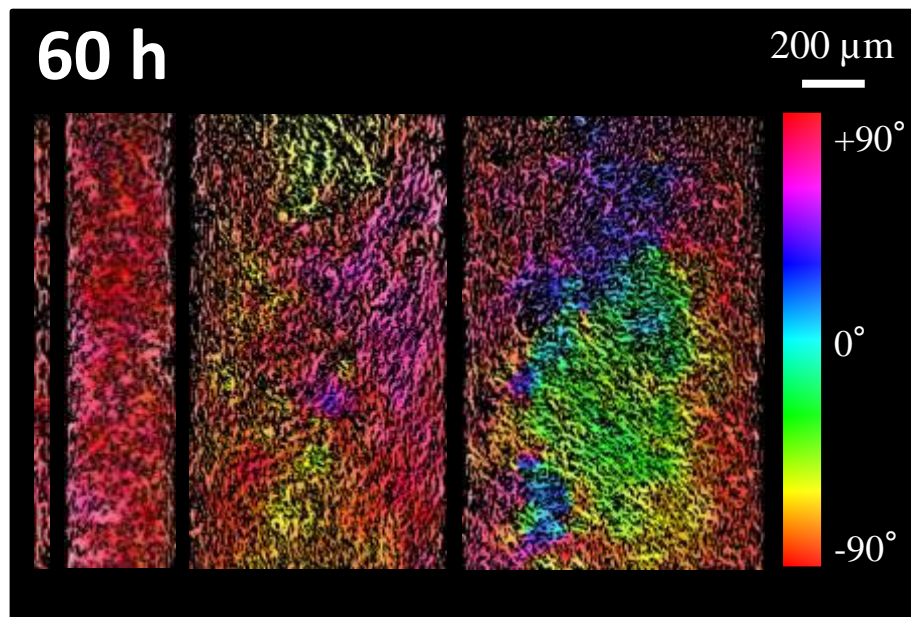
$Q \approx 0.98$

- Cells align at borders
- After confluence, this order propagates from the edges toward the center.



- Cells align at borders
- After confluence, this order propagates from the edges toward the center.



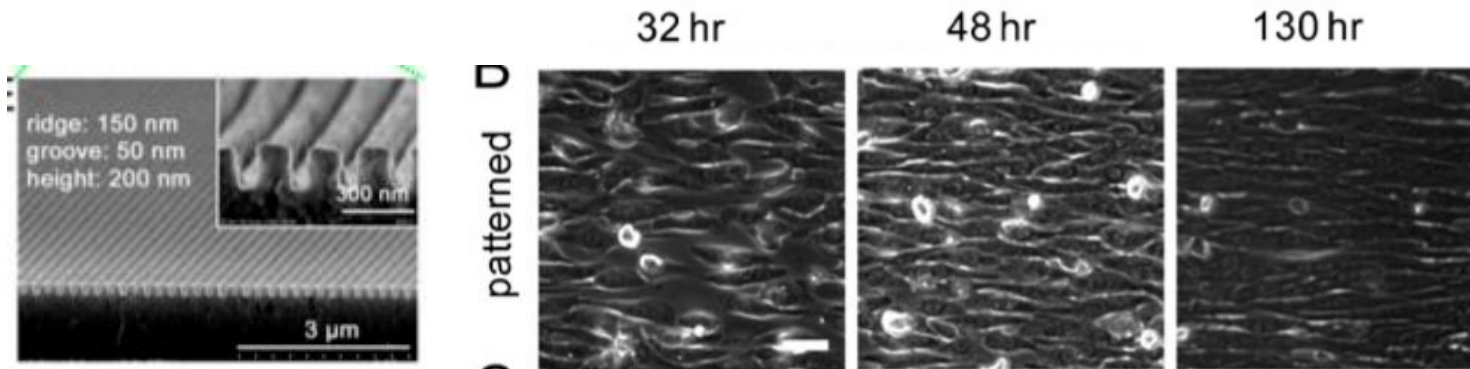


Population-scale vs. individual-scale template

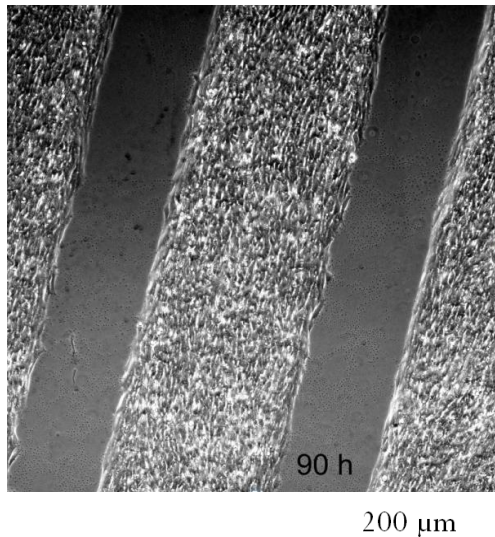
Nanoscale cues regulate the structure and function of macroscopic cardiac tissue constructs

PNAS | January 12, 2010 | vol. 107 | no. 2 | 565-570

Deok-Ho Kim^{a,b,1}, Elizabeth A. Lipke^{a,1}, Pilnam Kim^c, Raymond Cheong^{a,b}, Susan Thompson^a, Michael Delannoy^d, Kahp-Yang Suh^{c,2}, Leslie Tung^{a,2}, and Andre Levchenko^{a,b,2}



Micro-nano tracks



Mesoscopic pattern

Mesoscale contact guidance

Topological defects

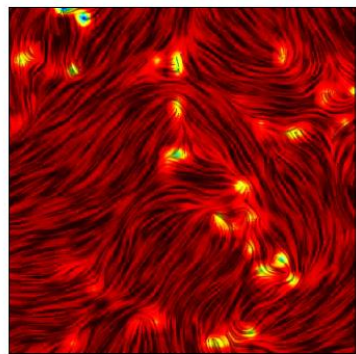
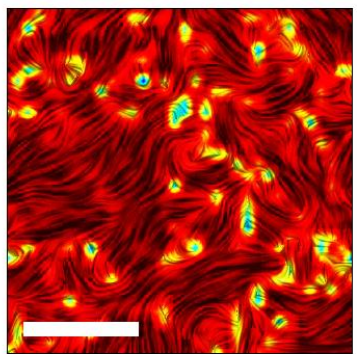
Duclos et al. Nat Phys 2017

Defect density decreases with time

Local order parameter

t = 10h

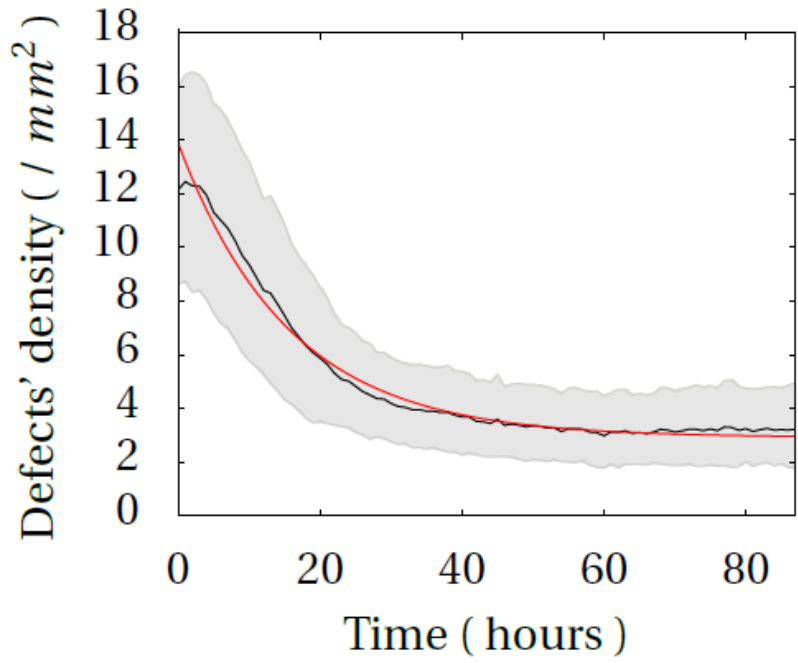
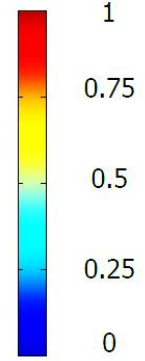
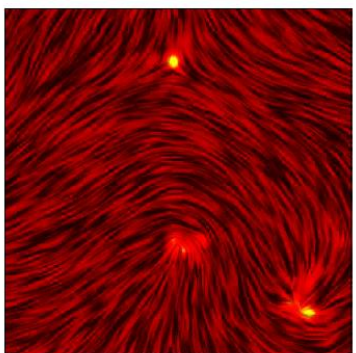
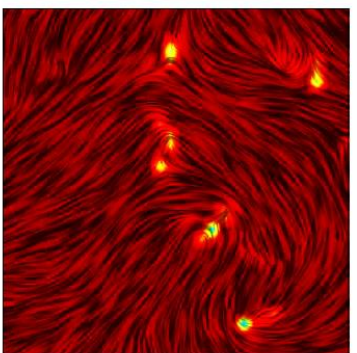
t = 20h



500 μm

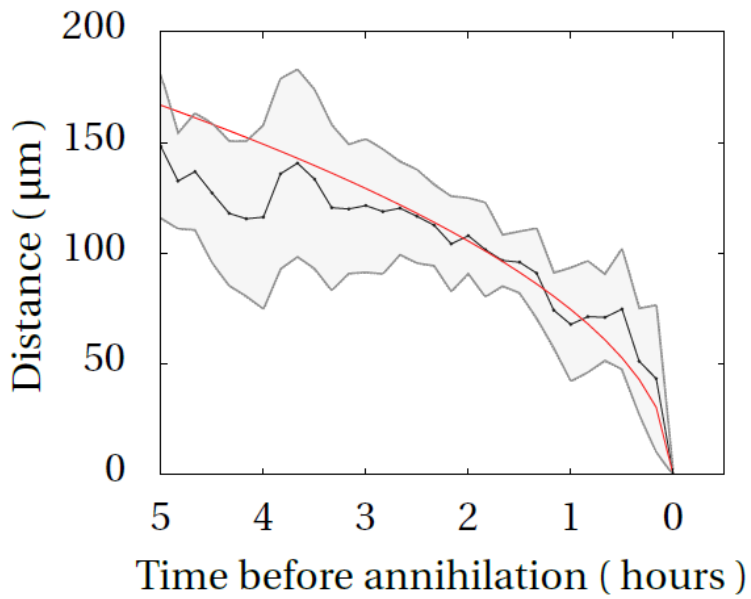
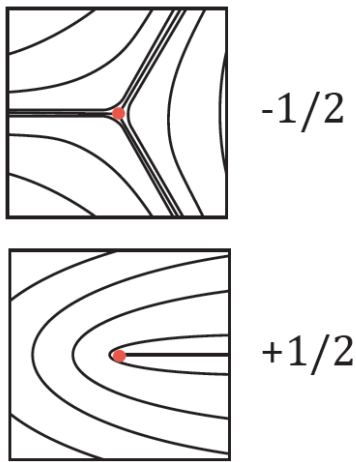
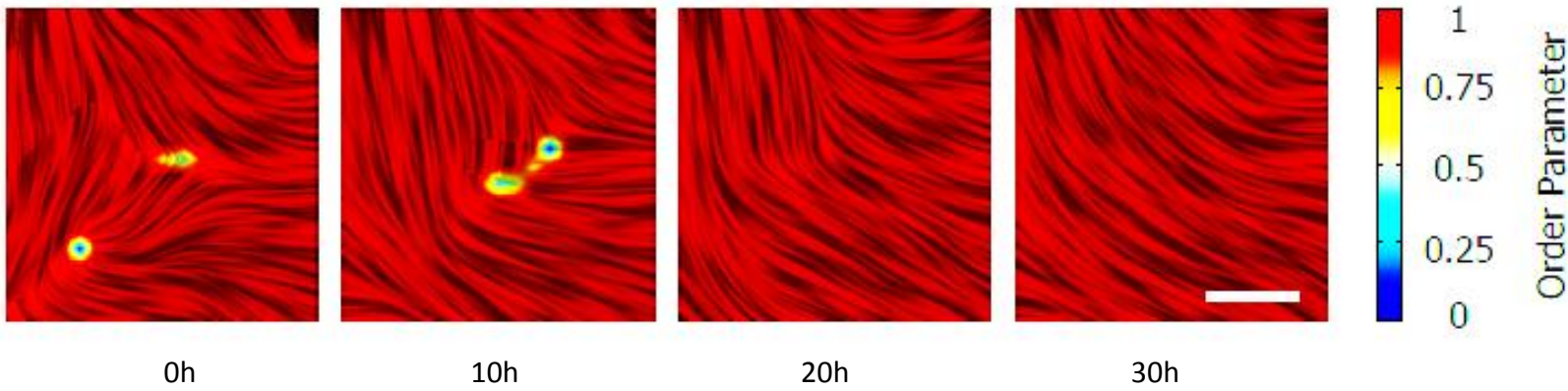
t = 30h

t = 60h



No defect creation
(consistent with contractile system)

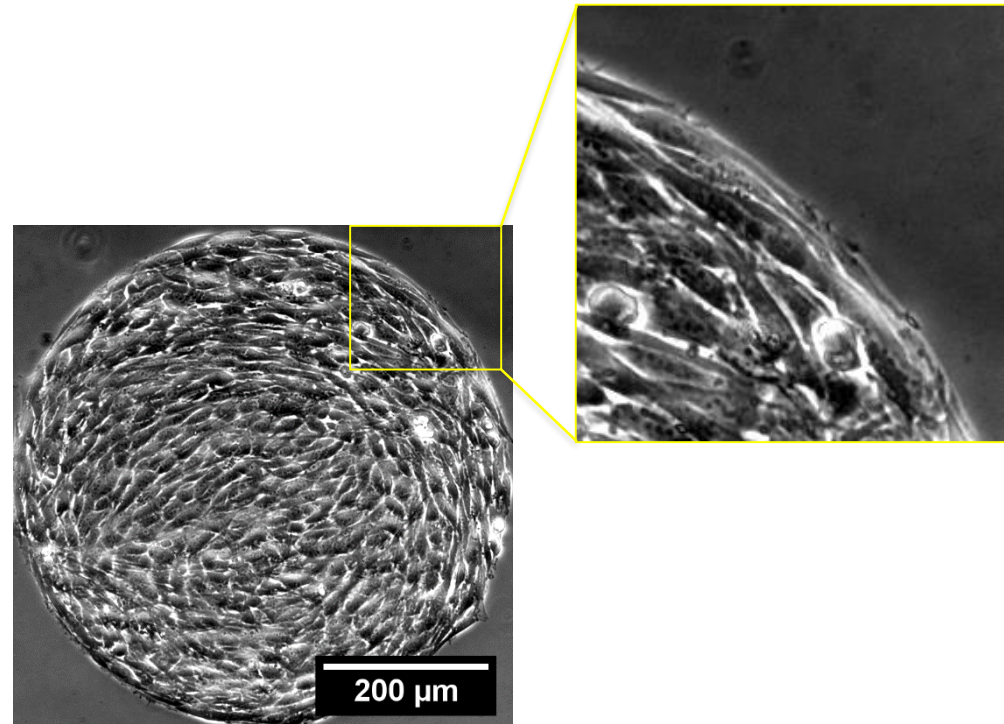
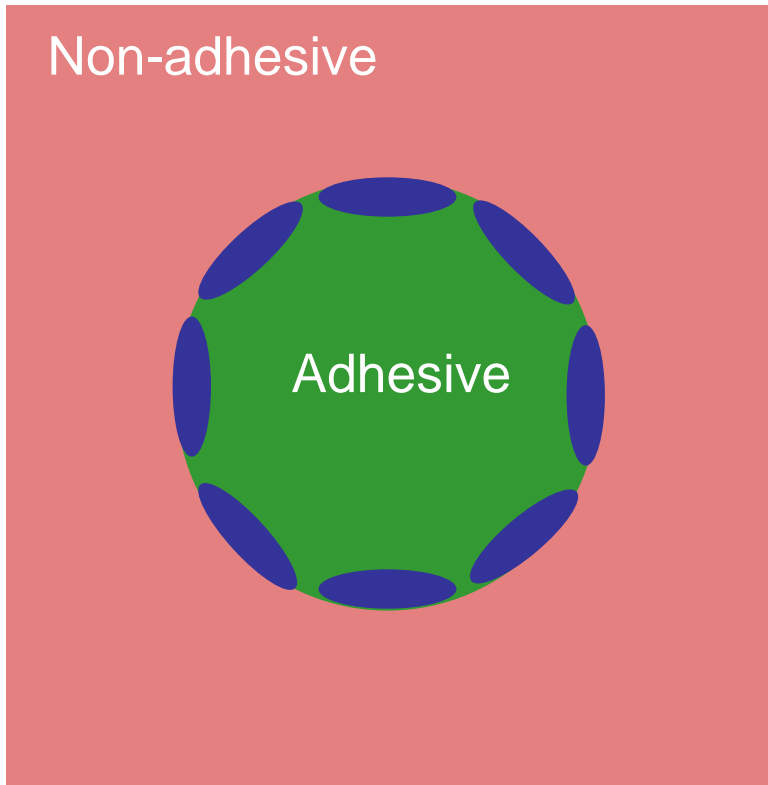
Pairwise defects annihilation



$$d \propto \sqrt{t}$$

(like passive LC)

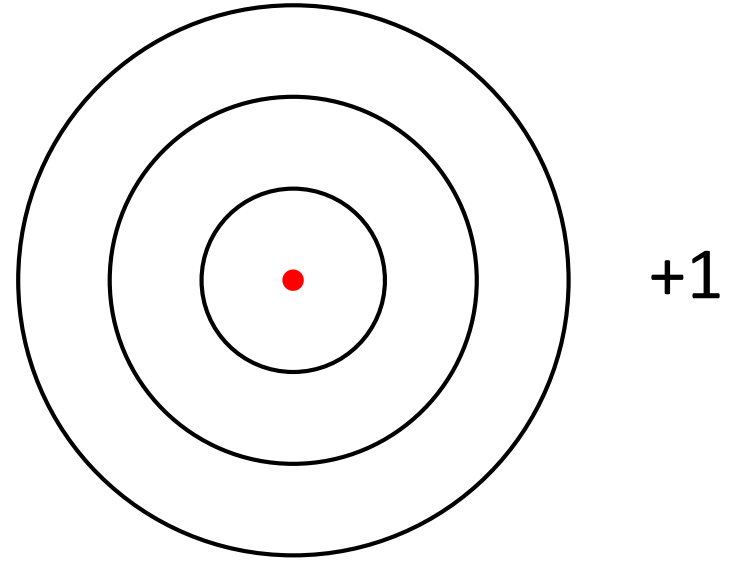
Confinement in micropatterned circular domains



