# **Organism and Tissue Growth**



### Course 5: Growth control and mechanics

Thomas Lecuit chaire: Dynamiques du vivant

COLLÈGE



### • Control of growth arrest at target size



- ---What is measured? Intrinsic « ruler » of growth: « size-meter » Organ intrinsic feedback of target organ size on cell growth/proliferation
- -Need to consider <u>scaling</u> between factors that promote growth and size itself
  - I. Energy and mass/volume
  - 2. Pattern and mass/volume
  - 3. Mechanics and mass/volume

-Need to consider feedback mechanisms operating across scales (organ/tissue to cell)



### • Conclusions

- Cell growth is controlled intrinsically
- -What are the mechanisms of organ specific tissue size sensing/measurement?
- --- What is measured? Intrinsic « ruler » of growth: « size-meter », scaling.

- Energy delivery/demand unbalance.
- Organ-size dependent negative feedback on cell growth/proliferation (chalone)
- Size-meter »: scaling of chalone concentration and tissue size distance between activator and feedback inhibitor mechanical signal that scales with size patterning cue: morphogen gradient
- Organ-size dependent growth inducer: Morphogen and growth arrest: Spatial model: gradient scaling and gradient slope. Temporal model: gradient scaling and rate of signal increase
- Importance of considering robustness of mechanisms: e.g. integral feedback



# Programmed vs Self-organised regulation of Growth



- hierarchical
- modular
- deterministic rules (ie. genetically encoded)



- no hierarchy
- feedbacks
- statistical rules



- Organs grow until they reach a fixed size characteristic of each organ/species
- Growth is often uniform in the face of non uniform growth factor distribution (morphogens)
- Tissues can compensate for growth heterogeneities and reach a normal final size arguing that cells assess their environment
- -How do cells compare their growth rates?
- -How is growth uniform on average in a tissue?
- -How is tissue size determined?

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• Cell-cell mechanical interactions





### Tissue Growth and Homeostasis





### Tissue Growth and Homeostasis





### Tissue Growth and Homeostasis

-Tissue growth depends on the balance between:

- Rate of cell division
- Rate of cell death and cell extrusion

—Tissue growth leads to homeostasis at a fixed size where cell growth/division and death/extrusion balance each other (unless growth is arrested): eg. gut.

- How is such a balance achieved?
- Feedback interactions
- Does growth impact on extrusion?
- Does extrusion lead to compensatory growth?



#### —What are the consequence of differential tissue growth rates?

- Cell and tissue competition (eg. tumours)
- Tissue buckling (morphogenesis)



### Biomechanical signalling driving tissue growth





### Control of cell division by mechanics: tension

### The control of cell division by tension or diffusion

The anchorage dependence of division in normal cells<sup>11,12,15</sup> and the fact that there is a minimum dimension of surface for this phenomenon<sup>16</sup> can be explained by this hypothesis if it is presumed, as Maroudas<sup>16</sup> first postulated, that the cells have to spread sufficiently to tense themselves, and that tension stimulates the cell cycle.

### Cells in culture are subjected to low frequency stretching (60 $\mu$ m) for 1 hour

Table 1 Effects of repetitive tensing of a cell sheet on mitotic index (Expt 1) and on <sup>3</sup> H-thymidine incorporation (Expt 2)							
Expt 1							
Frequency (Hz)	Experiment mitotic index	Control: probe moving, not tensing cells mitotic index					
0.25	$4.1 \pm 1.1$	$2.5 \pm 0.4$					
0.5	$5.1 \pm 1.5$	$2.3 \pm 0.7$					
1.0	$3.8 \pm 1.7$	$2.2 \pm 0.4$					
Control, no probe inserted Control, probe	$2.1 \pm 0.4$						
inserted, not moving	$1.9 \pm 0.6$						
Expt 2							
Frequency (Hz)	Experiment % labelled cells	Control: probe moving, not tensing cells % labelled cells					
0.25	$21.4 \pm 5.1$	$14.3 \pm 1.5$					
0.50	$22.5 \pm 5.6$	$14.1 \pm 3.2$					
1.00	$15.3 \pm 2.5$	$13.9 \pm 1.6$					
Control, probe not inserted	$14.7 \pm 2.2$						
Control, probe inserted	$14.3 \pm 2.2$						