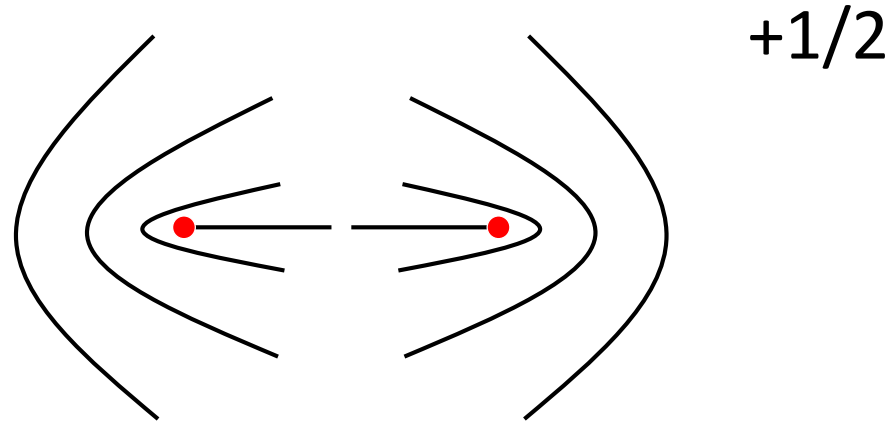
$R = 250 \mu\text{m} - 400 \mu\text{m}$

➔ Total charge = +1

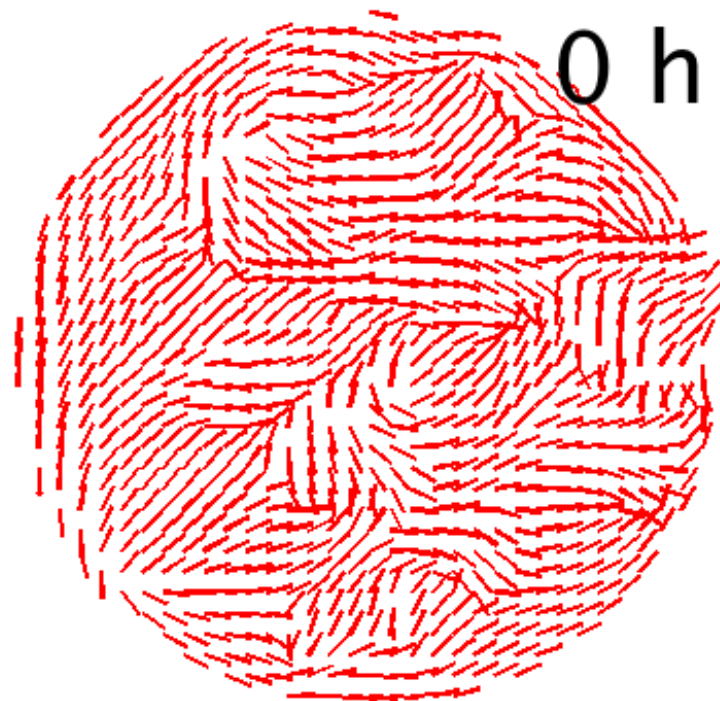
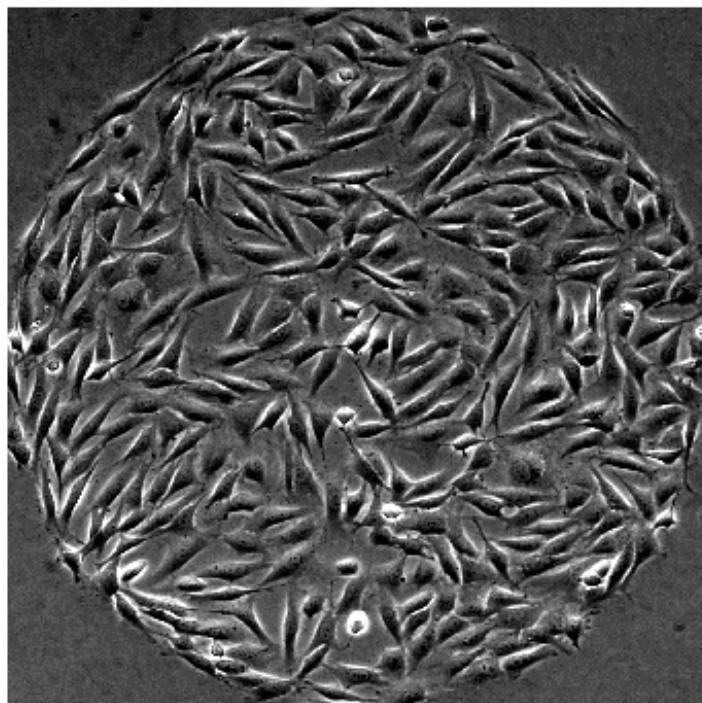
A central defect ...



... or two ?

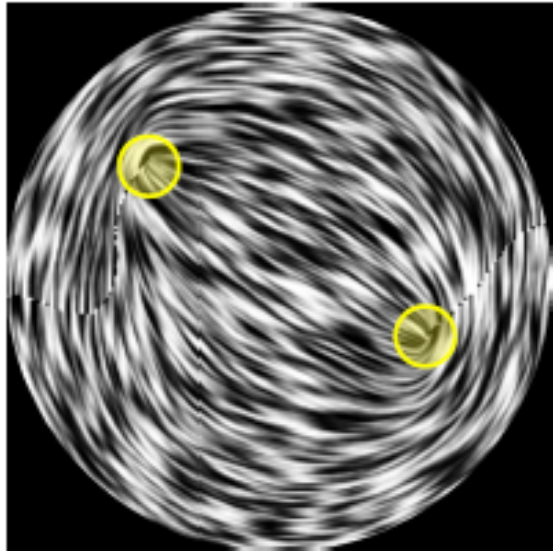


In 2D, $+1$ defects are not stable and give rise to two $+1/2$ defects

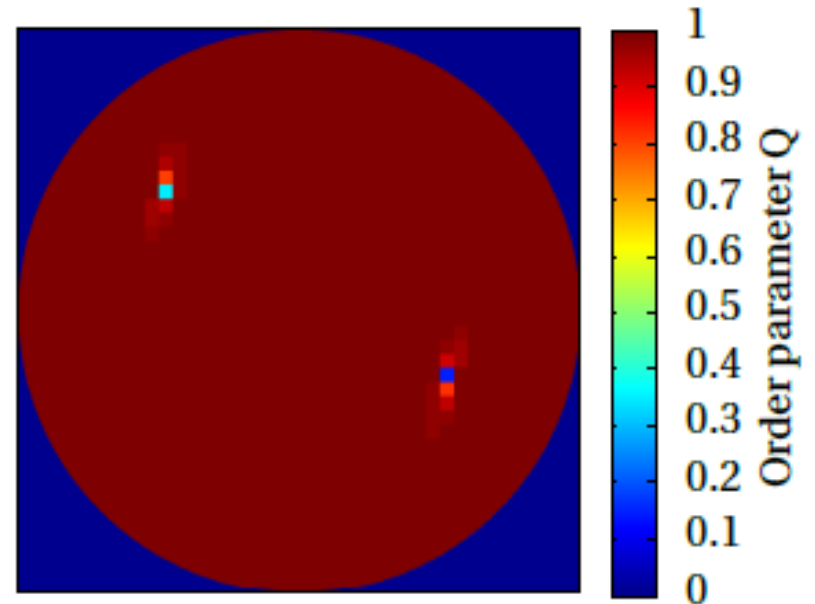


200 μm

Stationary state, 60h after confluence: two $+1/2$ defects



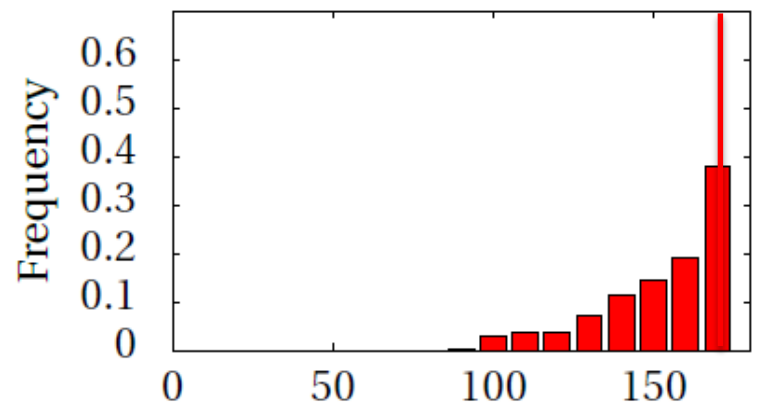
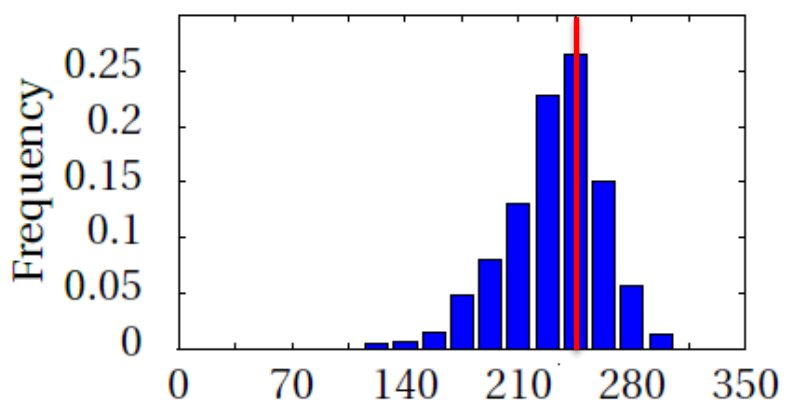
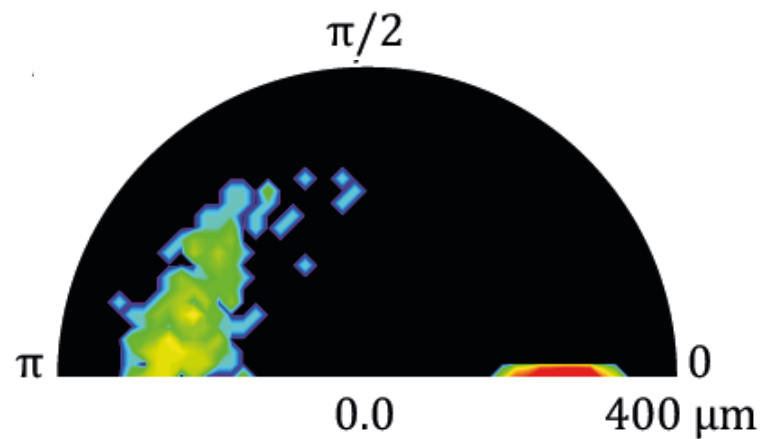
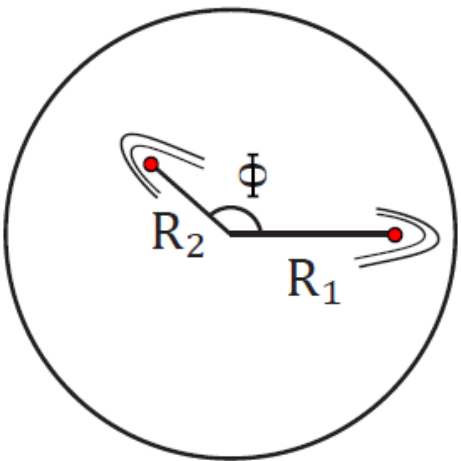
200 μm



Defects' positioning

Steady state - $R = 350 \mu\text{m}$

$R_1, R_2 > 0$
 $\Phi \in [0, \pi]$



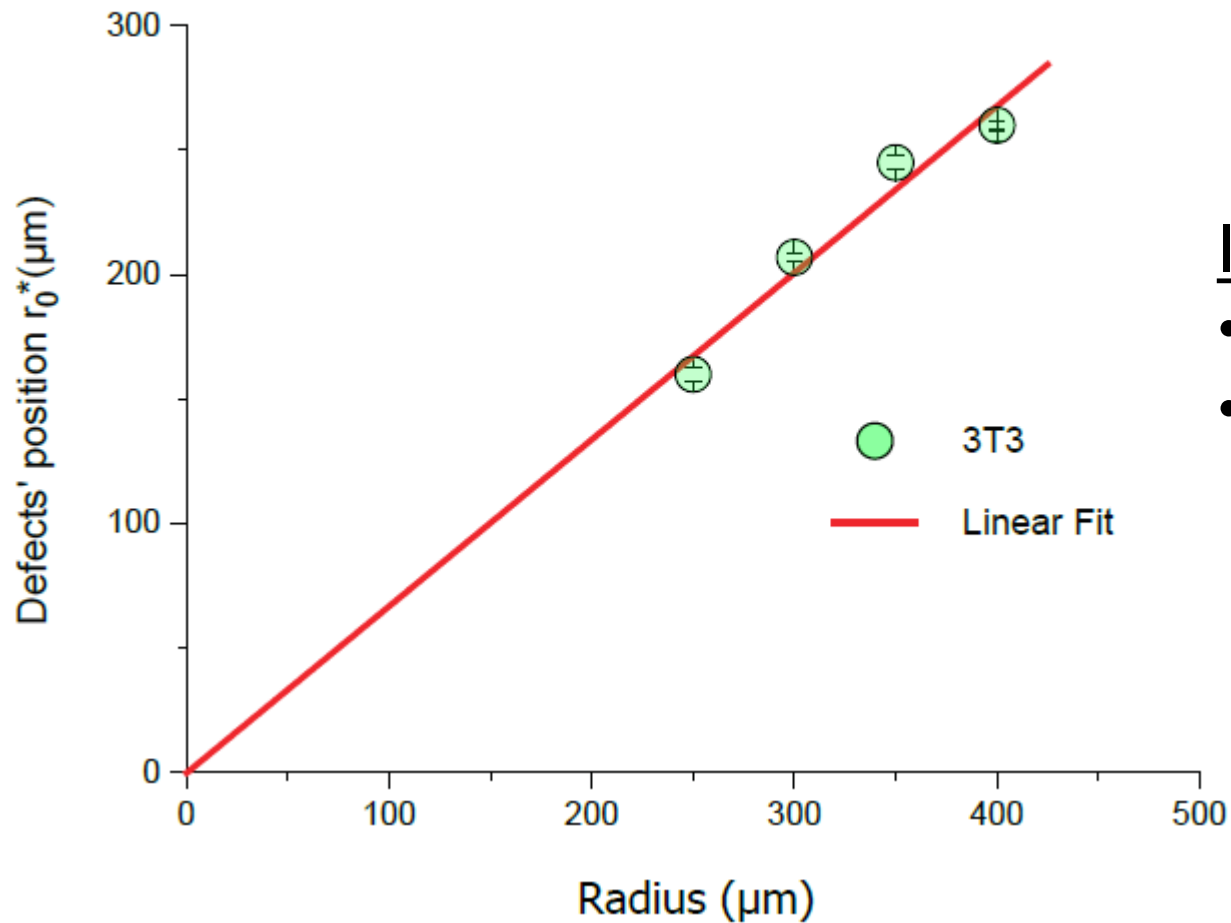
R_1, R_2 (μm) $N=357$

Φ ($^\circ$)

$$R_0^* = 245\mu\text{m} \pm 30\mu\text{m} / \Phi^* = \pi$$

The defects' radial position scales with the domain radius

$$r_0^* = \alpha \cdot R \text{ with } \alpha = 0.67 \pm 0.02$$



Independent of:

- Cell contractility
- Cell type

The nematic disk model

(Christoph Erlenkaemper, Jean-François Joanny)

Hypotheses:

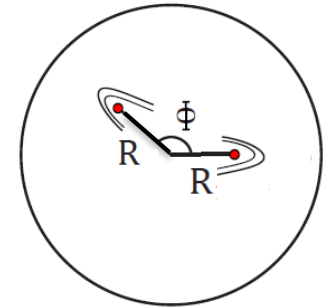
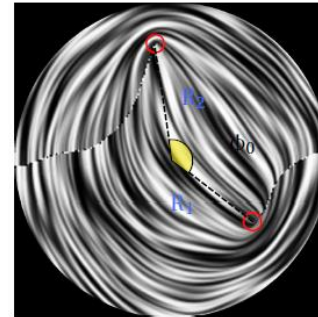
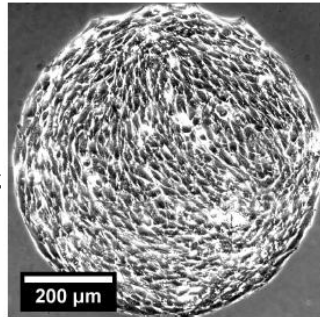
- Parallel alignment at the edges
- $K_1=K_3=K$
- No active stress

2 degrees of freedom

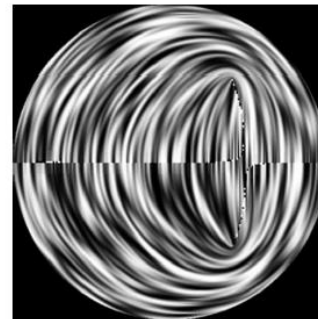
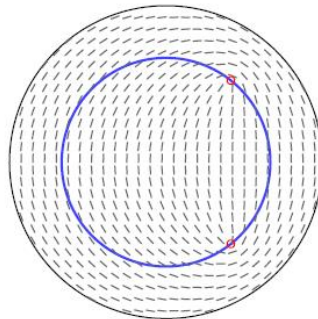
R and Φ

(Defects' positions uncorrelated)

Experimental data



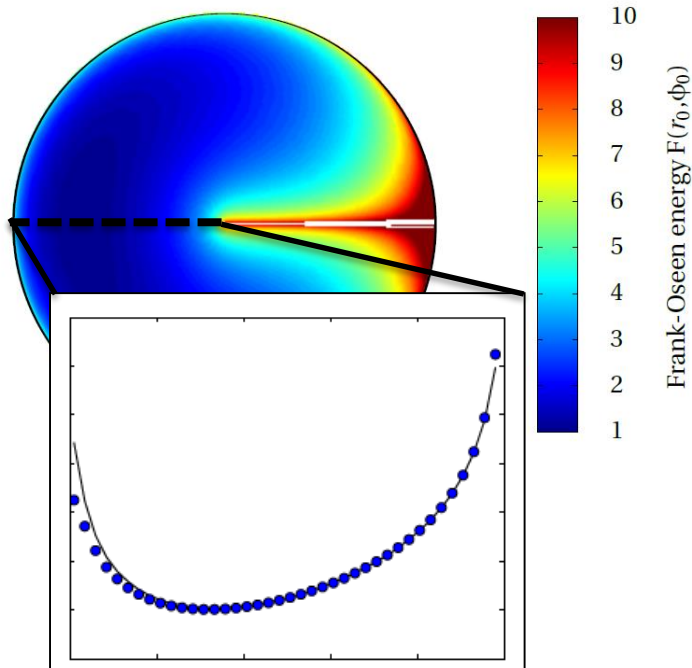
Model



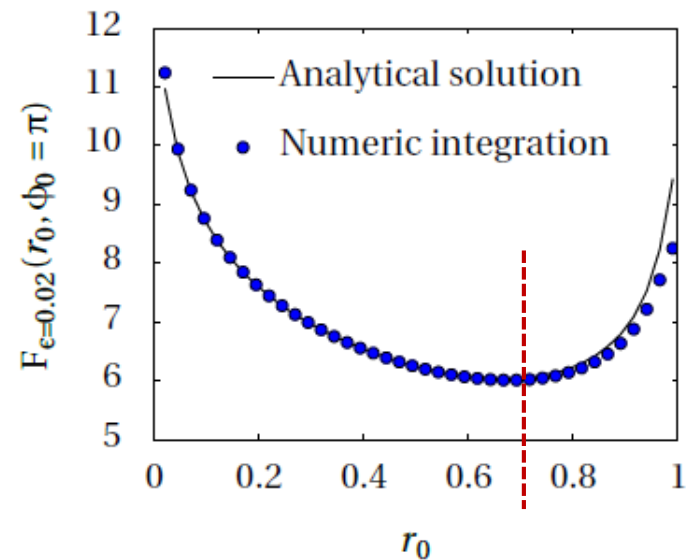
Frank-Oseen free energy

$$F(r_0, \phi_0) = \frac{K}{2} \int_{\Omega} (\nabla \vec{n})^2 dS$$

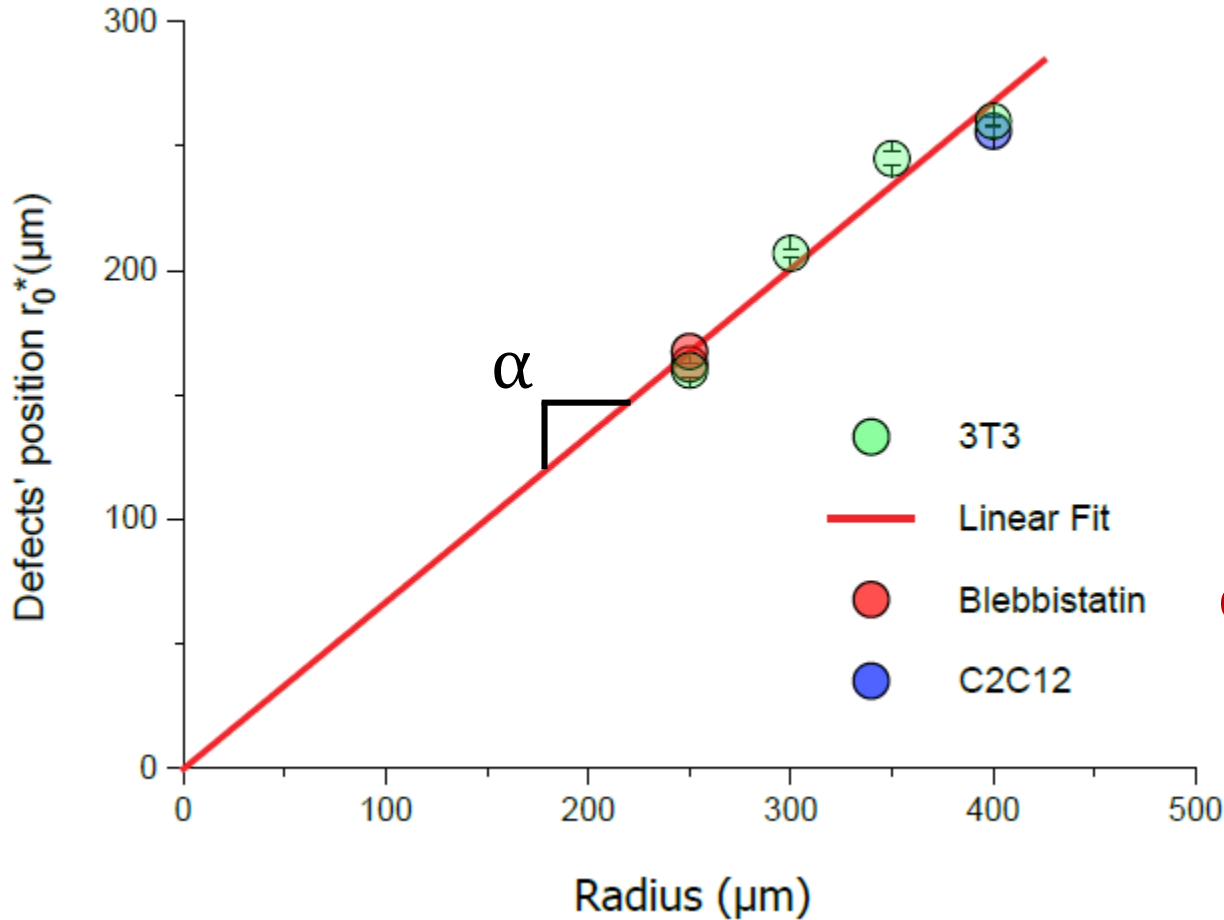
2D Energy map (polar)