### Control of cell division by mechanics: geometry

• Density dependence of cell proliferation and contact inhibition are characteristic of cells in culture

- Increased cell-substrate adhesion causes reduction of cell height and increased cell diameter
- Cell proliferation increases as cells spread/adhere more to the substrate
- Is it <u>adhesion per se or cell shape per</u> <u>se?</u>

Nature Vol. 273 1 June 1978

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#### Role of cell shape in growth control

#### Judah Folkman & Anne Moscona

Department of Surgery, Children's Hospital Medical Center and The Harvard Medical School, Boston, Massachusetts 02115



Table 1 Effect of crowding on cell shape and DNA synthesis (WI-38 cells)						
	Poly (HEMA)			Plastic		
Dilution of poly (HEMA)	Cell density (cells per cm <sup>2</sup> )	<sup>3</sup> H Incorporation (c.p.m.)	Cell height (µm)	Cell density (cells per cm <sup>2</sup> )	<sup>3</sup> H Incorporation (c.p.m.)	
1.00 4×10 <sup>-2</sup> 4×10 <sup>-3</sup>	15,000 15,000 15,000	$7 \pm 5$ $50 \pm 5$ $250 \pm 15$	22 15 6	250,000 (confluent) 100,000 (confluent) 30,000 (subconfluent)	7±1 55±9 254 ∤23	



# Control of cell division by mechanics: geometry or adhesion?

#### **Geometric Control of Cell Life and Death**

Christopher S. Chen, Milan Mrksich, Sui Huang, George M. Whitesides, Donald E. Ingber\*



- Cell division increases as the adhesive island surface increases
- Cell death by apoptosis decreases in parallel

—Disentangling cell geometry and substrate adhesion using a controlled micro fabricated substrate

- The projected surface area, but not the surface of adhesion to the ECM impacts on cell proliferation and cell death.
- Does this reflect cell shape per se or cell stresses?





0.20 0.15 0.10 0.05 0.00 BrdU **Proliferation** map 1.00 0.79 0.59 0.38 0.18 -0.03 Stress from FEM

endothelial cells on fibronectin

# —Disentangling the role of cell stress and cell shape

- using micro fabricated substrates on colony island of multiple cells (rather than single cells)
- Cell division patterns depend on the geometry colony islands
- -Mapping stress pattern
- Use of finite element modelling (FEM) to deduce stress patterns in colony islands.
- Cell division patterns correlate with stress patterns that emerge from cell contractility and connectedness between cells and tissue geometry.
- Traction Force microscopy confirms the deduced mechanical stress pattern.

C. Nelson *et al.* C. Chen. (2005) *P.N.A.S.* 102: 11594–11599 www.pnas.orgcgidoi10.1073pnas.0502575102





BrdU incorporation density maps

0.20 0.15 0.10



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----Identification of transcription factors that mediate mechanically induced cell proliferation

- YAP/TAZ are known components of the Hippo/LATS growth pathway that translocates in the nucleus once activated.
- Substrate stiffness induces YAP activation

• Cell substrate adhesion/geometry affects YAP/TAZ activation

• Cell projected area, but not adhesion per se activates YAP//TAZ





• The actomyosin cytoskeleton (but *not* the microtubule network) is required for YAP/TAZ activation.









---Identification of transcription factors that mediate mechanically induced cell proliferation

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Apoptosis (%) Proliferation (%)

- Cell stretching induces activation of cell proliferation and inhibition of apoptosis
- YAP/TAZ are both required for this process
- Constitutively active YAP expression induces cell proliferation and inhibits apoptosis
- This effect is independent of upstream Hippo/LATS pathway activity (not shown)