1D Energy map ($\Phi=\pi$)



$$r_0^* = 5^{-1/4} \cdot R \approx 0.67 \cdot R$$



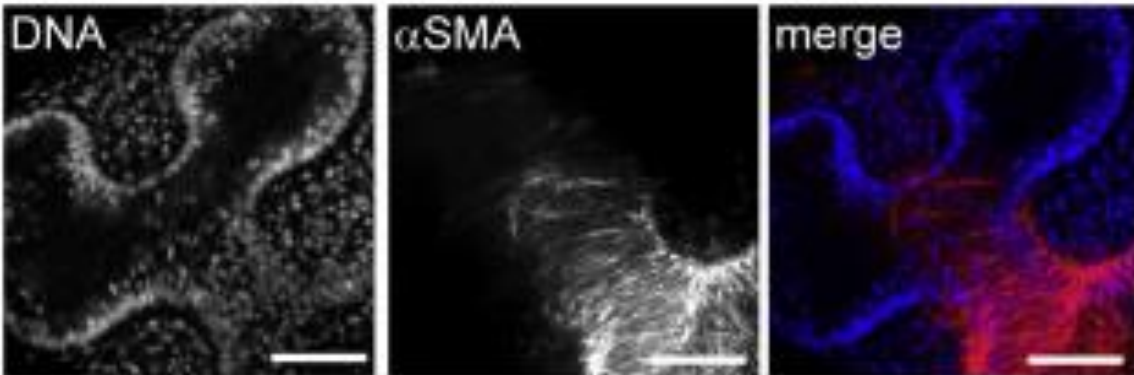
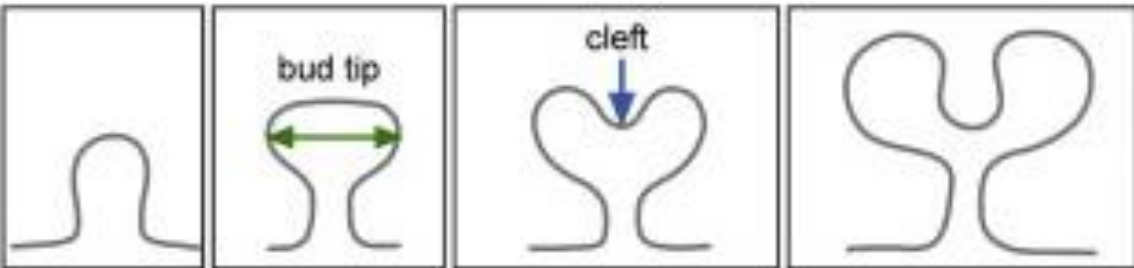
$$\alpha_{\text{exp}} = 0.67 \pm 0.02$$

$$\alpha_{\text{theory}} = 5^{-1/4} \approx 0.668$$

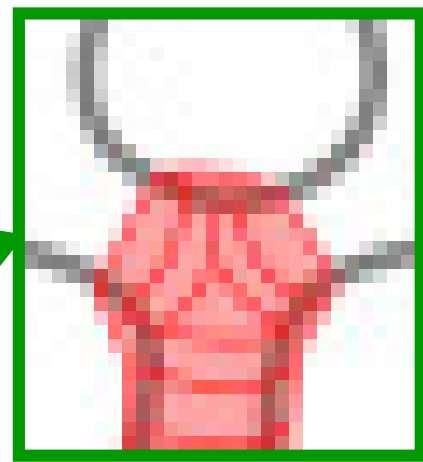
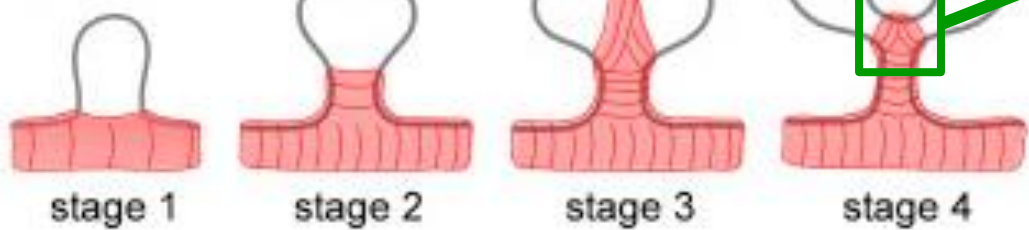
α independant of:

- Activity
- Nematic elasticity K

Epithelial branching (lung) (C. Nelson)



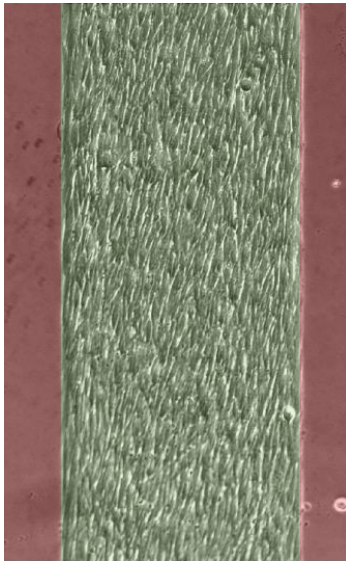
smooth muscle
epithelium



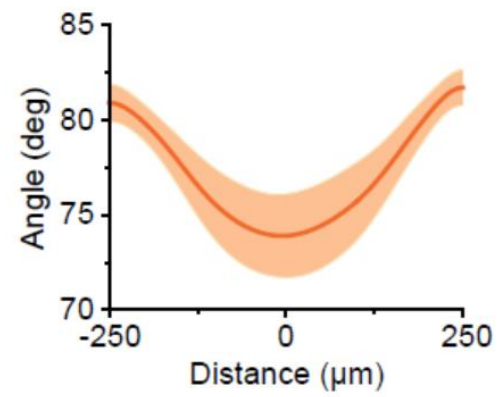
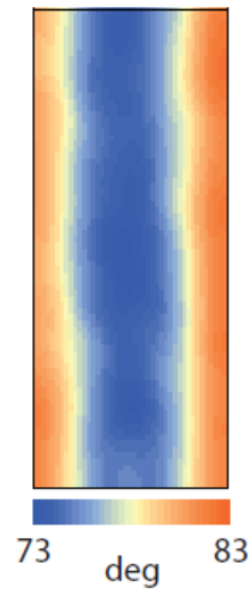
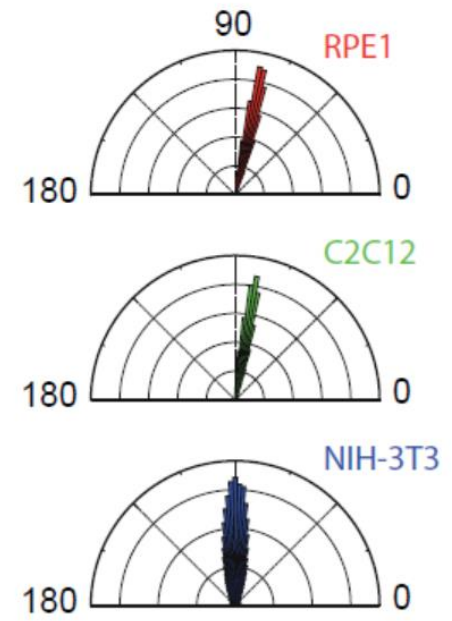
Spontaneous shear flows

Spontaneous tilt

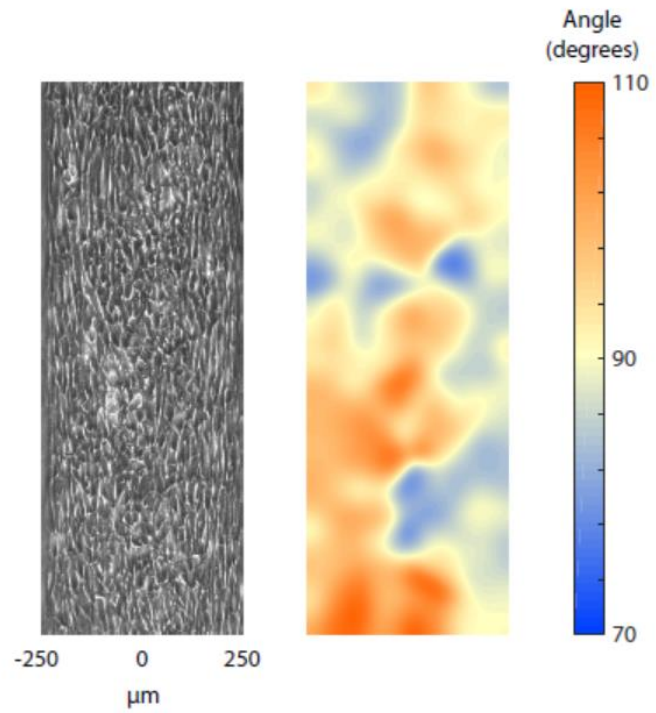
RPE1
C2C12



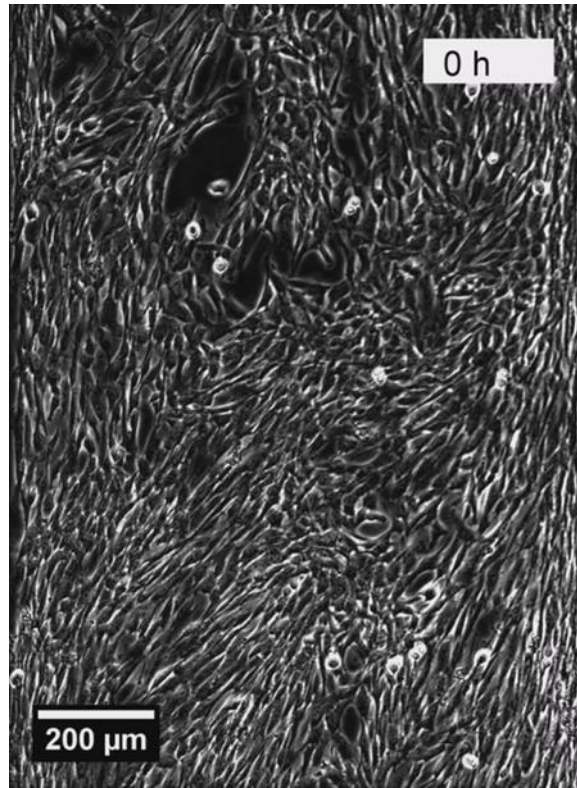
500 μm



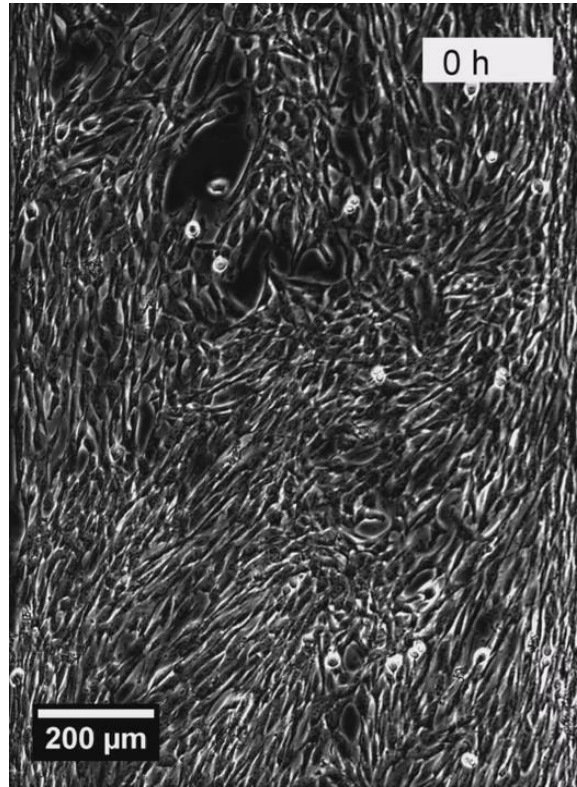
3T3



The dynamics: Spontaneous Flows



Spontaneous Flows



Supplementary Movie 4

Intra-strand dynamics of file-like collective invasion

Excitation: 850nm (25mW) and 910nm (30mW)

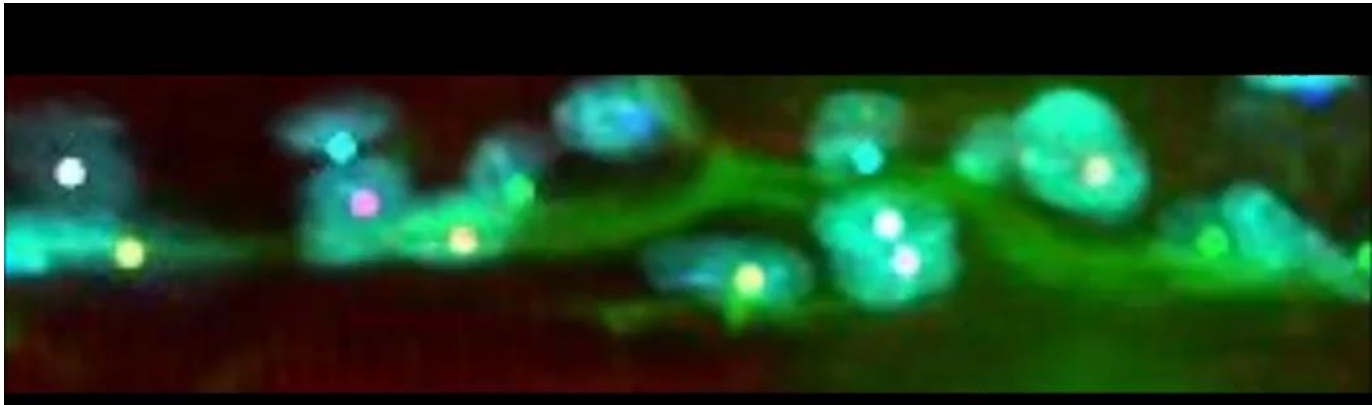
Frame rate: 1 frame / 10min

Total time: 70min

SHG (red): Collagen fibers, striated muscles

Histone-2B/eGFP (blue): Tumor cell nuclei

TM-Rhodamine (green): 70kDa-dextran labeling blood vessels, phagocytes



Supplementary Movie 4

Intra-strand dynamics of file-like collective invasion

Excitation: 850nm (25mW) and 910nm (30mW)

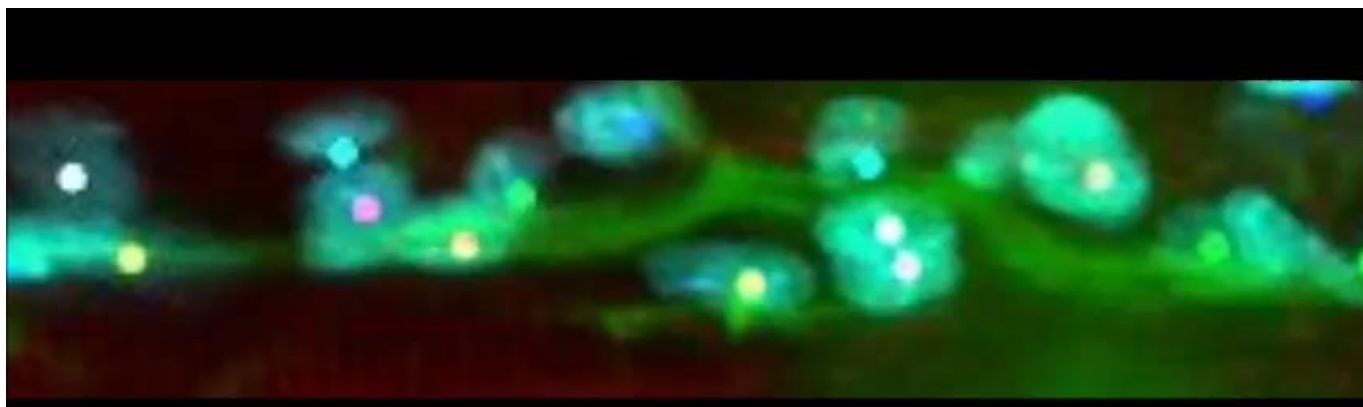
Frame rate: 1 frame / 10min

Total time: 70min

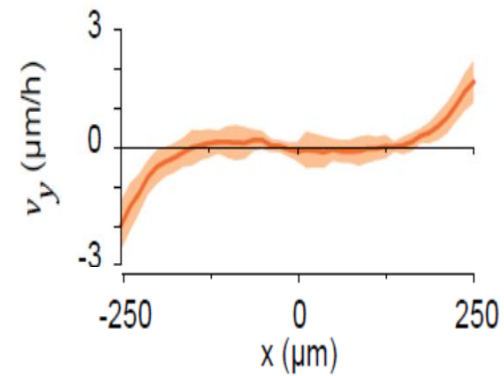
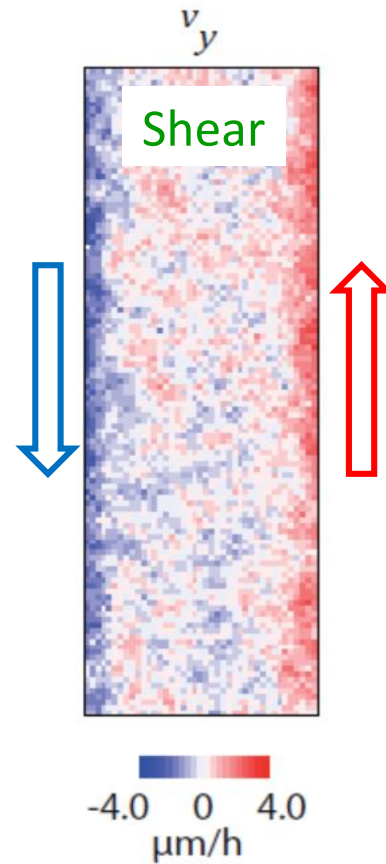
SHG (red): Collagen fibers, striated muscles

Histone-2B/eGFP (blue): Tumor cell nuclei

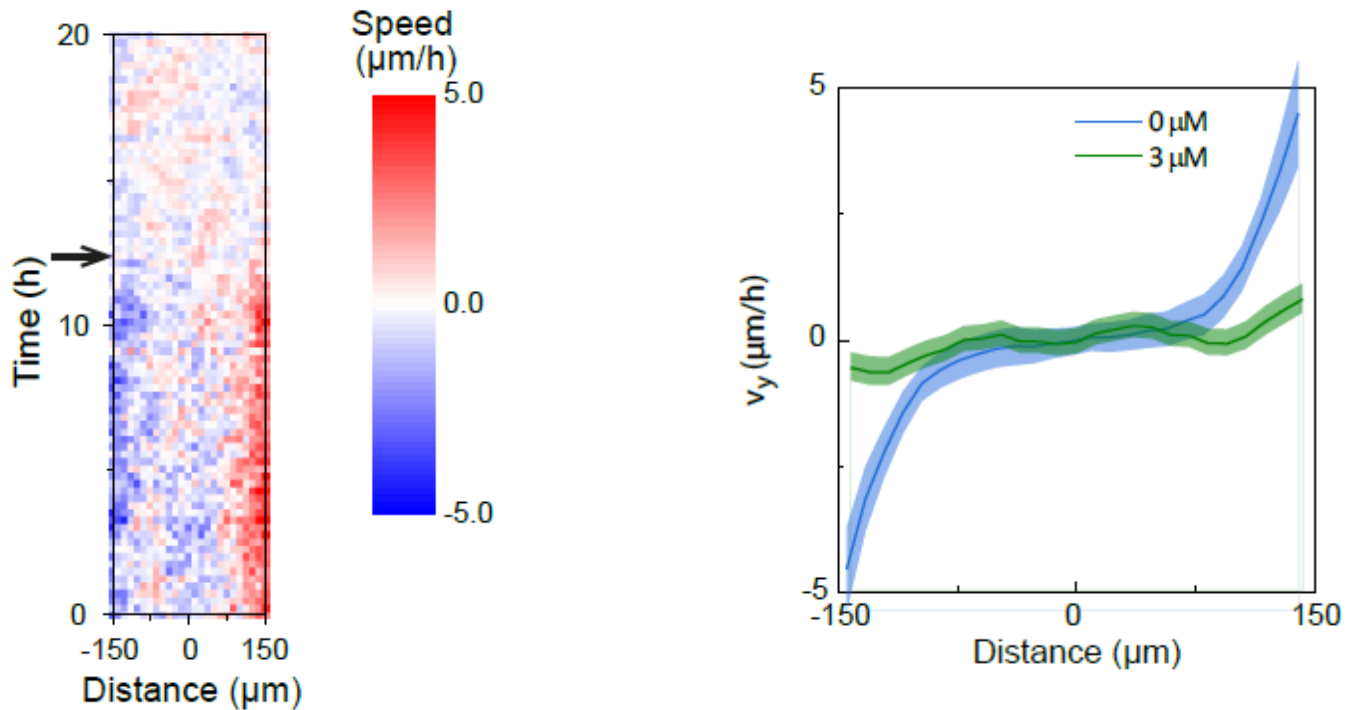
TM-Rhodamine (green): 70kDa-dextran labeling blood vessels, phagocytes



Spontaneous Flows



Inhibiting cell activity (blebbistatin)



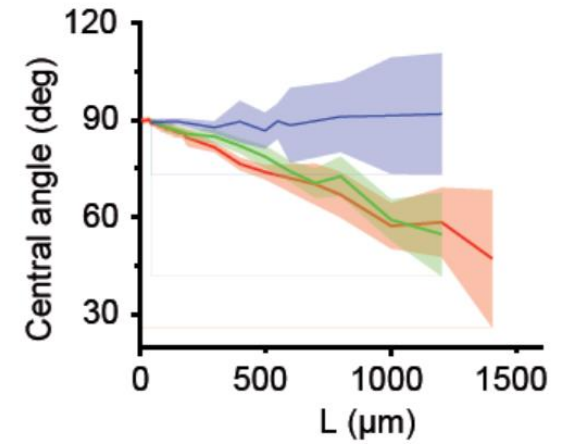
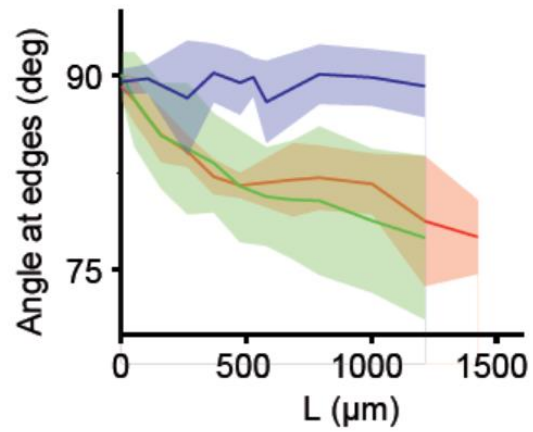
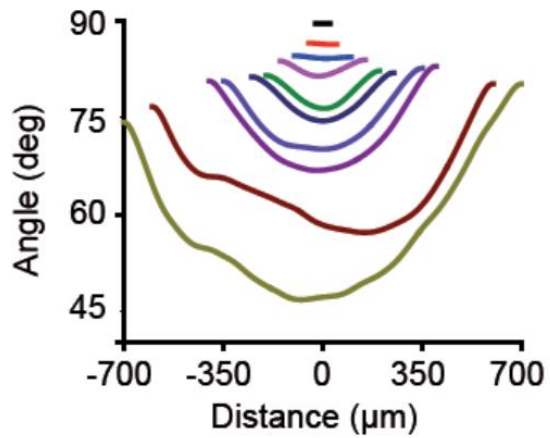
- Shear flow results from cell activity

Width dependence

NIH 3T3

C2C12

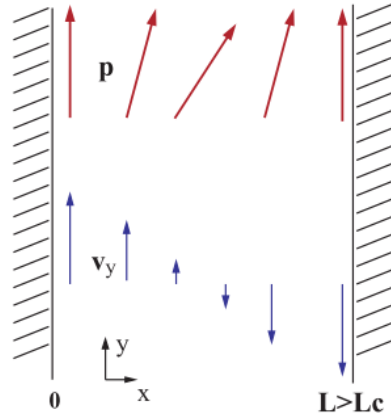
RPE1



Active gel theory

Spontaneous flows in actomyosin systems

(R. Voituriez, J.-F. Joanny, J. Prost EPL 2005)



+

New boundary conditions to account for finite edge angle:

$$\pm K \partial_x \theta + W_s \left(\theta - \frac{\pi}{2} \right) = 0 \Big|_{x=\pm L/2}$$

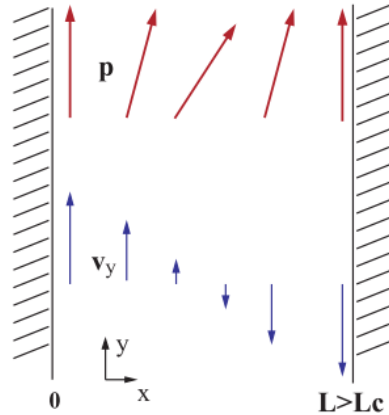
$$\partial_t \theta = \frac{h_{\perp}}{\gamma} \frac{(4\eta + \gamma(\nu^2 - 2\nu \cos 2\theta + 1))}{(4\eta + \gamma\nu^2 \sin 2\theta^2)} - \frac{\zeta \Delta \mu \sin 2\theta (\nu \cos 2\theta - 1)}{(4\eta + \gamma\nu^2 \sin 2\theta^2)} \quad \text{with} \quad h_{\perp} = K \partial_x^2 \theta.$$

Abstract. – We study theoretically the effects of confinement on active polar gels such as the actin network of eukaryotic cells. Using generalized hydrodynamics equations derived for active gels, we predict, in the case of quasi-one-dimensional geometry, a spontaneous flow transition from a homogeneously polarized immobile state for small thicknesses, to a perturbed flowing state for larger thicknesses. The transition is not driven by an external field but by the activity of the system. We suggest several possible experimental realizations.

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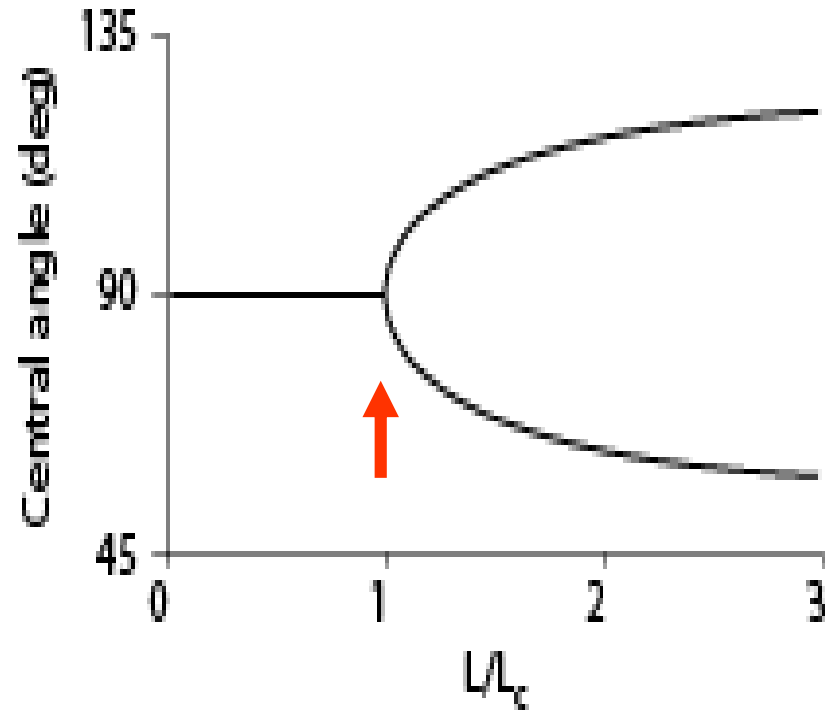
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Abstract. – We study theoretically the effects of confinement on active polar gels such as the actin network of eukaryotic cells. Using generalized hydrodynamics equations derived for active gels, we predict, in the case of quasi-one-dimensional geometry, a spontaneous flow transition from a homogeneously polarized immobile state for small thicknesses, to a perturbed flowing state for larger thicknesses. The transition is not driven by an external field but by the activity of the system. We suggest several possible experimental realizations.

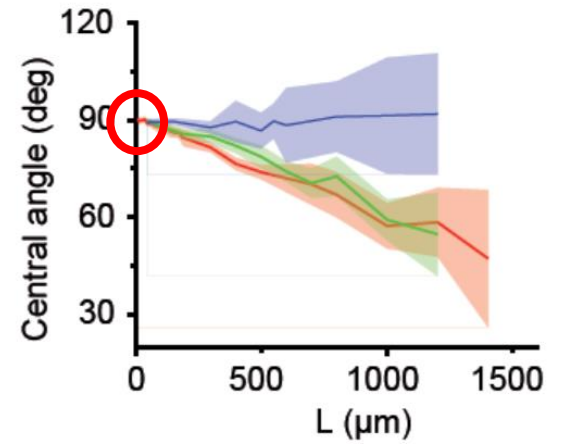
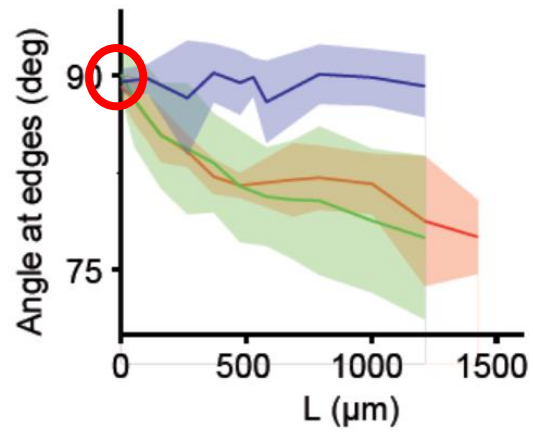
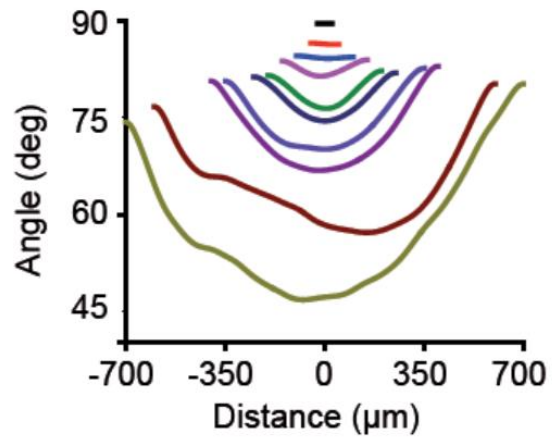
Active gel theory

$$L \gtrsim L_c \rightarrow \theta \sim \sqrt{L - L_c}$$

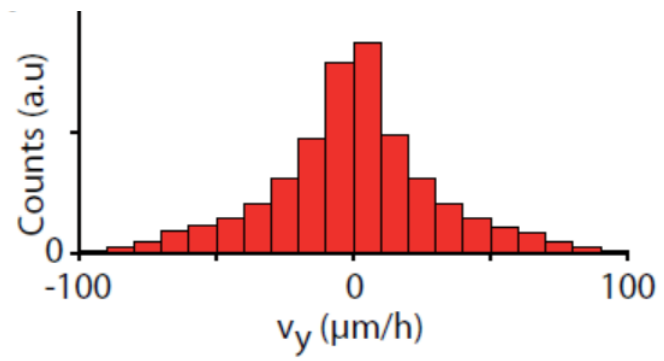
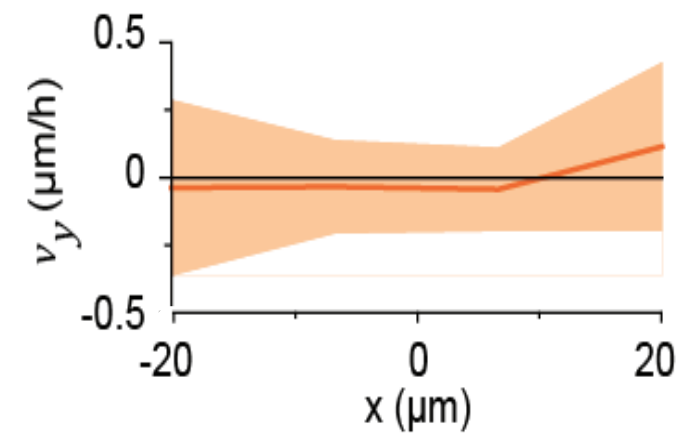
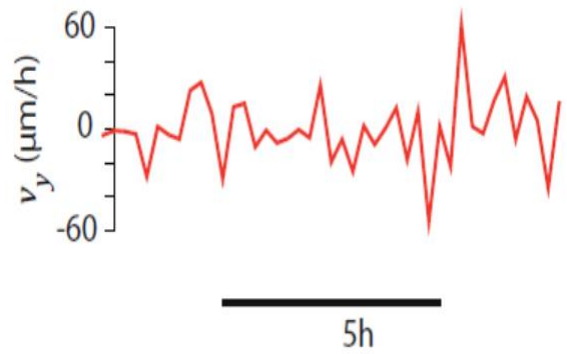
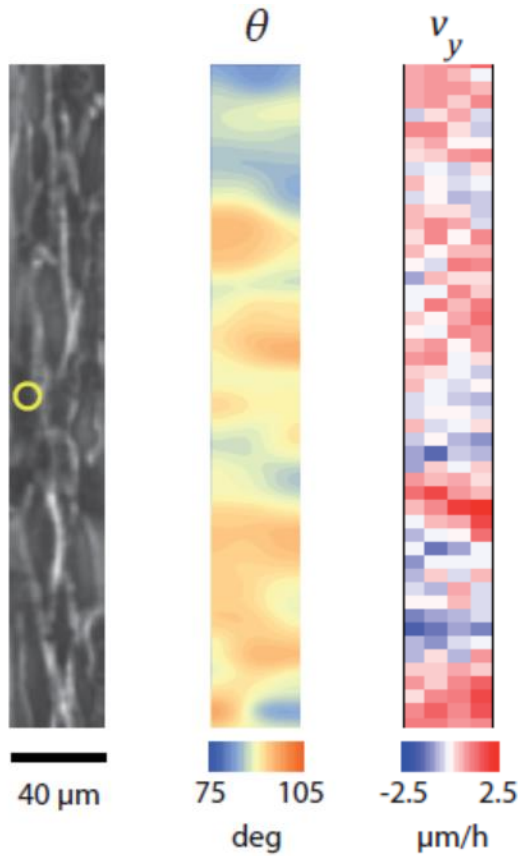


A Fréedericksz transition controlled by the activity of the cells

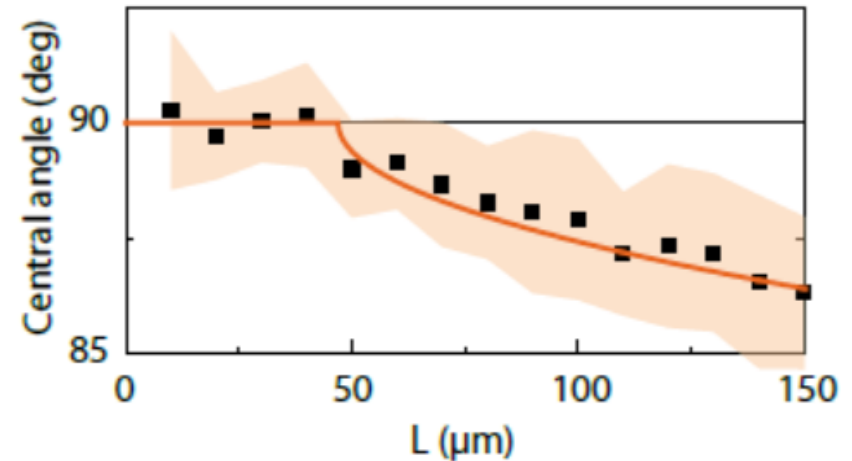
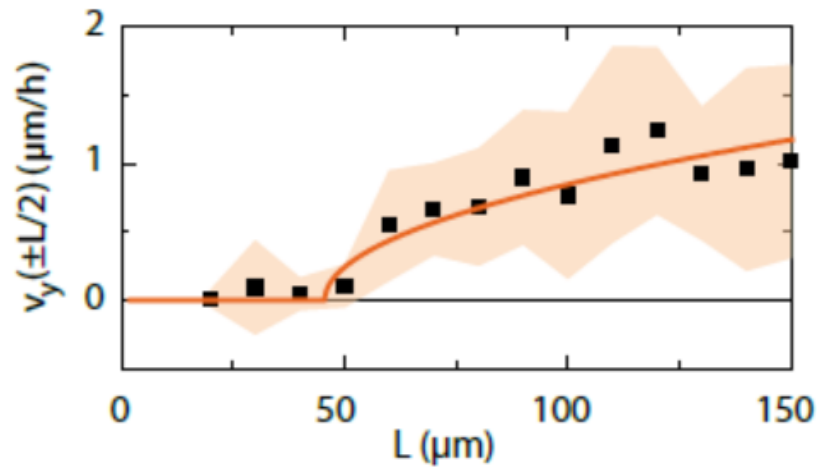
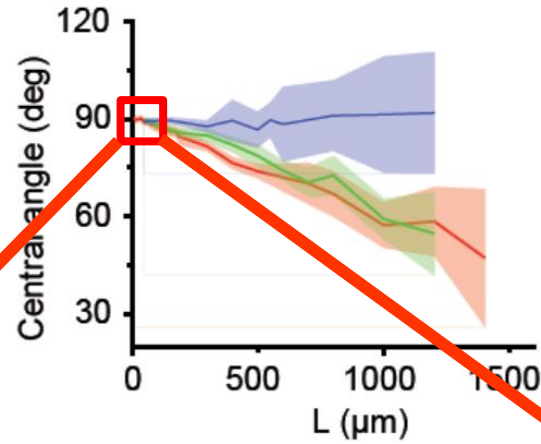
Width dependence