- YAP/TAZ ----YAP/TAZ \* (C.A.) siYAP/TAZ +5SA-YAP siCo. + C3 siControl Control Proliferation (%) 60 60 40 40 n.s. n.s. 20 20 0 30 Apoptosis (%) 30 20 20 10 0,000,02,02<sup>4</sup>,00 10,00 2 000 02 02 02 30C 10,00,02,02,02,00 2025,024.300 Adhesive island area (µm<sup>2</sup>) Adhesive island area (µm<sup>2</sup>) substrate stiffness 40 kPa **a** 40 kPa 40 kPa 40 kPa 1 kPa 40 kPa siYZ1 Osteogenesis (A.U.) 0 70 0 10 0 0 Adipogenesis (A.U.) b 60 20 40 40 40 40 (kPa) 40 40 (kPa) 40 1 STL r sille sico. έ<sup>ς</sup>Ο<sup>Ο.</sup> STA ico. ුඋි
- YAP/TAZ similarly mediates the cell differentiation of stem cells dependent on substrate stiffness
   On soft substrates a dia substrate an stiff substrates
- On soft substrates adipocytes, on stiff substrates osteocytes.

[see also A. Engler et al, D. Discher. (2006) *Cell* 126, 677–689]



#### -Cell spreading affects cell volume via water efflux

• Cells spread more on stiffer substrates and reduce concomitantly their volume

- Restricting cell area using smaller adhesive islands, increases cell volume
- Dynamic cell spreading is associated with dynamic change in cell volume
- The volume dependence on spread area is the same irrespective of how area is attained (substrate stiffness, adhesion area, dynamics of spreading)
- Volume could be changed by protein concentration changes or water efflux: no [protein] change

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• Volume change caused by water efflux







M. Guo et al. J. Lippincott-Schwartz and D. Weitz. PNAS. (2017) E8618–E8627 www.pnas.org/cgi/doi/10.1073/pnas.1705179114



-Volume changes induced by cell spreading or osmotic stress affect nuclear volume and molecular crowding in the nucleus.

- Hyper-osmotic stress (with high PEG concentration in the medium) causes a reduction in cell volume through water efflux irrespective of substrate stiffness.
- Blocking Cl ion channels (NPPB) blocks volume change as a function of substrate stiffness so volume adjustment involves changes in osmolytes.
- Blocking cell contractility also affects stiffness dependent volume change.
- Consistent with tension dependent regulation of mechanosensitive ion channels, affecting osmolyte concentration and water flux.

reviewed in R. Phillips, T. Ursell, P. Wiggins and P. Sens (2009) Nature 459: 379-385

- In all conditions nuclear volume adjusts to cell volume
- Histine H2::GFP mean square displacement is lower in compressed cells consistent with increased molecular crowding





M. Guo et al. J. Lippincott-Schwartz and D. Weitz. PNAS. (2017) E8618–E8627 www.pnas.org/cgi/doi/10.1073/pnas.1705179114



-Volume changed induced by osmotic stress affects differentiation of stem cells

- Differentiation of stem cells into osteocytes on a stiff substrate
- Hypertonic medium rescues differentiation on soft substrate.
- This is associated with volume reduction of cell and nucleus and increased cell stiffness
- On soft substrates, conversely cells differentiate into adipocytes
- Hypotonic medium rescues adipogenesis if on stiff substrate



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M. Guo et al. J. Lippincott-Schwartz and D. Weitz. PNAS. (2017) E8618–E8627 www.pnas.org/cgi/doi/10.1073/pnas.1705179114

#YAP forms liquid like nuclear condensates in response to molecular crowding induced by hyperosmotic stress

D. Cai et al. and J. Lippincott-Schwartz. bioRxiv. http://dx.doi.org/10.1101/438416. (2019)

now in press in *Nature Cell Biology Nat Cell Biol.* 2019 Dec;21(12):1578-1589. doi: 10.1038/s41556-019-0433-z.



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# Control of cell growth/division & death by mechanics

• Summary





### Mechanics of Tissue Growth and Homeostasis

-Tissue growth depends on the balance between:

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—Tissue growth leads to homeostasis at a fixed size where cell growth/division and death/extrusion balance each other (unless growth is arrested): eg. gut.

- How is such a balance achieved?
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—What are the consequence of differential tissue growth rates?