Narrow stripes



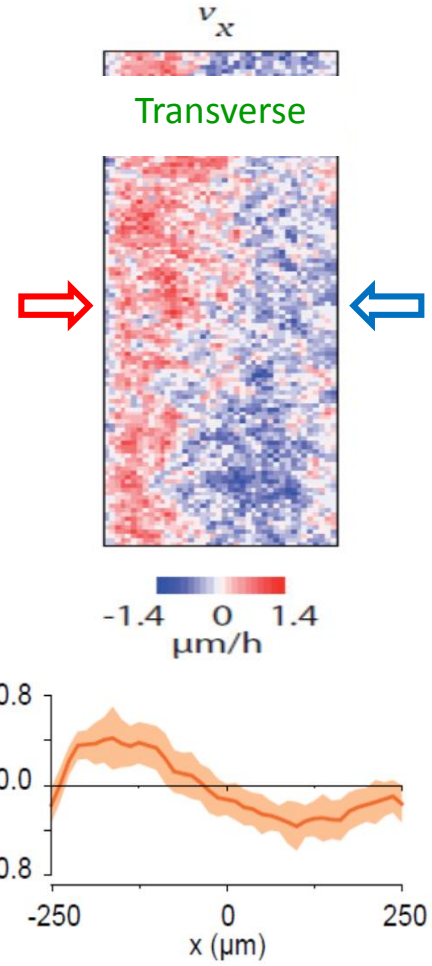
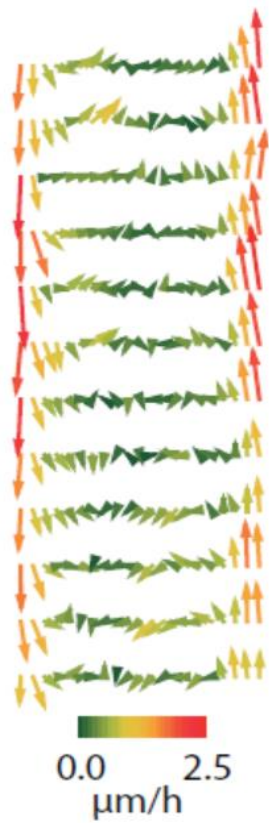
A transition at small widths



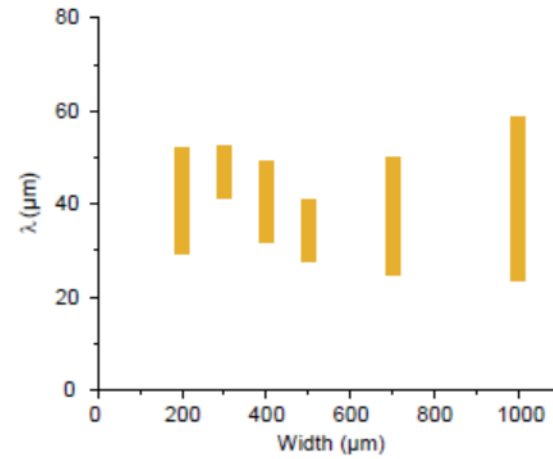
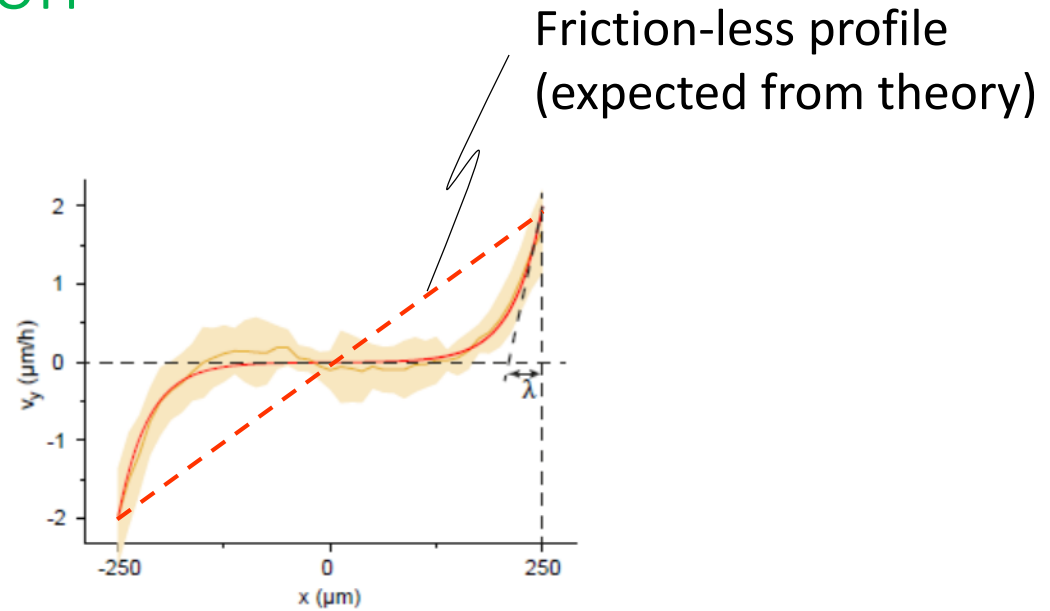
$$L \gtrsim L_c \rightarrow \theta \sim v_y \sim \sqrt{L - L_c}$$

A *Fréedericksz transition* controlled by the activity of the cells

Cell proliferation → Convergent flows

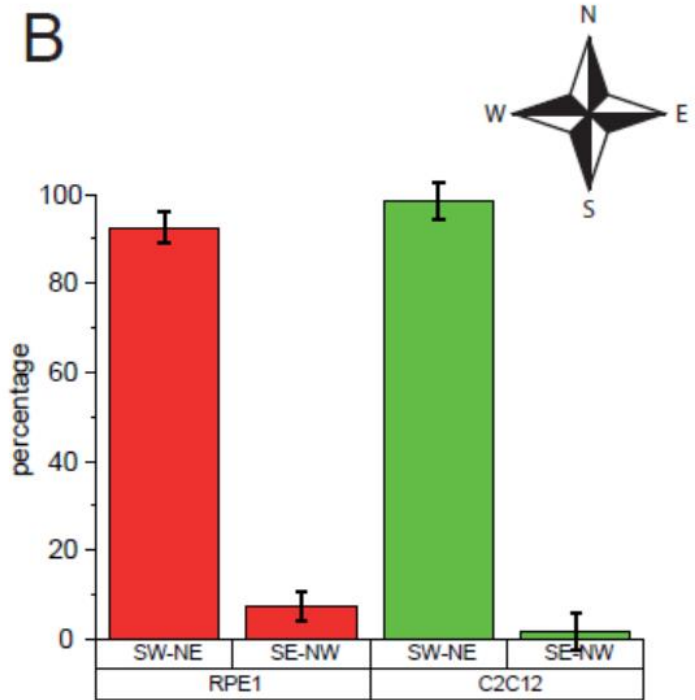
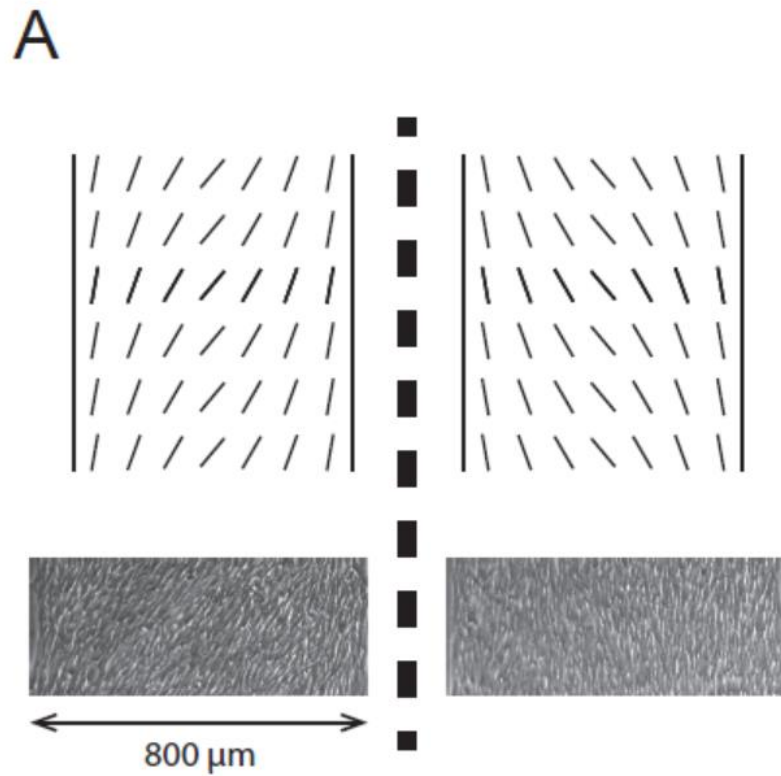


The return of friction



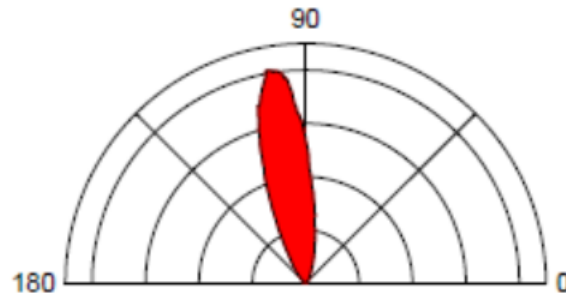
Friction is independent of geometry

A chiral arrangement

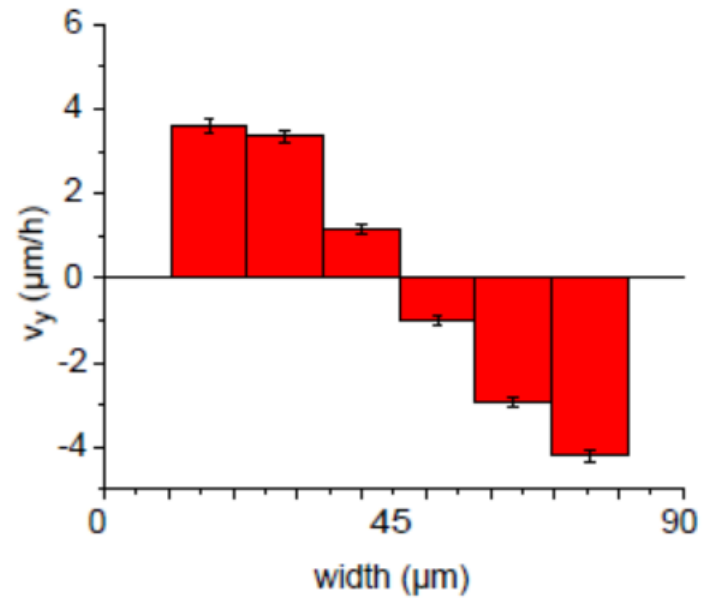


Single cell vs. population-scale chirality: origin ? Amplification ?

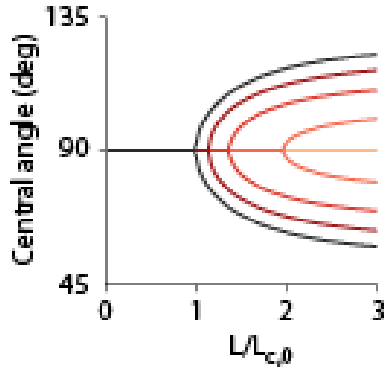
Cancer cells HT1080



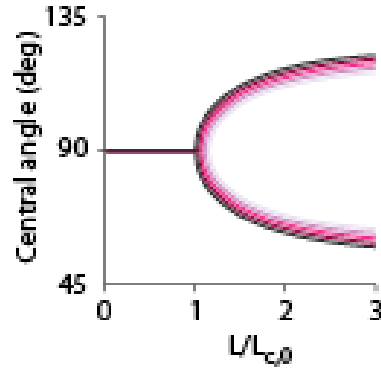
Opposite sign
of the
chirality !



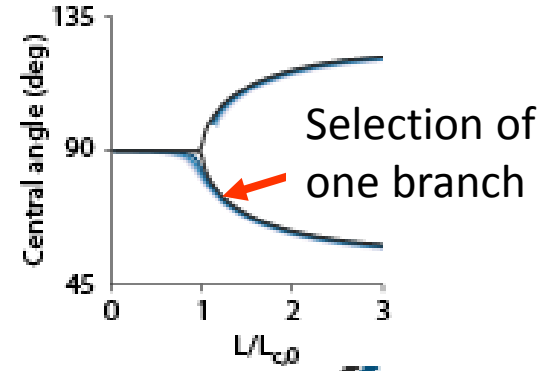
friction



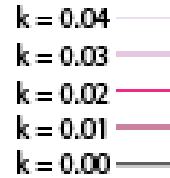
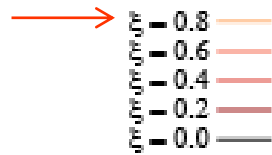
transverse flow



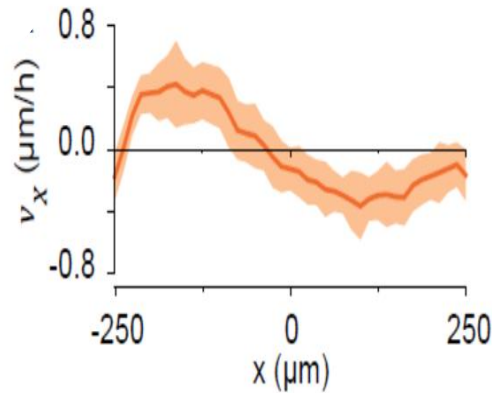
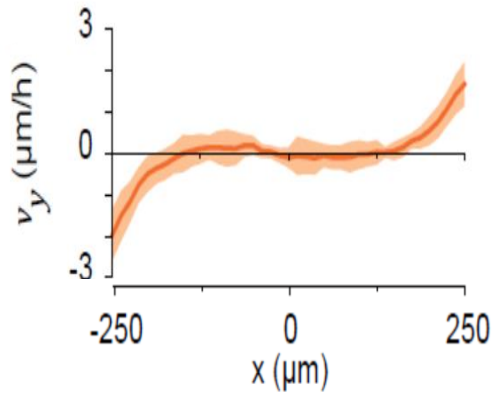
chirality



3T3 (?)



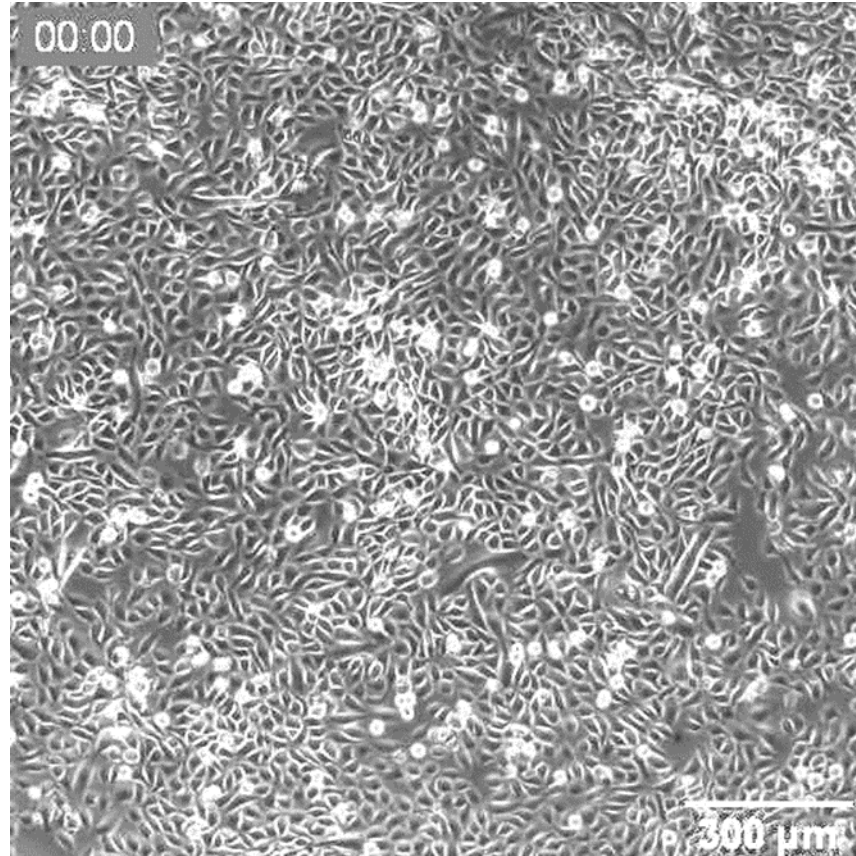
- Controls the dynamics of the director's orientation



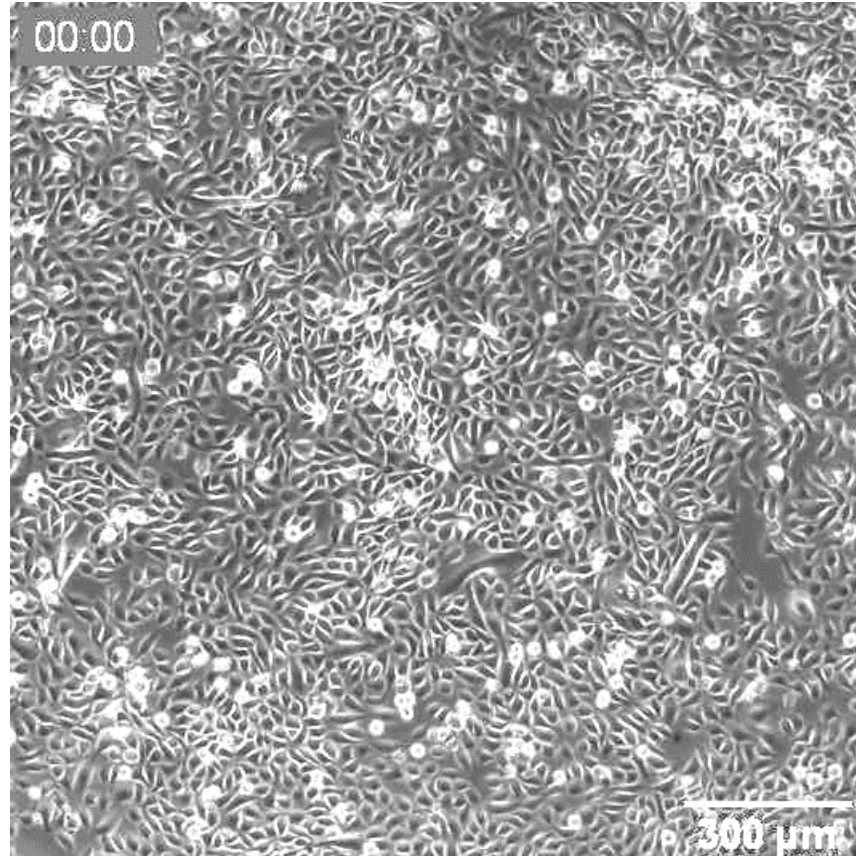
***Cranking up the activity:
Cellular turbulence***