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# Control of cell growth/division by mechanics

-Changes in cell behaviour during collective tissue growth *in vitro*: contact inhibition

- Growth of isolated colonies to disentangle cell contact from mechanics
- « free growth » regime is associated with exponential growth while cells are in contact
- Colony growth slows down, limited by constrained velocity of edge cells (ie. friction).
- This is associated with a transition to increased cell density and slower motility which are mechanically induced









cell density and size

A [um<sup>2</sup>]

A [µm<sup>2</sup>]



cell motility



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A. Puliafito et al. and B. Shraiman. (2012) PNAS 109:739-744

# Control of cell growth/division by mechanics

-Changes in cell behaviour during collective tissue growth in vitro: contact inhibition

—1D vertex mechanical model of selflimiting growth

- Short time elastic response of cells
- Friction coupling to the substrate (elastic restoring force)
- Active random cell motion (Langevin driving force) averages to zero in colony bulk but not at colony edges
- Cell growth is modelled by increasing the rest length of a cell edge when neighbouring cells stretch it.
- Cell division halves rest length
- Cell division rate depends on cell size



-Simulation results

- 2 phases: exponential and linear growth
- Cell division in the bulk and gradual arrests due to increased cell density
- Cell division persists at colony edges as cells are stretched.



#### -Compression induced cell extrusion



- Cells in the midline of a growing tissue delaminate.
- Tissue overgrowth (induced by activation of the Pi3K pathway) increases cell delamination
- Reducing tissue growth reduces cell delamination/ extrusion
- Tissue crowding induces cell delamination

-40 min -15 min -10 min 0 min

4

junction number

2

1.0

Normalized area 9.0 0.4 0.2 0.2

ſ

6

Cell delamination is associated with apical area reduction and junction remodelling (loss). E-cad-GFP





Insulin

Growth/Cell division

PTEN

-Compression induced cell extrusion a



• Mechanical vertex model of junction remodelling underlying compression induced cell delamination.











-Compression induced cell extrusion



- A tissue monolayer under compression adjusts its cell density in two phases:
- Phase I: Density first increases rapidly due to compression.
- Phase 2: Cell density then slowly decreases until reaching initial steady state.
- Adjustment of cell density is associated with cell extrusion by apoptosis and independent of apoptosis.
- Cell extrusion requires ROCK, the sphingosine kinase SK and sphingosine phosphate (SIP2).



Eisenhoffer, G.T., et al., and Rosenblatt, J. (2012). Nature 484, 546-549.



-Compression induced cell extrusion



- In whole organisms: gut villi and fins in zebrafish, cell crowding causes cell extrusion.
- This requires the mechanonosensitive receptors Piezo.

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Eisenhoffer, G.T., et al., and Rosenblatt, J. (2012). Nature 484, 546-549.



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-Stretch induced cell divisions



- Stretching of cell cultures (MDCK cells) induces cell division.
- Wound healing in a cell monolayer causes stretching of cells after closure and induces a burst of cell division after closure
- The mechanosensitive receptor Piezo I is required for cell division in MDCK culture cells and in zebrafish fins
- Piezo activation gives rise to calcium signalling, ERK activation and Cyclin B induction





#### -Stretch induced cell divisions

Regulation of cell cycle progression by cell-cell and cell-matrix forces

#### Cdt1::RFP: G1-S Geminin::GFP: S-G2-M

- MDCK cell culture expand at a constant cell density
- When cell division is blocked (G1/S arrest), cell area increases



• Cell division is sustained while the colony expands and thereby maintains a constant density.









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M. Uroz et al. and X. Trepat. (2018) Nature Cell Biology 20:646–654

#### 



- Cells sense compression and stretch via the same mechanosensitive receptor Piezo and adjust their density depending on their mechanical state.
- Cell density evolves until the population reaches a steady state X where cell death/extrusion and cell division balance each other



X: steady-state density where cell death/extrusion and cell division balance each other



# Balancing cell growth and death by mechanics: tumours

#### -In 3D, homeostatic pressure: a tissue property where cell growth/division balances apoptosis

• Homeostatic pressure



Tissue grows against wall resisted by a spring. Growth proceeds until internal pressure  $p = p_h$  where growth and apoptosis balance each other.

The spring position is stable, since further growth increases the pressure above this value  $p_h$  and induces apoptosis, whereas recession of the piston decreases the pressure and causes cell division.

• Tumour growth



Tissue H



Tissue T

• Cell competition (see 26 Nov 2019)





A. Matamoro-Vidal and R. Levayer *Current Biology* (2019) 29, R762–R774,

Markus Basan , Thomas Risler , Jean-François Joanny , Xavier Sastre-Garau & Jacques Prost (2009) *HFSP Journal*, 3:4, 265-272, DOI: 10.2976/1.3086732



### Balancing cell growth vs death by mechanics: competition

#### —Differential growth by cell competition requires cell intercalation

- <u>Differential cortical tension</u> between winner-winner cell interfaces and loser-loser cell interfaces causes cell intercalation at the winner/loser interface, cell isolation and cell death.
- Mechanics (differential cortical tension) drives cell competition and differential growth.





Levayer, R., Hauert, B., and Moreno, E. (2015). Nature 524, 476-480.



RFP (WT loser in *tub-dmyc*) E-cad::GFP

### Balancing cell growth vs death by mechanics: competition

—Differential growth by cell competition requires tissue compaction induced cell delamination





### Balancing cell growth vs death by mechanics: competition

Mechanical cues affects cell competition in 3 ways:

• Modulating the geometry of cell signalling leading to loser cell elimination,





• Stimulating the compensatory growth of winner cells around extruded/dying loser cells via cell stretching.





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Reviewed in: A. Matamoro-Vidal and R. Levayer *Current Biology* (2019) 29, R762–R774.

*Current Biology* (2019) *29*, R762–R774, https://doi.org/10.1016/j.cub.2019.06.030

- Organs grow until they reach a fixed size characteristic of each organ/species
- Growth is often uniform in the face of non uniform growth factor distribution (morphogens)
- Tissues can compensate for growth heterogeneities and reach a normal final size arguing that cells assess their environment
- -How do cells compare their growth rates?
- -How is growth uniform on average in a tissue?
- -How is tissue size determined?





- -How do cells compare their growth rates?
- -How is growth uniform on average in a tissue?
- Mechanical Feedback



B. Shraiman PNAS, (2005) 102:3318-3323

- Regulation as an integral feedback:

$$\frac{dp(r,t)}{dt} = \frac{\mu K}{1+\mu} \left[ \gamma(r,t) - \langle \gamma(t) \rangle \right] - \frac{1}{\tau} p(r,t),$$

- Compression or stretching of a group of cells is dependent on integral over the history of the clone of its growth rate with respect to average surrounding growth.
- Cell rearrangements limits the « memory » of this feedback so adaptation of growth rate is not perfect.



- Initial rate of growth is faster than that of a background clone.
- At later stages, the rate of growth is reduced to match that of the background, resulting in a ratio of mutant to background clone sizes (the overgrowth ratio) independent of time



— Non-autonomous effect: The mechanical feedback as a possible mechanisms underlying cell competition

- Dynamics of a fast-growing clone in a background of slow growing cells
- Fast growing cells increase their pressure *p* to *p*<sub>0</sub> and reduce their growth rate until they reach the growth rate of the surrounding
- Neighbouring slow growing cells may die as a consequence of high pressure
- Non-autonomous cell death around fast growing clone





— Integration of morphogen and mechanical feedback



 $M(r) = me^{-r/\lambda}$ 

- 1) growth rate goes down with increasing stress ie. negative mechanical feedback.
- 2) growth rate has a maximum
- 3) the maximum depends on morphogen concentration above a threshold value *M*<sub>0</sub>
- 4) the morphogen M does not scale



Increased pressure reduces perimeter and increases height Stress p is proportional to  $\xi_{\alpha} - 1$   $\xi_{\alpha} - 1 > 0$  :compression  $\xi_{\alpha} - 1 < 0$  :stretch





— Integration of morphogen and mechanical feedback

- Growth homogeneity: The mechanical feedback suppresses the effect of the high concentration of the morphogen at the center of the tissue and ensures uniform growth
- Growth arrest: when M falls below threshold at edges, growth arrests there and pressure increases in the bulk thereby causing growth arrest.