Blanch-Mercader, Yashunsky et al. PRL 2019

HBE cells (Human Bronchial Epithelial)

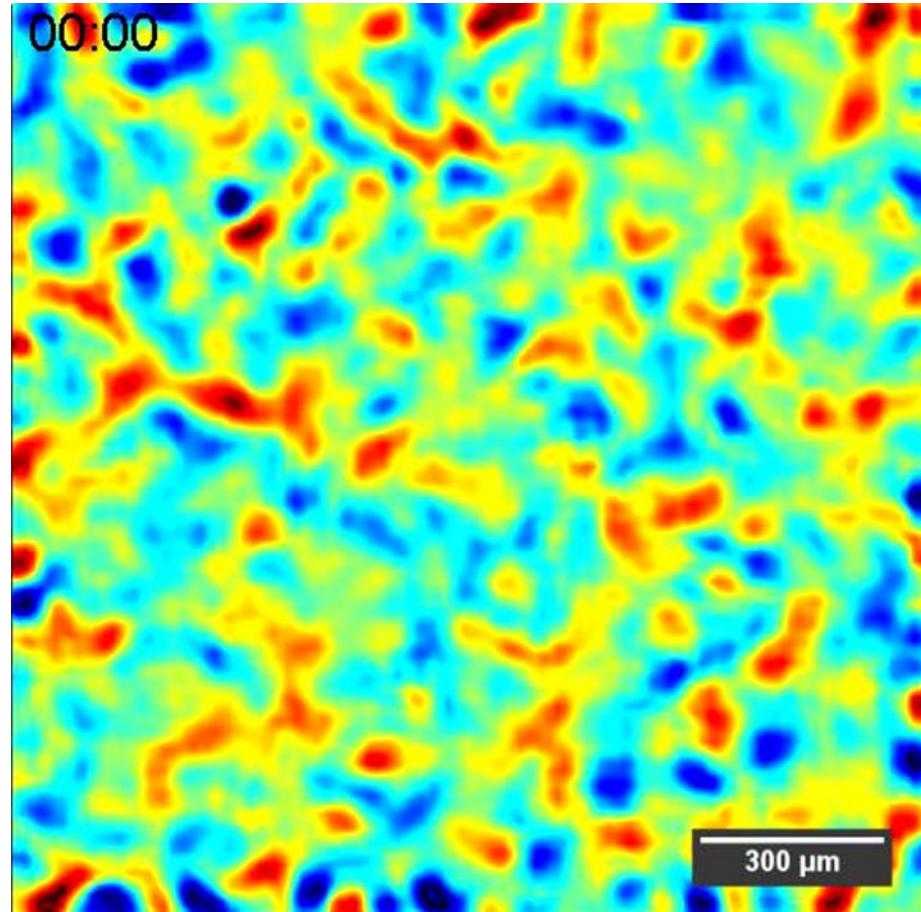


HBE cells (Human Bronchial Epithelial)



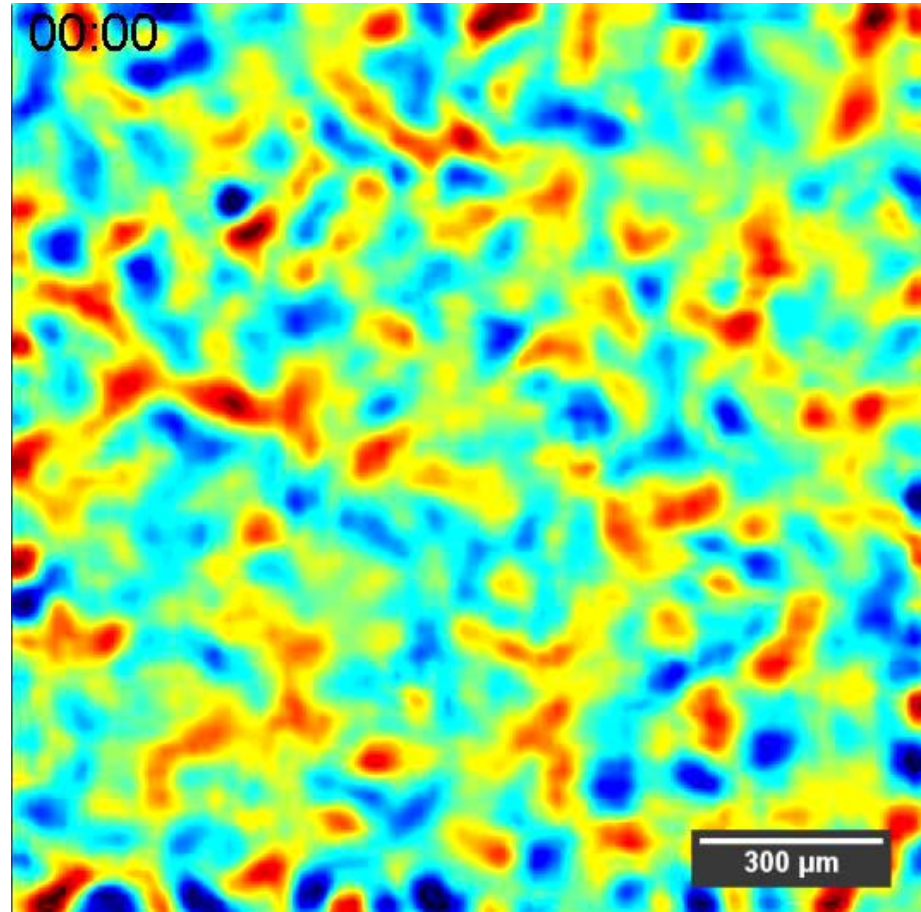
Vorticity field

$$\omega = \partial_x v_y - \partial_y v_x$$



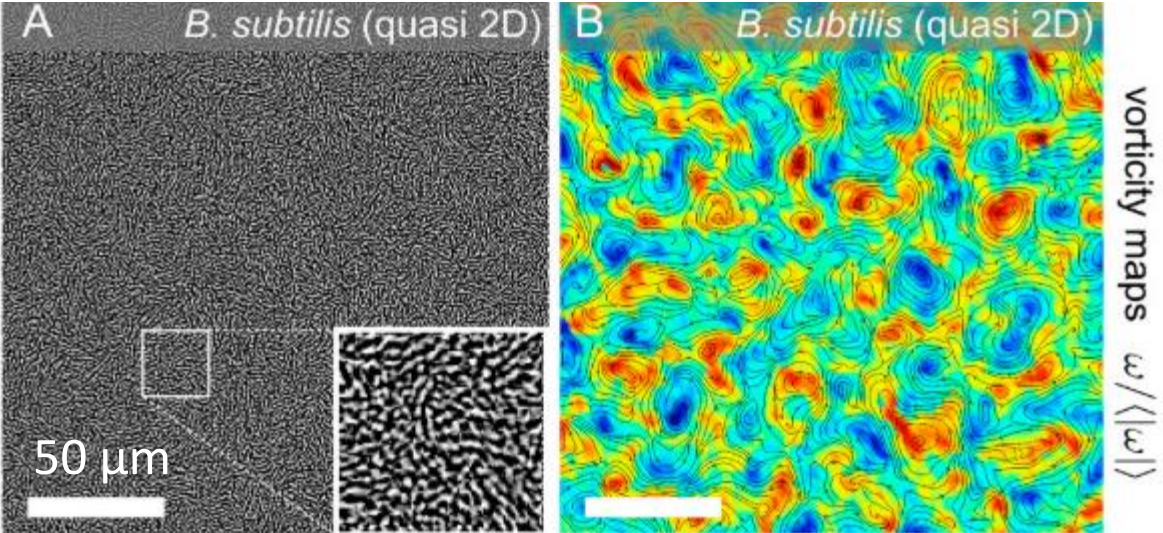
Vorticity field

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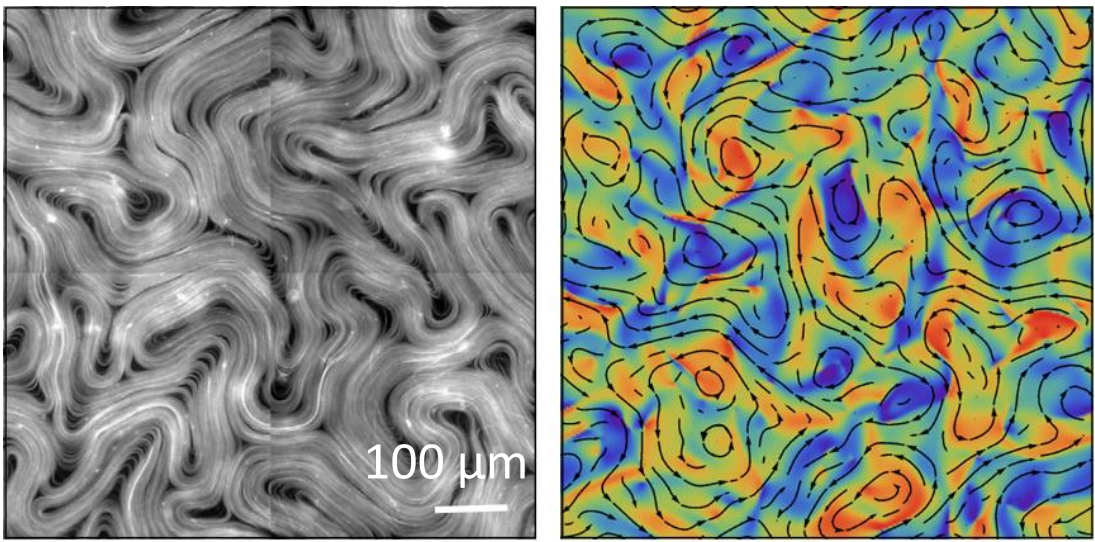
Examples of low-Reynolds number turbulence

Bacterial turbulence



Wensink PNAS 2012

Microtubules + kinesin



Z. Dogic lab

Low Reynolds number turbulence - Theory

L. Giomi PRX 2015:

$$\begin{cases} \rho \frac{Dv_i}{Dt} = \eta \nabla^2 v_i - \partial_i p + \partial_j \sigma_{ij}, \\ \frac{DQ_{ij}}{Dt} = \lambda S u_{ij} + Q_{ik} \omega_{kj} - \omega_{ik} Q_{kj} + \gamma^{-1} H_{ij}. \end{cases}$$

Q_{ij} alignment tensor

S nematic order parameter

p pressure

ν shear viscosity

λ flow alignment

γ rotational viscosity

u_{ij} strain rate tensor

ω_{ij} vorticity tensor

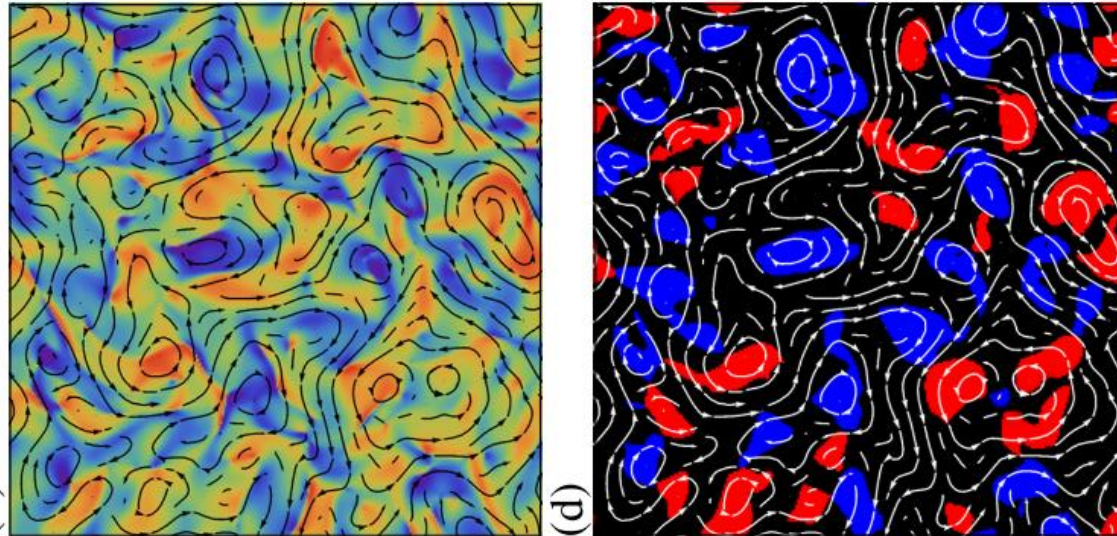
H_{ij} molecular tensor = $-\delta F / \delta Q_{ij}$ where $F_{\text{LdG}} = \frac{1}{2} \int d^2 r [K |\nabla \mathbf{Q}|^2 + C \text{tr} \mathbf{Q}^2 (\text{tr} \mathbf{Q}^2 - 1)]$,

stress tensor $\sigma_{ij} = \sigma_{ij}^e + \sigma_{ij}^a = \underbrace{-\lambda H_{ij} + Q_{ik} H_{kj} - H_{ik} Q_{kj}}_{\text{Elastic stress}} + \underbrace{|\zeta \Delta \mu| Q_{ij}}_{\text{Active stress}}$

Elastic stress

Active stress

Numerical simulation :



vorticity
 $\omega = \partial_x v_y - \partial_y v_x$

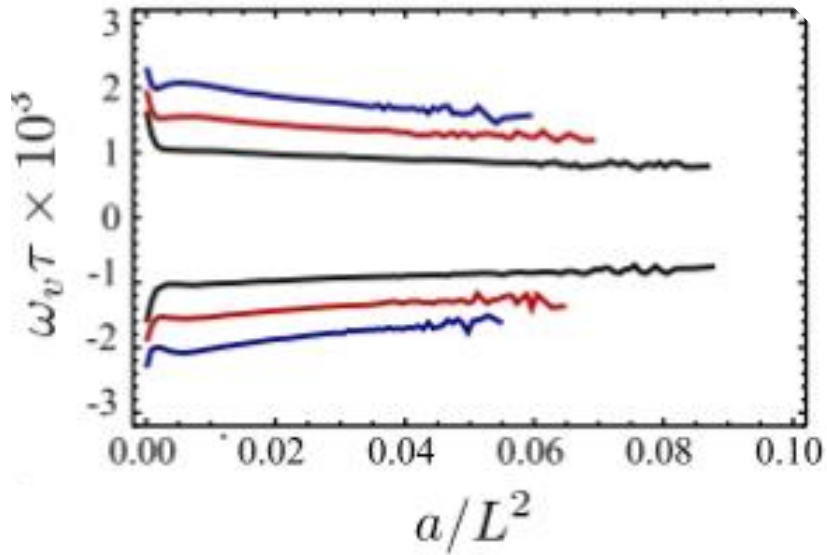
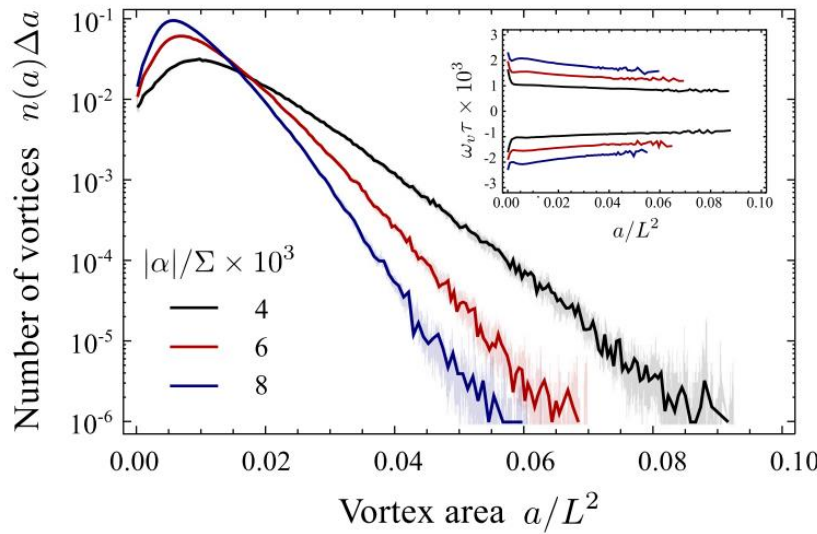
Okubo-Weiss field

Measurements:

- Number of vertices, size, mean vorticity per vortex
- «Kinetic» energy $\mathcal{E} = \langle (v_x^2 + v_y^2)/2 \rangle$
- Enstrophy $\Omega = \langle \omega^2/2 \rangle$

Characteristic active length $l \sim \sqrt{\frac{K}{|\zeta \Delta \mu|}}$ Chaotic if $l \ll L$

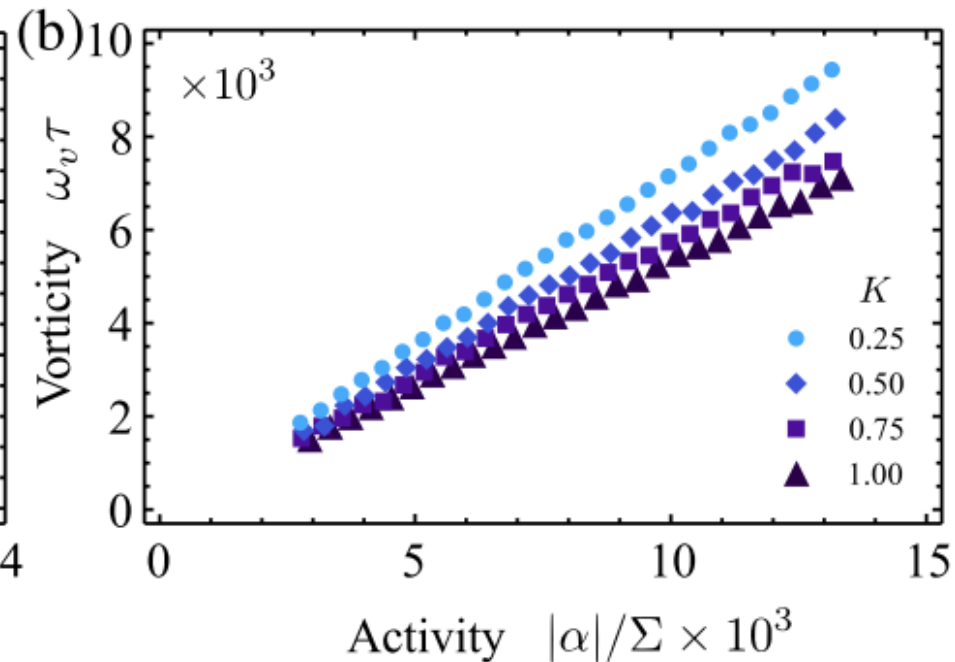
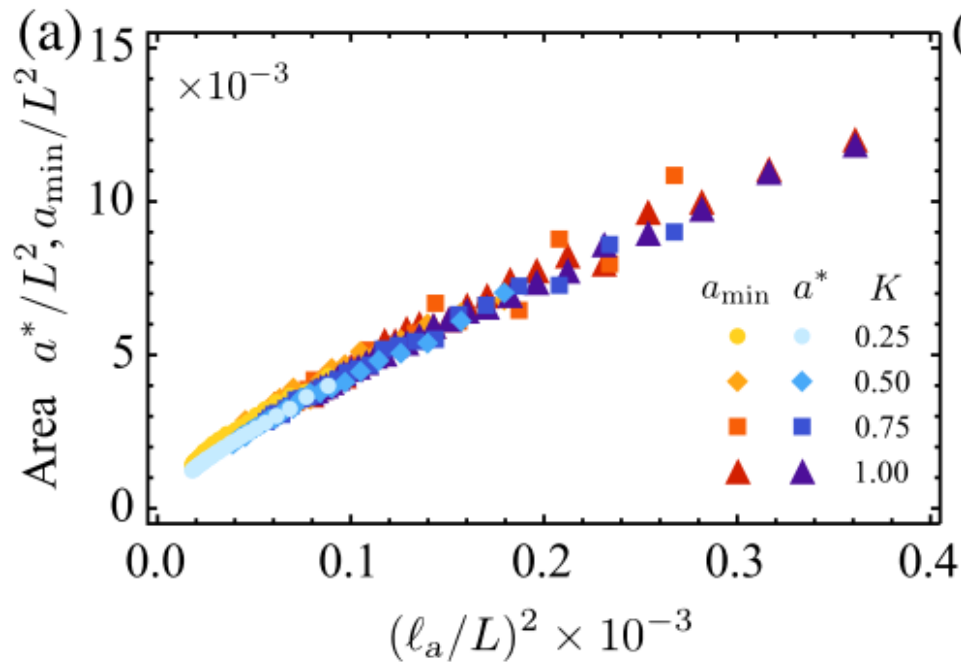
Simulations: Vertices area and vorticity