Division follows iso-growth lines in the M, p phase plane









— Integration of morphogen and mechanical feedback

#### Assumptions:

- Two morphogens that scale with tissue size
- Cell stretching induces growth above a threshold
- No growth if both stretch and morphogen are too low

#### **Results:**

- Initially growth higher in center due to higher growth factors GF
- As a result cells are stretched at periphery, which promotes growth.
- Growth lowers stretching but cells remain stretched and compress the center, thereby reducing growth.
- The wider the periphery the greater the compression.
- Growth arrests when compression overcomes the effect of growth factors







T. Aegerter-Wilmsen et al. *Mechanisms of Development* 124 (2007) 318–326

- Pattern of tissue deformations in growing imaginal discs

- Gradient of cell area consistent with compression at the centre.
- Evidence of tissue anisotropy in a growing epithelium at tissue periphery





L. Le Goff et al . and T. Lecuit Development 140, 4051-4059 (2013) doi:10.1242/dev.090878

Y. Mao et al. and N. Tapon. The EMBO Journal (2013) 32, 2790–2803. doi:10.1038/emboj.2013.197

**Texture tensor** 
$$M_i = \frac{1}{N_i} \sum_{j=1}^{N_i} \begin{pmatrix} \Delta x_{ij}^2 & \Delta x_{ij} \Delta y_{ij} \\ \Delta x_{ij} \Delta y_{ij} & \Delta y_{ij}^2 \end{pmatrix}$$

Graner, F., et al. (2008). Eur. Phys. J. E 25, 349-369.



- Stress patterns in growing imaginal discs

 Cells are stretched tangentially in the growing tissue

 laser ablation experiments and





• Planar polarised distribution of actomyosin contractile network



L. Le Goff et al . and T. Lecuit Development 140, 4051-4059 (2013) doi:10.1242/dev.090878



Y. Mao et al. and N. Tapon. The EMBO Journal (2013) 32, 2790–2803. doi:10.1038/emboj.2013.197

- Stress patterns in growing imaginal discs В 0.25 • The induction of a fast-growing anisotropy of cells 0.20 clone causes non-autonomous tissue deformation (anisotropy) 0.15 C'Myoll-GFP ea e.g. activation of the Hippo pathway (Yorkie++ or Expanded 10 mutant) or of targets of Hippo 20 10 30 40 vivoll-GFF distance from the center (um) pathway (mRNA bantam)



L. Le Goff et al . and T. Lecuit *Development* 140, 4051-4059 (2013) doi:10.1242/dev.090878
Y. Mao et al. and N. Tapon. *The EMBO Journal* (2013) 32, 2790–2803. doi:10.1038/emboj.2013.197
Y. Pan et al and B. Shraiman and K. Irvine. (2016) PNAS www.pnas.org/cgi/doi/10.1073/pnas.1615012113



- Cytoskeletal actomyosin tension modulates tissue size

- Activation of Rho I/Rok affects wing size
- Modulates Yorkie (YAP) pathway activity (Expanded expression)
- and Adjuba recruitment at cell junctions















— Cytoskeletal actomyosin tension increases tissue size via activation of the Hippo growth pathway

Tension at E-cadherin complexes recruits Adjuba and the kinase Warts at cell junctions and inhibits its activity via « sequestration » at cell cortex.
This releaves Yorkie repression by Warts and causes Yorkie nuclear

translocation, hence activation of the pathway.





#### — Tension is reduced in fast growing clones

#### Mechanical Feedback

I. overgrowth compresses cells;
2. compression decreases cytoskeletal tension (conversely, stretching increases actomyosin network and tension)
2. decreases in subschedatal tension trianges

3. decrease in cytoskeletal tension triggers reduction in the cortical recruitment of Jub and Wts, and in Yki activity hence on growth.





— This emerges from the mechanical feedback model where mechanical cell response adapts to external mechanical stress pattern.