- Exponential distribution of the vertex areas (multiscale)
- Near-constant mean vorticity

Larger activity means smaller and faster vertices

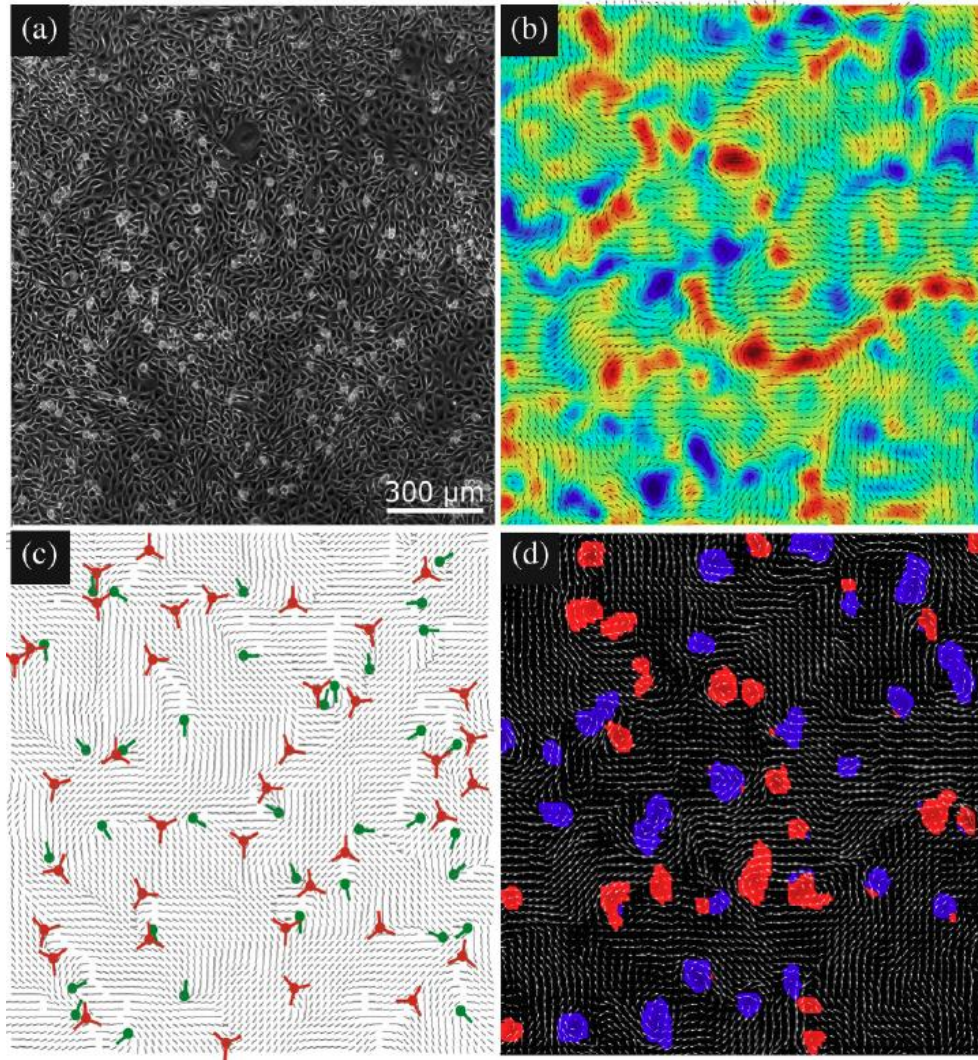
Impact of activity



$$P(a) \propto \exp\left(-\frac{a}{l^2}\right)$$

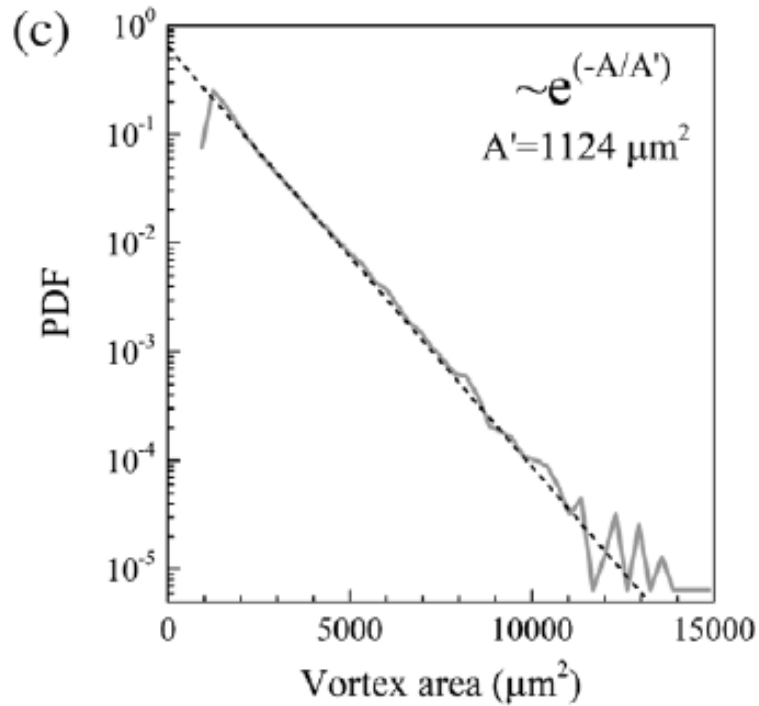
$$\begin{aligned} \omega_v &\propto |\zeta \Delta \mu| \\ \rightarrow \omega_v &= |\zeta \Delta \mu| / \eta \end{aligned}$$

Experiments (HBECs)

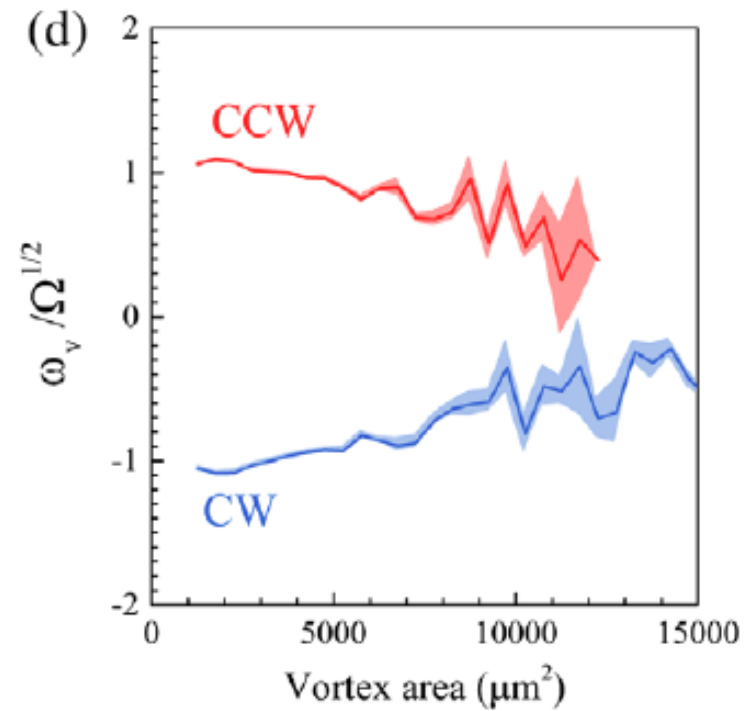


Okubo-Weiss
field

Experiments

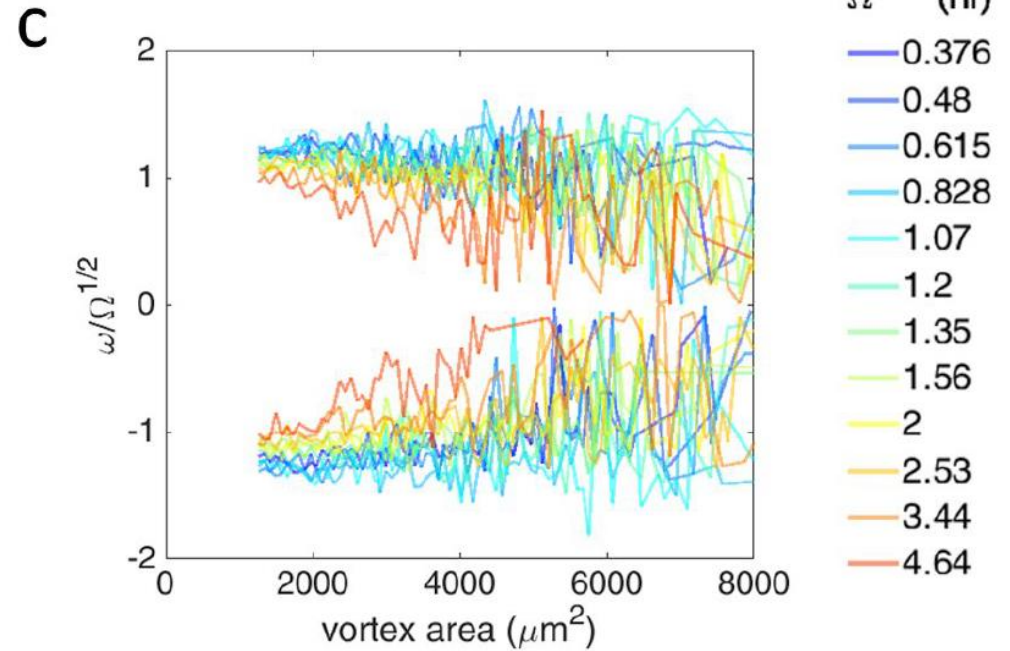
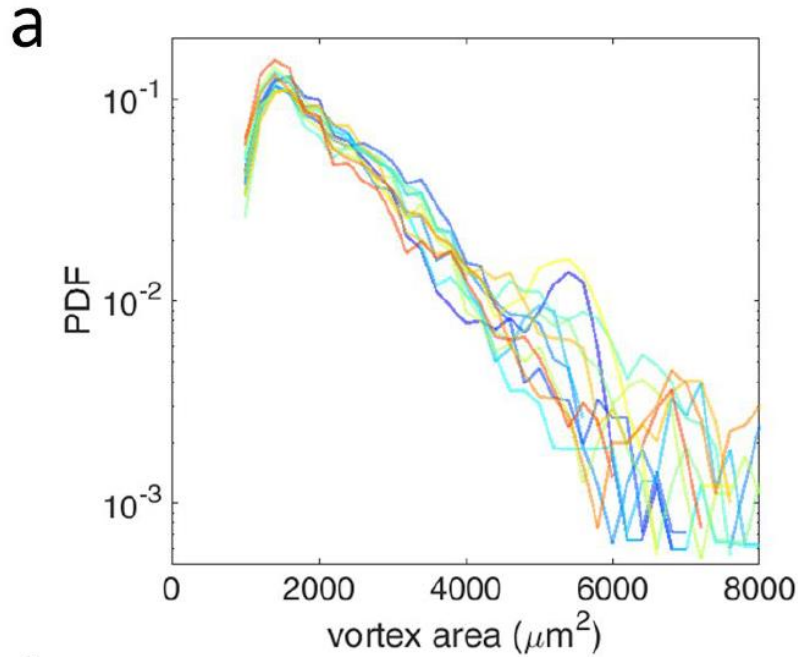


Exponential distribution



Low dependence of mean vorticity
on vortex size

Signature of turbulence in the system



Activity and elastic constants don't vary with time

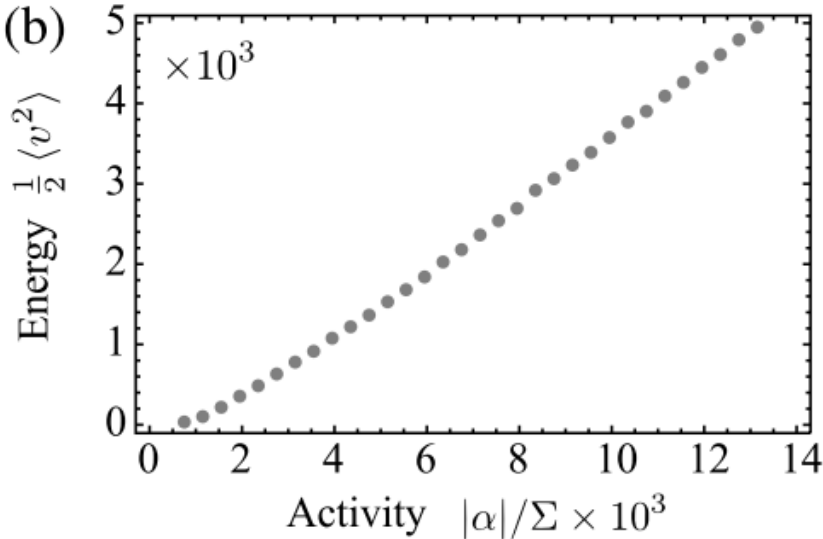
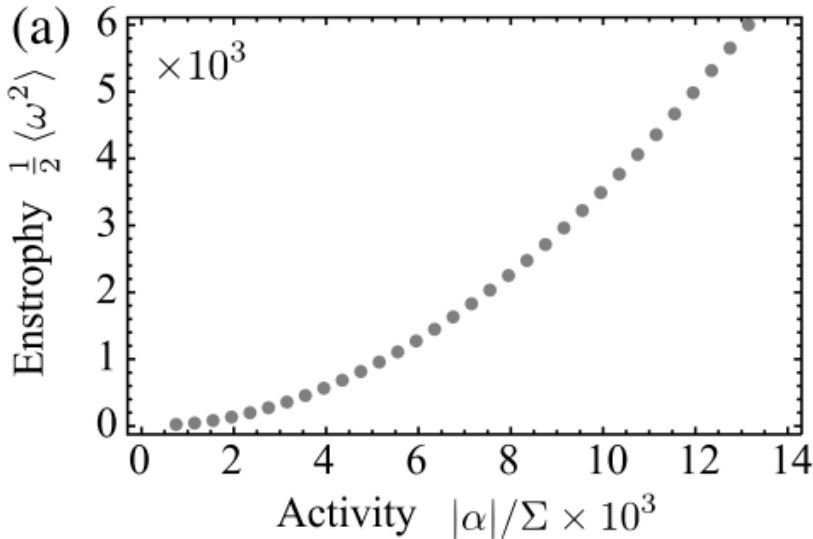
$$\sqrt{A'} \sim l \sim 50\mu\text{m}$$

$$\longrightarrow \omega_v \sim |\zeta\Delta\mu|/\eta$$

Simulations: enstrophy and energy

$$\Omega = \langle \omega^2 / 2 \rangle$$

$$\mathcal{E} = \langle (v_x^2 + v_y^2) / 2 \rangle$$

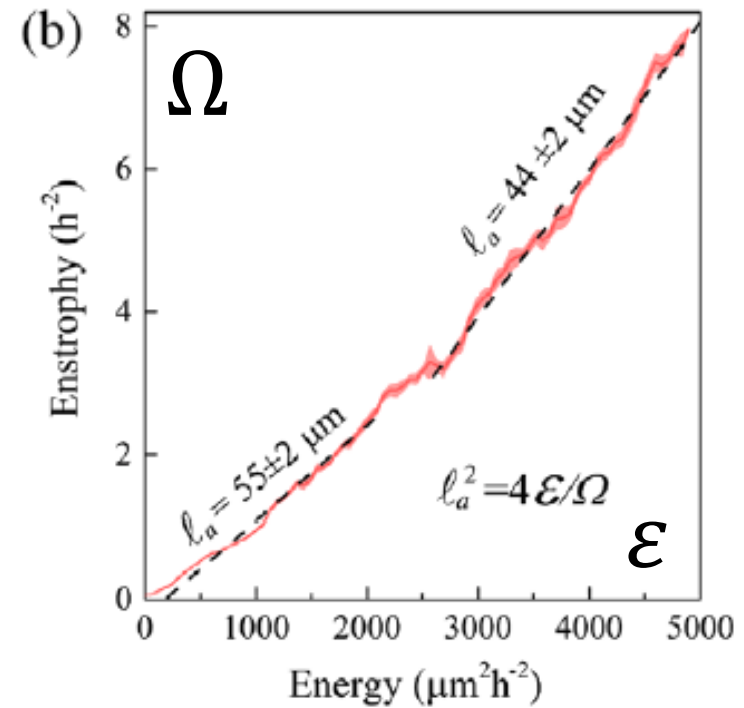
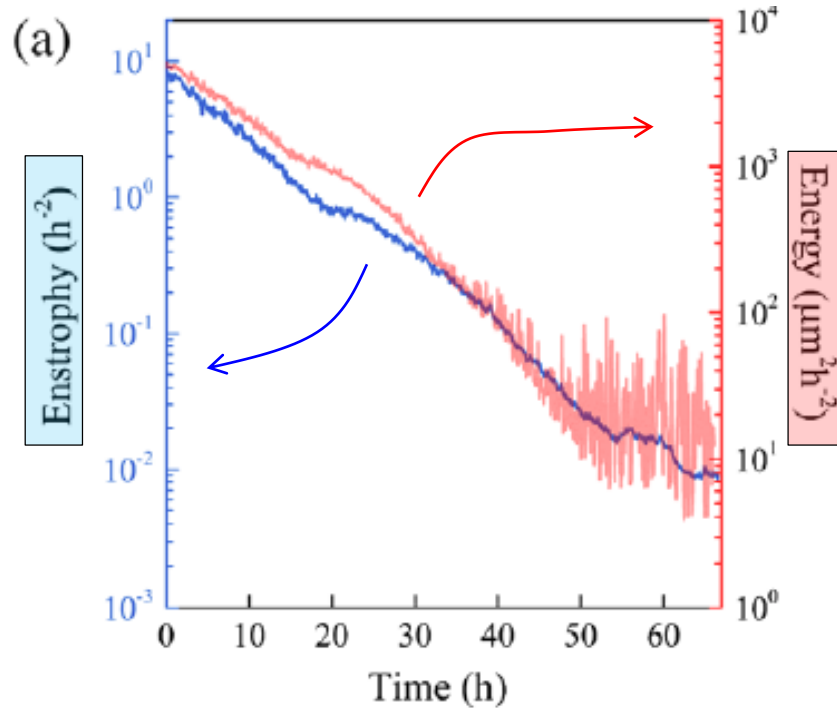