Cell shape reflects balance between edge tension and internal pressure



- Adjuba and Warts localization at cell junction are decreased in overgrowing clone of cells

and rescued when Myosin2 is constitutively active (and tension increased in these overgrowing cells)









Y. Pan et al and B. Shraiman and K. Irvine. (2016) PNAS www.pnas.org/cgi/doi/10.1073/pnas.1615012113

-Yorkie induced overgrowth downregulates Yorkie activity (evidence of negative feedback)



- Blocking the feedback affects causes growth heterogeneity
  - Forcing Bantam expression independent of mechanical feedback
  - Removing Warts so it cannot be subjected to feedback regulation
  - Forcing Myosin2 constitutively











Y. Pan et al and B. Shraiman and K. Irvine. (2016) PNAS www.pnas.org/cgi/doi/10.1073/pnas.1615012113

- Integration of chemical and mechanical signalling

Feedback mechanisms responsible for uniform growth:

-increased growth leads to increased compression, which decreases growth,

-it leads to a compression gradient that induces growth in regions with lower growth rates



• vertex model with cell shape governed by







• Growth rate  $G_{\alpha} = (cg_1 + cg_2 * [Yki]_{\alpha} + cg_3 * [Vg]_{\alpha} + cg_4 * [N]_{\alpha} - cg_5 * [Brk]_{\alpha}) * cg_6$ 

Tinri Aegerter-Wilmsen et al. and C. Aegerter and K. Basler. Development 139, 3221-3231 (2012) doi:10.1242/dev.082800



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Growth

- Integration of chemical and mechanical signalling

Non-trivial model predictions:



Tinri Aegerter-Wilmsen et al. and C. Aegerter and K. Basler. *Development* 139, 3221-3231 (2012) doi:10.1242/dev.082800



#### Mechanical feedback:

- corrects the consequence of non uniform growth rates, and smoothens out heterogeneities: growth homogeneity.
- may be involved in compensatory mechanisms (eg. cell competition) in the context of tissue growth and homeostasis.
- part of growth arrest mechanisms at the scale of organs.

Yet, tissue growth can be highly heterogeneous.

- normal development: growth zone (limb bud, beak morphogenesis), tissue folding.
- disease: tumour evolution.



### Differential growth and tissue folding

• Cortex convolutions



#### physical model of brain like instability



swelling of outer layer (growth)

Gl: gyration index (total area/exposed area)



### Differential growth and tissue folding

• Gut vilification:

- Epithelium and mesenchyme form annealed surfaces ensheathed by sequential layers of smooth muscles
- Formation of **Ridges** in epithelium and mesenchyme parallel formation of circumferential smooth muscles
- Formation of **Zigzags** parallels formation of longitudinal muscles
- Formation of Villi formation of longitudinal muscles





### Growth induced mechanical instabilities: Gut vilification

### • Gut vilification

- —Theoretical studies:
- E. Hannezo J. Prost and J-F. Joanny PRL 107, 078104 (2011)
- M. Ben Amar and F. Jia. PNAS 110: 10525–10530 (2013)
- -Computation model
- A. Shyer et al, C. Tabin and L. Mahadevan. Science 342: 212-218 (2013)







# Summary

#### Mechanical regulation of cell division and cell death/extrusion

- Cells respond differently to compressive and tensile stresses:
- Cell growth, cell division, cell extrusion or simple cell shape changes, cell intercalation.
- These processes underlie viscoelastic properties of tissues: energy dissipation in homeostatic conditions
- Cell mechanics underlies tissue homeostatic pressure.
- Not well understood what factors determine these properties
- Sensing: mechanosensitive receptors (Piezo), cortical tension and YAP pathway
- Cellular response: mechanics of junction remodelling (see Courses 2018)

#### Tissue growth and mechanical feedback:

- corrects the consequence of non uniform growth rates, and smoothens out heterogeneities: growth homogeneity.
- may be involved in compensatory mechanisms (eg. cell competition) in the context of tissue growth and homeostasis.
- required for growth arrest at the scale of organs.