$$\Omega \propto |\zeta \Delta \mu|^2$$

$$\mathcal{E} \propto |\zeta \Delta \mu|$$

so: $\boxed{\mathcal{E} / \Omega \sim l^2}$

Experiments: Energy, Enstrophy

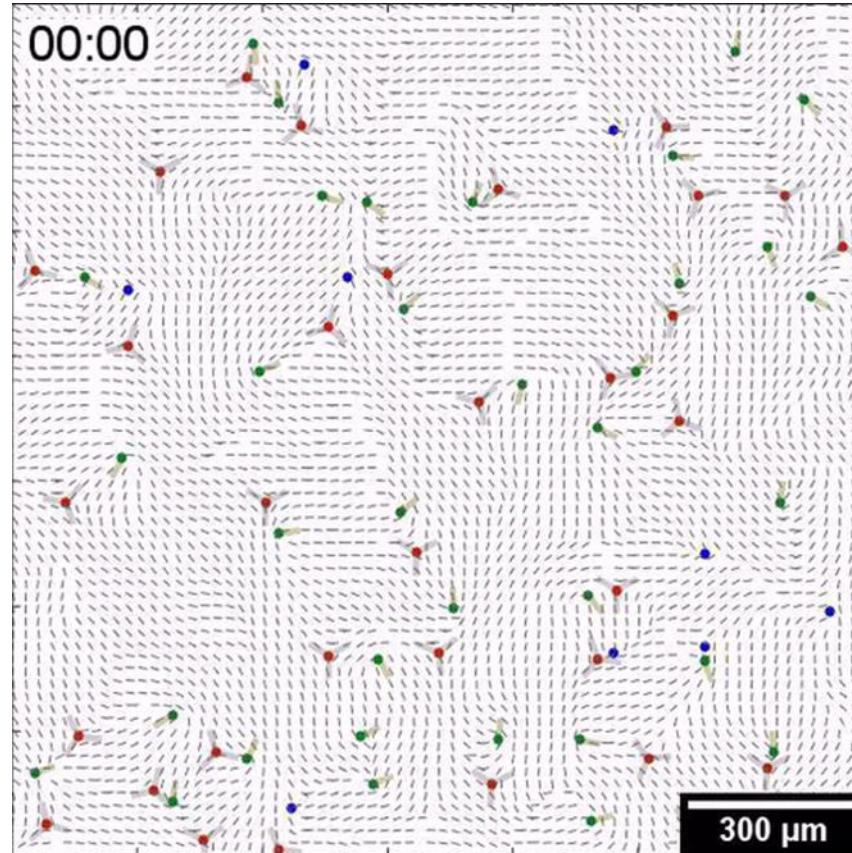


$$\Omega \sim \omega_v^2 \sim (|\zeta \Delta \mu| / \eta)^2$$

$$\epsilon / \Omega \sim l^2$$

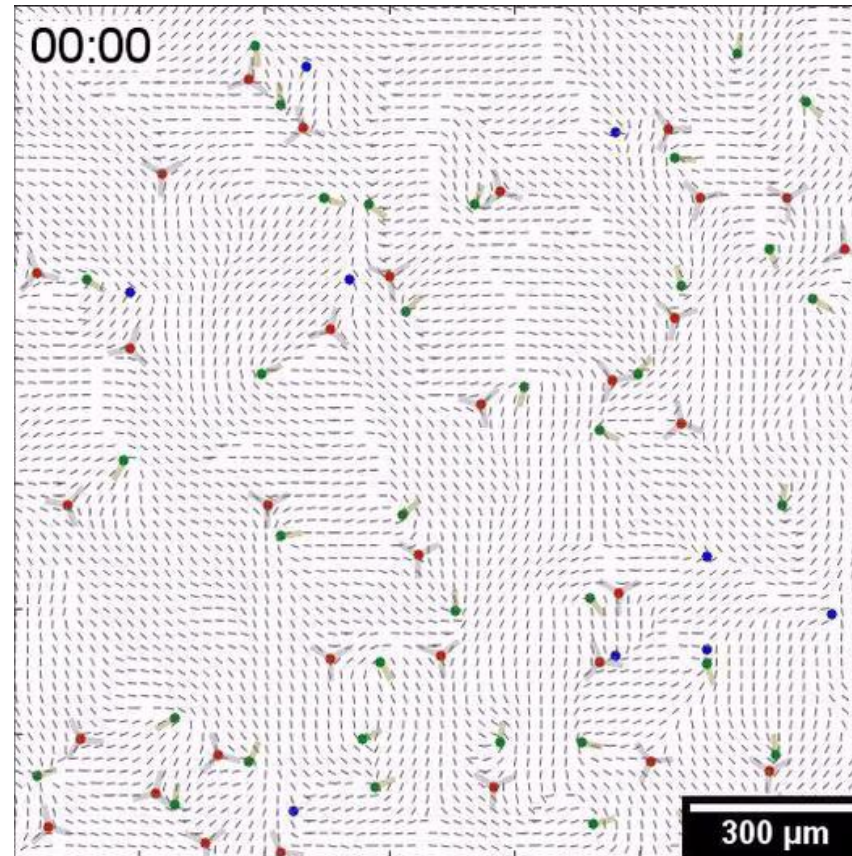
Increase of viscosity (x10) as the system enters jamming

Defects



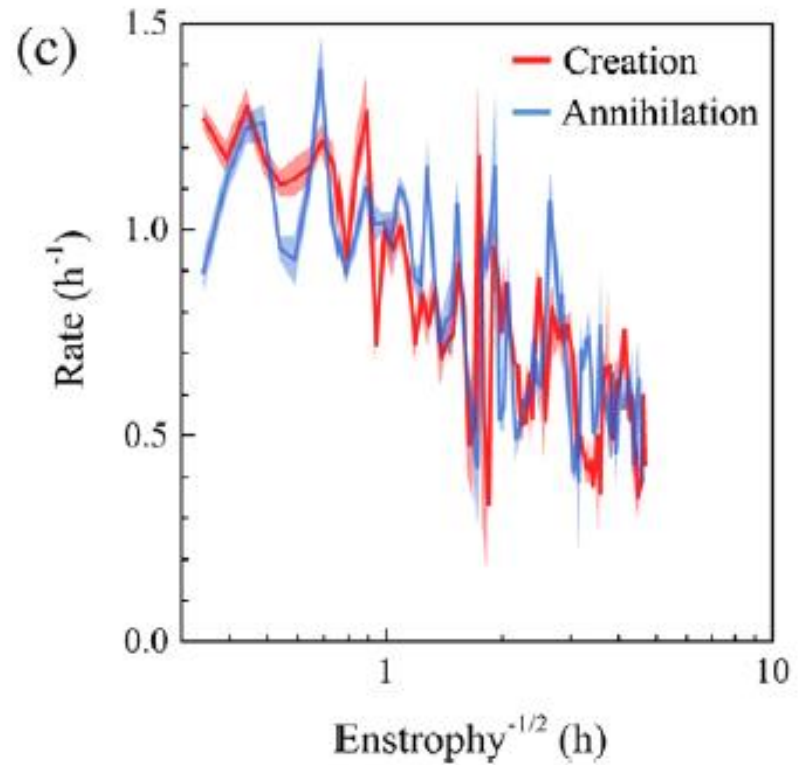
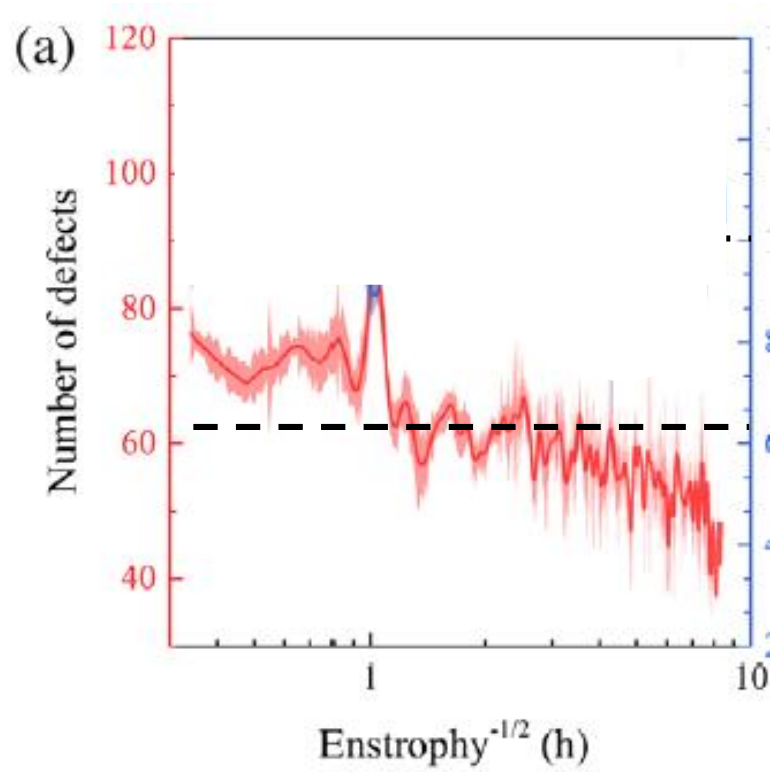
+1/2
-1/2

Defects



+1/2
-1/2

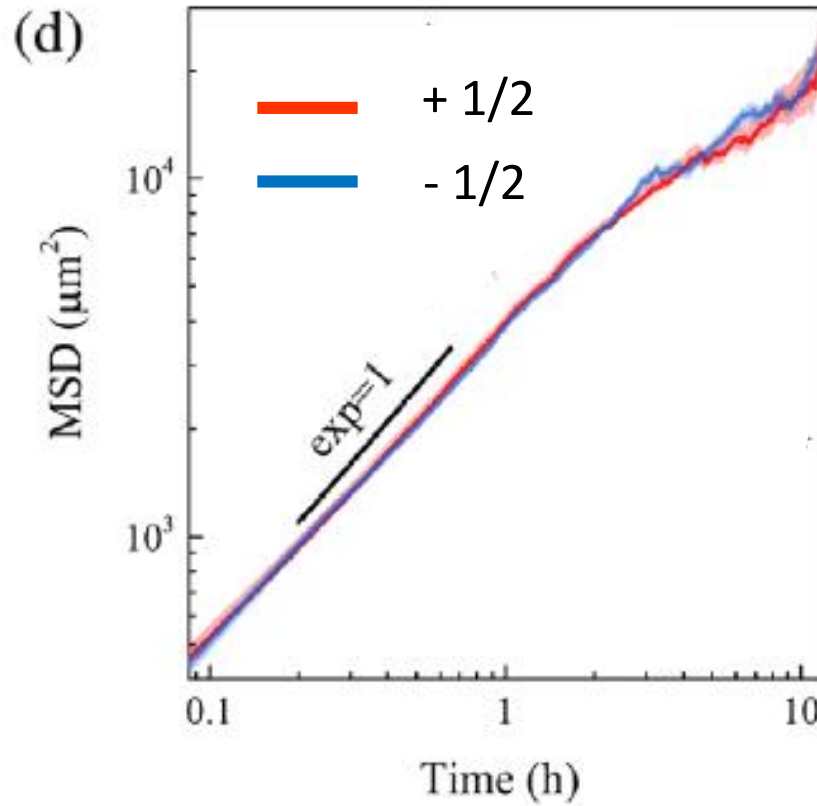
Defects



Dynamical balance creation-annihilation

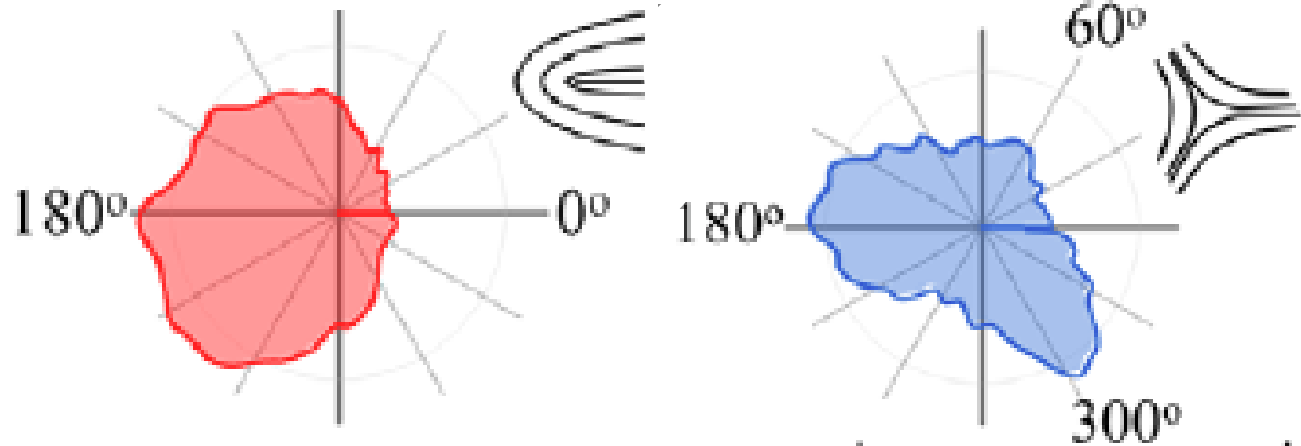
Defects

MSD



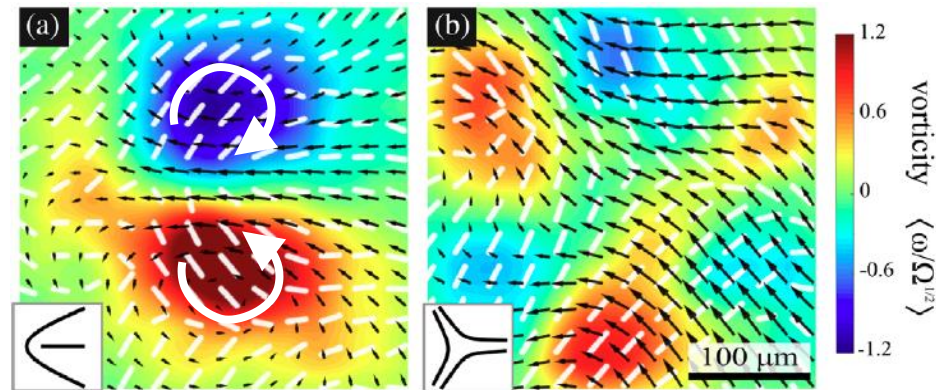
Diffusive

Direction of the flow field with respect to the defect axis

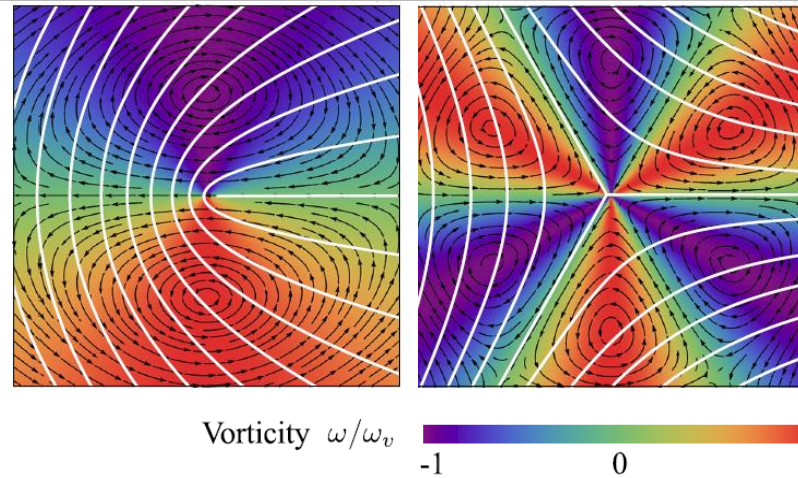


Flows and topological defects

Experiments
 $N > 10\,000$



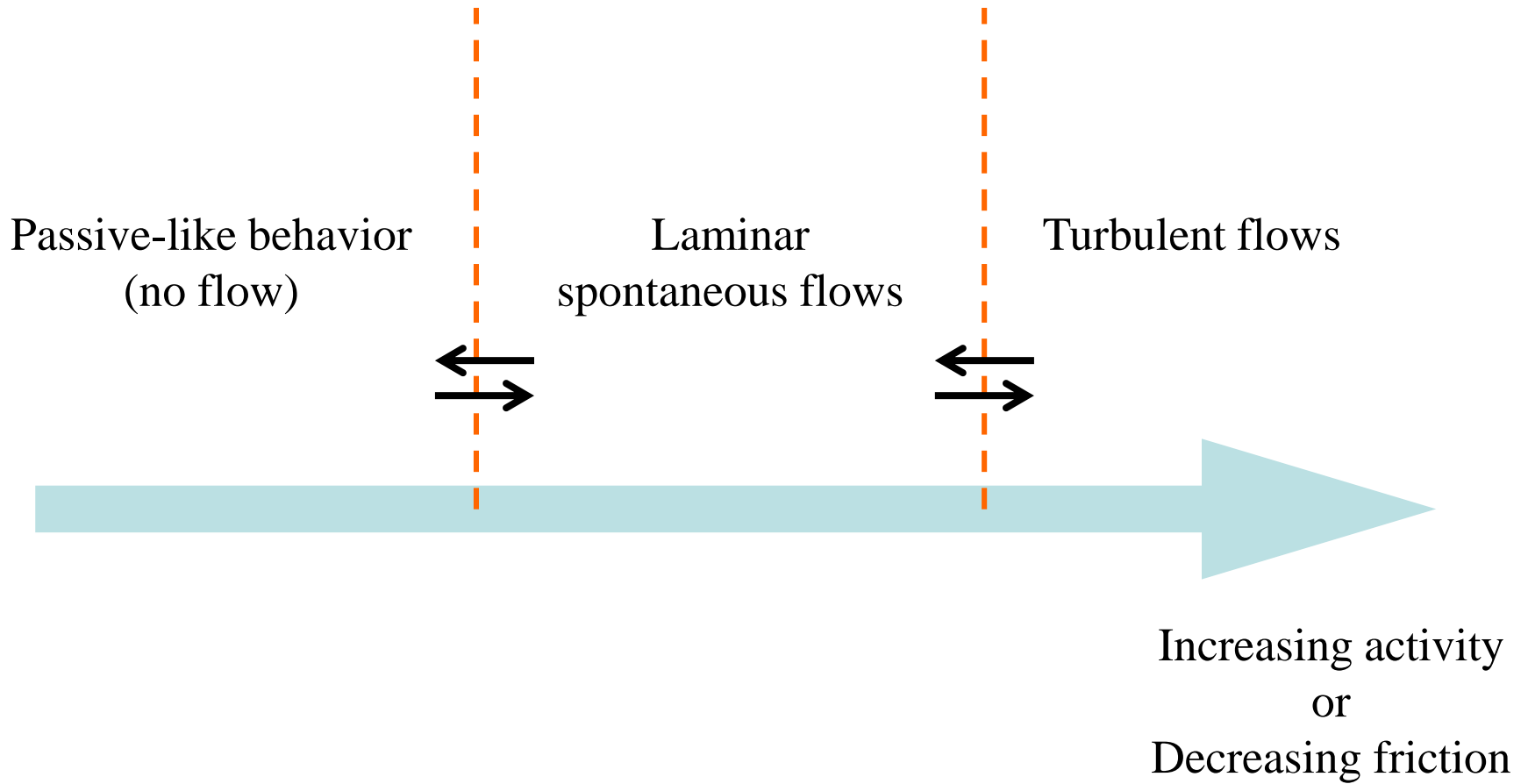
Theory
(Giomi 2015)



- Interplay between the defects and the vertices :
Defects \rightarrow vertices \rightarrow transport of defects and chaotic mixing

Conclusions

- A cellular monolayer can undergo a transition from a low-density disordered state to a highly **ordered nematic state after confluence / “Meso-contact guidance”**
- In the final state, **activity is secondary to nematic elasticity** (jamming). Activity screened out by **friction**.
- Spontaneous **active Fréedericksz transition** {tilted/shear} to {untilted/immobile}. Very general physical principles / parameters' values
- **Turbulent** collective flows, Chaotic dynamics. Coupling of defects with the vertices - Transition to jamming (increase of η). **Physiological function?**





- A. Buguin, I. Bonnet, S. Coscoy
- G. Duclos, T. Aryaksama, T. Sarkar, M. Lacroix
- V. Yashunsky, T. Vourc'h, F. Ascione, B. Smeets
- N. Sepulveda, V. Hakim (ENS, Paris)
- J. Camonis, M. C. Parrini (Curie)
- J.-F. Joanny, J. Prost, C. Blanch-Mercader, C. Erlenkaemper (Curie)
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